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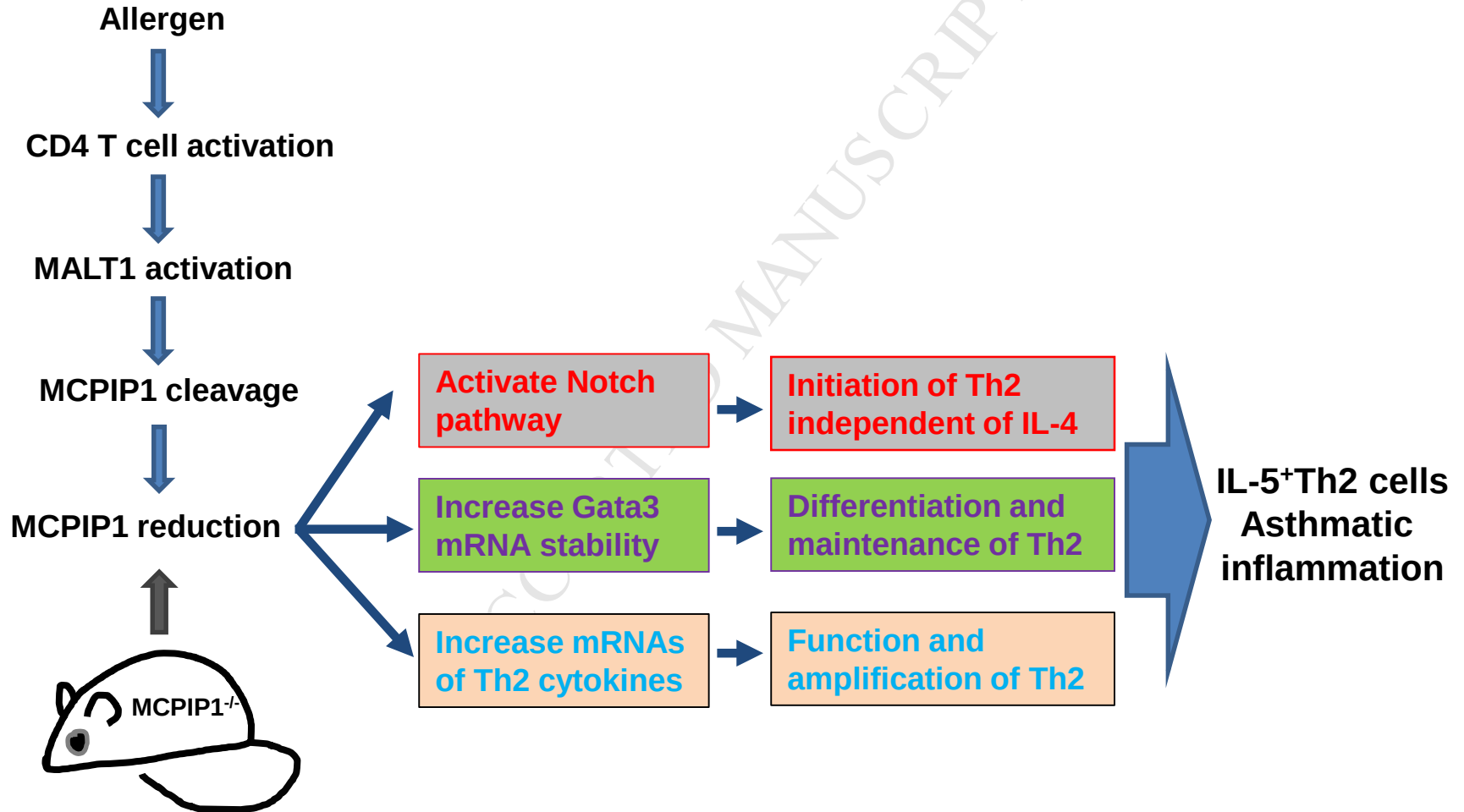
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Running title: MCPIP1 inhibits IL-5⁺Th2 cells via Notch

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Abstract

Background: Asthmatic and allergic inflammation is mediated by Th2 cytokines (IL-4, IL-5 and IL-13). Though we have learned much about how Th2 cells are differentiated, the Th2 checkpoint mechanisms remain elusive.

Objectives: In this study, we investigate how monocyte chemotactic protein induced protein-1 (MCPIP1, encoded by *zc3h12a* gene) regulates IL-5-producing Th2 cell differentiation and Th2-mediated inflammation.

Methods: The functions of *zc3h12a*^{-/-} CD4 T cells were evaluated by checking the expression of Th2 cytokines and transcription factors *in vivo* and *in vitro*. Allergic airway inflammation of *zc3h12a*^{-/-} mice was examined with murine asthma models. In addition, antigen-specific CD4 T cells deficient in MCPIP1 were transferred to WT recipient mice, challenged with OVA or HDM, and assessed for Th2 inflammation.

Results: *Zc3h12a*^{-/-} mice spontaneously develop severe lung inflammation, with an increase mainly in IL-5- and IL-13-producing but not IL-4-producing Th2 cells in the lung. Mechanistically, the differentiation of IL-5-producing *zc3h12a*^{-/-} Th2 cells is mediated through Notch signaling and Gata3 independent of IL-4. Gata3 mRNA is stabilized in *zc3h12a*^{-/-} Th2 cells. MCPIP1 promotes Gata3 mRNA decay via the RNase domain. Furthermore, deletion of MCPIP1 in OVA- or HDM-specific T cells leads to significantly increased Th2-mediated airway inflammation in OVA or HDM murine models of asthma.

Conclusions: Our study reveals that MCPIP1 regulates the development and functions of IL-5-producing Th2 cells through Notch/Gata3 pathway. MCPIP1 represents a new promising target for the treatments of asthma and other Th2-mediated diseases.

Key Messages

- MCPIP1 specifically inhibits IL-5/IL-13-producing Th2 cell differentiation and function.
- MCPIP1 suppresses the early differentiation of IL-5-producing Th2 cells through Notch.
- MCPIP1 inhibits IL-5-producing Th2 development by targeting Gata3 and Th2 cytokine transcripts for degradation.

Capsule Summary

This study reveals that MCPIP1 regulates the development and functions of IL-5-producing Th2 cells through Notch/Gata3 pathway. MCPIP1 represents a new promising target for the treatments of asthma and other Th2-mediated diseases.

Key words

RNA-binding protein, MCPIP1, mRNA decay, Th2 cells, IL-4, IL-5, IL-13, Gata3, Notch, asthma.

Abbreviations

BALF (bronchoalveolar lavage fluid), EGFP (enhanced green fluorescent protein), KO (knockout), WT (wild type), MCPIP-1 (monocytes chemoattractant induced protein 1), MCP-1 (monocytes chemoattractant protein 1), FSH (forward scatter), HDM (house dust mites), OVA (ovalbumin), P/I (PMA and ionomycin), Ab (antibody), DLN (draining lymph nodes), MFI (median fluorescence intensity), H&E (hematoxylin and eosin), PAS (periodic acid-Schiff).

INTRODUCTION

The hallmark of asthma is bronchial hyperreactivity caused by Th2 cell dysregulation¹. Th2 cytokines initiate and maintain key pathophysiological features of asthmatic inflammation, with each cytokine exerting different functions in the induction of inflammation^{2, 3}. IL-4 plays a critical role in allergic sensitization and IgE production^{4, 5}; IL-5 is crucial for eosinophil survival^{6, 7}; and IL-13 plays a central role in the development of airway hyperresponsiveness and tissue remodeling^{8, 9}. Expression of Th2 cytokines is controlled by transcription factors through multiple signaling pathways. Th2 cells induce type 2 immunity characterized by high antibody titers and eosinophilia¹⁰⁻¹². Th2 cells can also be induced by parasitic helminths¹³ and are shown to associate with atopic diseases, allergy and asthma^{11, 14}. Though the network of transcription factors required for activation and differentiation of Th2 cells are well understood¹⁵⁻²⁰, how Th2 development and Gata3 expression are negatively regulated, especially through posttranscriptional mechanisms, is largely unknown. Control of gene expression at the posttranscriptional level is a faster event than transcriptional control because of a direct targeting at the protein coding mRNA in the cytoplasm^{21, 22}.

Monocyte chemotactic protein induced protein-1 (MCPIP1) encoded by *zc3h12a* gene was initially discovered as the most highly induced mRNA by monocyte chemotactic protein-1 (MCP-1) in human peripheral blood monocytes²³. MCPIP1 protein is highly expressed in lymph node, thymus and spleen^{24, 25}, and rapidly induced in macrophages upon stimulation with inflammatory stimuli, such as TNF, IL-1 β and LPS²⁵⁻²⁷. MCPIP1 inhibits the expression of proinflammatory cytokines (IL-1, IL-6, and IL-12) by binding to their 3'UTRs for mRNA degradation. MCPIP1 is also named as Regnase-1 based on its RNase activity²⁵. MCPIP1 reduction is required for T activation²⁸. Loss of MCPIP1 in CD4 T cells leads to severe splenomegaly and lymphadenopathy, suggesting that MCPIP1 is essential for inhibiting the development of lethal autoimmune responses^{24, 25}. MCPIP1 functions through regulating mRNA decay and affecting NF- κ B signaling^{25, 29-31}, leading to reduced expression of proinflammatory mediators^{24, 32}. Therefore, MCPIP1 is believed to be a key negative regulator involved in the control of inflammation and maintenance of immune homeostasis. Mice deficient of MCPIP1 develop complex phenotypes, including autoimmune disorders,

anemia and severe inflammatory responses^{24, 25}. In addition, it was recently reported that MCPIP1 can degrade viral RNA and thus acts as a host defense against virus infection³³⁻³⁵. MCPIP1 is also involved in controlling cytokines-induced endothelial inflammation³⁶ and inducing endothelial dysfunction³⁷. Though MCPIP1 inhibits CD4 T cell activation and cytokine production^{28, 30}, it is largely unknown whether MCPIP1 plays a role in Th2 development and Th2 inflammation.

In this study, we report that *zc3h12a*^{-/-} mice spontaneously develop severe airway inflammation, with increased airway Th2 cells capable of producing large amounts of IL-5. *Zc3h12a*^{-/-} CD4⁺ T cells are mostly effector T cells and produce significant amounts of IL-5 and IL-13 but not IL-4. The early differentiation of IL-5-producing *zc3h12a*^{-/-} Th2 cells depends on Notch signal. IL-5-producing *zc3h12a*^{-/-} Th2 cells express high levels of Gata3, and deletion of Gata3 by shRNA/lentivirus results in IL-5 reduction. MCPIP1 suppresses Gata3 expression by promoting its mRNA decay via the RNase domain. Moreover, deletion of MCPIP1 in OVA- or HDM-specific T cells significantly increases Th2-mediated airway inflammation in murine models of asthma. Transferring *zc3h12a*^{-/-} CD4⁺ T cells into congenic CD45.1 mice induces severe Th2 inflammation and accumulation of Th2 cells in the airway of the recipient mice. Our study reveals that MCPIP1 regulates the development and functions of IL-5-producing Th2 cells through Notch/Gata3 pathway. MCPIP1 represents a new promising target for the treatments of asthma and other Th2-mediated diseases.

METHODS

Mice

Zc3h12a^{+/-} mice were provided by Dr. Pappachan E. Kolattukudy (University of Central Florida, Orlando, FL). Homozygous *zc3h12a*^{-/-} mice were bred from *zc3h12a*^{+/-} breeding pairs and genotyped with PCR by detecting exon 4,5,6, with a band of 300 bp indicating WT and 500 bp for knockout. The primer sequences were listed in Table E1. *Zc3h12a*^{+/-} mice and wild type littermates were housed in pathogen-free conditions and used at 5-8 weeks old. The CD45.1 congenic WT mice were used at age 6-12 weeks old. All mice were on C57BL/6 background and housed in animal facility at Saint Louis University. All mouse experiments were approved by Institution Animal Care and Use Committee of the Saint Louis University (animal protocol#: 2443).

In vitro T cell differentiation

Naïve CD4⁺ T cells were purified by EasySep™ Mouse Naïve CD4⁺ T Cell Negative Isolation Kit (Catalog#: 19765, Stemcell Technologies). The purity of naïve CD4 T cells was up to 95%. Naïve CD4 T cells were treated with plate-coated anti-CD3 (2 µg/ml) and anti-CD28 (2 µg/ml) Abs under the following conditions for three days. Th1 conditions: anti-IL-4 (10 µg/ml), IL-12 (10 U/ml); Th2 conditions: anti-IFN-γ (10 µg/ml), IL-4 (20 ng/ml); Th17 conditions: IL-6 (50 ng/ml), TGF-β (1 ng/ml), IL-23 (20 ng/ml), anti-IL-4 (10 µg/ml), and anti-IFN-γ (10 µg/ml). Then, cells were rested in fresh medium with IL-2 (1 ng/ml) for additional three days. On day 6, cells were stimulated with PMA (50 ng/ml) and Ionomycin (0.5 µg/ml) in the presence of Monensin (GolgiStop; 0.7 µl/ml) for 4 h for analysis of cytokines and surface markers by flow cytometry.

mRNA stability

CD4⁺ T cells were treated with plate-coated anti-CD3/CD28 Abs for 4 hours and then stimulated with Actinomycin D (5 µg/ml) and 5,6-dichlorobenzimidazole riboside (DRB, 10 µg/ml) for different times. CEM-Tet-on cells expressing WT or mutant MCP1P1 were stimulated with Dox (100 ng/ml) for 24 hours, then treated by PMA and Ionomycin for 3 h,

and by Act D and DRB for different times. Total RNAs were collected and cDNA were used to measure the remaining mRNAs for calculating half-life.

Murine models of allergic asthma

OVA (Sigma-Aldrich) was adsorbed onto Imject Alum adjuvant (Pierce) to a final concentration of 500 µg/ml. Mice were immunized i.p. of 100 µg of OVA-alum or 10 µg HDM extract (*Dermatophagoides pteronyssinus*; Greer Labs) on day 0, and boosted with OVA on day 7 and 14, or with HDM on day 14. One week later, mice were challenged by intranasal injection (i.n.) of 25 µg OVA or HDM extract in PBS per nostril daily for five days. On day 26, mice were euthanized, and lungs were fixed, sectioned and stained with H&E and PAS. Cells from draining lymph nodes were stimulated with OVA (0, 10, 50, 250 µg/ml) or with HDM extract (30 µg/ml) for 72 hours, and then supernatants were collected for measuring cytokines by ELISA. Splenocytes were cultured with 200 µg/ml OVA or 30 µg of HDM extract in the presence of 10 ng/ml recombinant mouse IL-2 (eBioscience). After 72 hours, CD4⁺ T cells were isolated and resuspended in PBS. 4×10⁶ CD4⁺ T cells were injected i.v. into recipient mice, followed by giving 50 µg OVA or 50 µg of HDM extract i.n. daily for five times starting on days 3. BAL fluid was collected one day after the last challenge for assessment of inflammatory cell infiltration. Lung sections were stained with H&E and PAS, and examined for pathological changes (see additional Methods information in the Online Repository).

Statistical analysis

Data were analyzed with Prism software 5.0 (GraphPad). For standard data sets, an unpaired two-tailed Student's *t*-test was used.

RESULTS

MCPIP1 deficient mice display severe Th2 inflammation in the airway

To test whether MCPIP1 deficient mice have airway inflammation, we stained lung sections with H&E, found a significant accumulation of inflammatory cells and airway remodeling in the lungs of *zc3h12a*^{-/-} mice compared to WT mice (Fig 1, A). There were more mucin-positive cells (goblet-cell metaplasia) in lung epithelium of *zc3h12a*^{-/-} mice than WT mice (Fig 1, B). Total number of cells in BAL were also significantly increased in *zc3h12a*^{-/-} mice compared to WT mice (Fig E1), indicating an asthmatic-like airway inflammation in *zc3h12a*^{-/-} mice. Since Th2 cytokines are critical for asthmatic inflammation, we measured the levels of Th2 cytokines in BAL and blood. IL-5 and IL-13 in serum (Fig 1, C) and BAL (Fig 1, D) were much higher in *zc3h12a*^{-/-} mice than in WT mice, while IL-4 was undetectable in both (data not shown). When collecting BALF, we noticed a reduction in “elastic recoil” of the lungs of MCPIP1 knockout mice. Namely, we could hardly retrieve any PBS after 0.7 ml of PBS was injected into lungs of MCPIP1 knockout mice at the first time. For the second flushing, we could only retrieve 0.1-0.3 ml out of 0.7 ml of PBS. For the final third flushing, we could retrieve 0.4-0.5 ml out of 0.7 ml of PBS. Altogether, we could only retrieve about 30% of the total PBS used for BAL flushing, and most of it was from the last flushing which contained the least levels of cytokines among three BALF. We reason that the excessive Th2 inflammation in the lungs of MCPIP1 knockout mice causes a reduction in lung elastic recoil and an increase in airway permeability, resulting in low levels of cytokines in BALF of the MCPIP1 knockout mice. Nevertheless, even with the low levels of cytokines in BALF, the levels of IL-5 and IL-13 in BALF were still significantly increased in MCPIP1 knockout mice compared to WT mice. The expression of Muc5ac and Gob5 mRNA were also increased in the lungs of *zc3h12a*^{-/-} mice (Fig 1, E). The increased IL-5 and IL-13 in *zc3h12a*^{-/-} mice compelled us to check Th2 cells. Much more IL-5- and IL-13-producing CD4⁺ T cells were presented in the lungs of *zc3h12a*^{-/-} mice than in WT mice; whereas IL-4-producing cells were minimally increased and the cells producing IFN-γ and IL-17A were comparable (Fig 1, F). These data indicate that MCPIP1 deficiency leads to an increase in IL-5/IL-13-producing Th2 cells in the lung, which could cause asthmatic-like airway inflammation.

MCPIP1 deficient CD4 T cells produce significant amounts of Th2 cytokines

In line with the role of MCPIP1 in preventing CD4 T cell activation²⁸, CD4⁺ T cells deficient in MCPIP1 displayed a significant increase in effector T cells (CD62L^{lo}CD44^{hi}) compare with WT mice (Fig E2, A). CD4 and CD8 T cell development were similar between WT and *zc3h12a*^{-/-} mice (Fig E2, B). To test whether *zc3h12a*^{-/-} CD4⁺ T cells produce more Th2 cytokines *in vivo*, we stimulated splenocytes with PMA/Ionomycin and then measured cytokine production. The percentages of IL-5- and IFN- γ -producing effector CD4⁺ T cells were increased in *zc3h12a*^{-/-} CD4⁺ T cells compared to WT CD4⁺ T cells (Fig 2, A). Similar to lung CD4 T cells, splenic *zc3h12a*^{-/-} CD4⁺ T cells also produced more IL-5 and IL-13 than WT CD4⁺ T cells when stimulated with PMA/Ionomycin (Fig 2, B&C). Interestingly, when splenic CD4 T cells were activated with plate-bound anti-CD3/CD28 antibodies, the production of IL-4, IFN- γ and IL-17 became comparable (Fig 2, D). Then, we tested the responses of CD4⁺ T cells to different strengths of TCR signal. Maximal IL-5 and IL-13 production by *zc3h12a*^{-/-} CD4⁺ T cells were induced even by low TCR signaling, while IL-4 production was caught up by WT CD4⁺ T cells and IFN- γ production remained comparable (Fig 2, E and Fig E2, C). These data indicate that once becoming effector cells, WT and *zc3h12a*^{-/-} CD4⁺ T cells have similar capacity to produce cytokines except IL-5 and IL-13 that are still overproduced in *zc3h12a*^{-/-} CD4⁺ T cells. Indeed, IFN- γ -producing cells were comparable between WT and *zc3h12a*^{-/-} effector T cells, while IL-5-producing cells remained increased in *zc3h12a*^{-/-} cells (Fig E2, D). Taken together, these data indicate that MCPIP1 specifically regulates IL-5/IL-13-producing Th2 cells.

MCPIP1 regulates Th2 cytokine production in a cell-intrinsic manner

Zc3h12a^{-/-} CD4⁺ T cells produce significant amounts of Th2 cytokines which might have an autocrine effect on Th2 cytokine production. Therefore, we cultured the WT and *zc3h12a*^{-/-} CD4⁺ T cells at a ratio of 1:1 in the same well to let them first be exposed to the same levels of cytokines. This setting will exclude the possible effects of different levels of cytokines on Th2 cytokine production in comparisons between WT and KO cells. IL-5- and IL-13-producing cells were significantly higher in *zc3h12a*^{-/-} CD4⁺ T cells (CD45.2⁺) than in

WT CD4⁺ T cells (CD45.2⁻), whereas IL-4-, IFN- γ - and IL-17-producing cells were comparable (Fig 3, A&B). Next we measured the mRNAs of Th2 cytokines and transcription factors important for Th2 cell differentiation. Th2-related transcription factor Gata3, IRF4, and Gfi mRNA were higher in *zc3h12a*^{-/-} CD4⁺ T cells than in WT CD4⁺ T cells (Fig 3, C). The master transcription factors important for Th17 cells (Rorc) and for Th1 (Tbx21) and Treg (Foxp3) cells were reduced and slightly increased in *zc3h12a*^{-/-} cells, respectively (Fig 3, C). The mRNA expression of Th2 cytokines (IL-4, IL-5 and IL-13) was increased in *zc3h12a*^{-/-} CD4⁺ T cells, so as the expression of IFN- γ and IL-17A mRNAs, while TGF- β transcript remained unchanged in *zc3h12a*^{-/-} CD4⁺ T cells, compared to WT CD4⁺ T cells in response to anti-CD3/CD28 stimulation (Fig 3, D). In addition, mRNAs encoding costimulatory molecules (*ox40l* and *icosl*) and chemokine receptors (*ccr3* and *cxcr4*) associated with Th2 cells were also highly expressed in *zc3h12a*^{-/-} CD4⁺ T cells (Fig. E3). Taken together, these data indicate an intrinsic effect for MCPIP1 on Th2 cytokine production, especially IL-5 and IL-13.

MCPIP1 regulates IL-5-producing Th2 cell differentiation under unfavorable condition

To determine the effects of MCPIP1 on Th2 development, we differentiated naïve CD4⁺ T cells from *zc3h12a*^{-/-} mice and wild-type littermates toward Th1 and Th2 cells, and then assessed lineage-specific cytokines. Compared with WT cells, the percentages of IL-4-producing CD4⁺ T cells were increased in *zc3h12a*^{-/-} CD4⁺ T cells under non-polarizing condition, while this difference was abolished under Th2-polarizing condition (Fig 4, A-C). However, naïve *zc3h12a*^{-/-} CD4⁺ T cells expressed much higher levels of IL-5 and IL-13 under both non-polarizing and Th2 polarizing conditions (Fig 4, B&C), indicating that naïve *zc3h12a*^{-/-} CD4⁺ T cells were programmed to produce IL-5 and IL-13 once activated even under unfavorable conditions. Indeed, higher levels of IL-5, IL-13 and Gata3 mRNA were detected in *zc3h12a*^{-/-} cells than in WT T cells under unfavorable Th1- and Th17-polarizing conditions (Fig 4, D).

To define the role of IL-4 for IL-5 production in *zc3h12a*^{-/-} CD4⁺ T cells, we respectively added recombinant IL-4 and IL-4 neutralizing antibody during expansion of *zc3h12a*^{-/-} CD4⁺

T cells, and found that neither adding IL-4 nor neutralizing IL-4 had any effects on IL-5 expression (Fig 4, E&F). Neutralizing IL-4 inhibited the expression of IL-4 and Gata3 in non-IL-5-producing *zc3h12a*^{-/-} T cells (Fig 4, E&F), suggesting that IL-4 is not required for the differentiation of IL-5-producing *zc3h12a*^{-/-} CD4 T cells. In addition, to confirm the role of MCPIP1 in Th2 cytokine expression, we treated WT and *zc3h12a*^{-/-} CD4⁺ T cells with MI-2, a specific inhibitor of MALT1 known to increase MCPIP1 levels by blocking its degradation²⁸, and measured cytokine responses. Neither increasing MCPIP1 by MI-2 treatment nor deleting MCPIP1 affected IL-4 expression in CD4⁺ T cells (Fig E4, A), further suggests the independence of MCPIP1 in IL-4 expression. Increasing MCPIP1 by MI-2 treatment inhibited IL-5 and IL-13 expression in WT CD4⁺ T cells but not in *zc3h12a*^{-/-} CD4⁺ T cells (Fig E4, B&C). These data further indicate that MCPIP1 selectively regulates the development of IL-5/IL-13-producing Th2 cells but not the IL-4-producing Th2 cells.

Notch signaling mediates the initiation of IL-5-producing Th2 cells by MCPIP1 deficiency

Notch-mediated Th2 cell differentiation and Th2 immunity³⁸⁻⁴⁰ are also IL-4 independent. Notch signaling is initiated by binding of Notch ligands to its receptors. There are five mammalian Notch ligands, three of which belong to the Delta-like family (DLL1, DLL3 and DLL4) and two belong to the Jagged family (Jag1 and Jag2)⁴¹. We found that the expression of Jag1, Jag2 and DLL4 were increased in naïve *zc3h12a*^{-/-} CD4⁺ T cells compared with WT T cells (Fig 5, A). To determine whether Notch signaling is involved, we pretreated naïve *zc3h12a*^{-/-} CD4⁺ T cells with GSI, a γ -secretase inhibitor that blocks cleavage of the Notch intracellular domain (NICD) from the membrane, stimulated the cells with anti-CD3/CD28 antibodies, and then measured Th2 cytokines. IL-5 and IL-4 production were significantly inhibited while IFN- γ production enhanced after blocking Notch signaling by GSI (Fig 5, B&C), demonstrating the involvement of Notch pathway in MCPIP1-mediated Th2 inhibition. Next, we tested whether Notch signaling is required for cytokine production in effector T cells. We purified total CD4⁺ T cells (most are effector cells) from *zc3h12a*^{-/-} mice, added GSI, and then measured Th2 cytokines and Gata3. Blockade of Notch pathway did not affect IL-5 and IL-4 production as well as Gata3 expression in *zc3h12a*^{-/-} effector CD4 T cells

(Fig 5, D and Fig E5, A). Furthermore, we co-cultured total WT and *zc3h12a*^{-/-} CD4⁺ T cells in the same wells with or without GSI. Blockade of Notch signaling by GSI inhibited the production of IL-4 and IL-13 in WT CD4⁺ T cells but not in *zc3h12a*^{-/-} CD4⁺ T cells, while IL-5 production was not affected by GSI in *zc3h12a*^{-/-} CD4⁺ T cells. (Fig 5, E and Fig E5, B). These data indicate that Notch mediates the initiation but not the maintenance of IL-5-producing Th2 cells with MCPIP1 deficiency.

MCPIP1 targets Gata3 to control IL-5 production by Th2 cells

Since Gata3 is the master transcription factor for Th2 cell development, we checked Gata3 mean fluorescence intensity (MFI) and found that Gata3 protein was highly expressed in *zc3h12a*^{-/-} CD4⁺ T cells compared to WT CD4⁺ T cells (Fig 6, A). Based on Gata3 levels, we divided *zc3h12a*^{-/-} CD4⁺ T cells into three populations, Gata3^{hi}, Gata3^{mid} and Gata3^{lo}. Gata3^{hi} cells produced higher levels of IL-5 than Gata3^{mid} and Gata3^{low} cells (Fig 6, B). To confirm whether Gata3 accounts for IL-5 overproduction in *zc3h12a*^{-/-} CD4⁺ T cells, we knocked down Gata3 with Gata3-shRNA/lentivirus and found that Gata3 knockdown resulted in reduced IL-5 expression in a dose-dependent manner (Fig 6, C and Fig E6, A), indicating that Gata3 mediates IL-5 production in *zc3h12a*^{-/-} T cells. In addition, we generated a human T cell line (CEM) with an inducible MCPIP1 expression Tet-On system. Addition of Doxycycline (Dox) dose-dependently induced GFP/MCPIP1 fusion protein expression in CEM/MCPIP1-Tet-On cells (Fig E6, B). As a consequence, the expression of Gata3 protein (Fig 6, D) and mRNA (Fig 6, E and Fig E6, C) was reduced significantly. Since MCPIP1 is an RNase²⁵, we measured the half-life of Gata3 mRNA and found that the half-life of Gata3 mRNA was increased in *zc3h12a*^{-/-} CD4⁺ T cells compared to WT CD4⁺ T cells (Fig 6, F) and decreased in CEM/MCPIP1-Tet-On cells after inducing MCPIP1 (Fig 6, G). The Th2-related transcription factor IRF4 mRNA was also stabilized, and STAT6 mRNA unaffected in *zc3h12a*^{-/-} CD4⁺ T cells (Fig E6, D). These data indicate that MCPIP1 inhibits Gata3 mRNA expression through promoting its mRNA decay.

Fully differentiated Th2 cells expressed high levels of Gata3 and low levels of MCPIP1 (Fig E6, E). To test whether MCPIP1 binds to Gata3 mRNA, we purified the

MCPIP1-binding complex with anti-MCPIP1 antibody from cytoplasmic extracts of the Th2 cells and then detected the bound mRNAs by RT-PCR. Compared with controls, Gata3 mRNA was enriched in the mRNAs pulled down with anti-MCPIP1 antibody (Fig E6, F), indicating a direct binding of MCPIP1 to Gata3 mRNA. Next, we transfected a Gata3 3'UTR luciferase construct into CME-Tet-On cells, induced MCPIP1 by adding dox, and then measured luciferase activity. Induction of MCPIP1 suppressed luciferase activity (Fig 6, H), demonstrating that MCPIP1 promotes Gata3 mRNA decay through 3'UTR. As illustrated in Fig E6, G, there are several domains in MCPIP1 protein. To determine which domain mediates Gata3 mRNA decay, we generated CEM-Tet-On stable clones expressing several MCPIP1 mutants, transfected the Gata3-3'UTR luciferase construct into these stable CEM-Tet-On cells, and then induced their expression by adding Dox and measured luciferase activity. Induction of both WT MCPIP1 and mutant MCPIP1 except the mutant D141N inhibited luciferase activity (Fig 6, I). We also measured the levels of Gata3 mRNA in CEM-Tet-On cells after adding Dox. Consistent with the luciferase results, both WT and mutant MCPIP1 except the mutant D141N inhibited the endogenous Gata3 mRNA expression (Fig E6, H). The expression of Gata3 protein displayed similar patterns after expressing WT and mutant MCPIP1 (Fig E6, I). To further confirm whether mutant D141N mediate Gata3 mRNA decay, we measured half-life of Gata3 mRNA in the above CEM-Tet-On cells. The half-lives of Gata3 mRNA were shortened in cells expressing WT MCPIP1 but not in cells expressing the D141N mutants (Fig E6, J), indicating that the RNase domain is required for Gata3 mRNA degradation by MCPIP1. Taken together, these data demonstrate that MCPIP1 via RNase activity destabilizes Gata3 mRNA, therefore inhibits IL-5 and IL-13 production by CD4 T cells.

MCPIP1 deficiency in CD4 T cells enhances Th2-mediated inflammation in allergen-induced asthma models

To determine the physiological consequence of MCPIP1-mediated Th2 inhibition, we immunized WT and *zc3h12a*^{-/-} mice with OVA mixed with adjuvant aluminum hydroxide, and then challenged them with intranasal OVA (Fig 7, A). One day after the last challenge, we stained lung tissue by H&E and PAS. Compared to WT mice, there were increased

infiltrating inflammatory cells and mucin-positive cells (goblet-cell metaplasia) in lungs of the *zc3h12a*^{-/-} mice (Fig E7, A&B). Effector CD4⁺ T cells were also increased after OVA-immunization and challenge in both mice (Fig E7, C). In addition, IL-4- and IL-5-producing effector CD4 T cells were highly increased in the lungs of *zc3h12a*^{-/-} mice compared with WT mice (Fig E7, D). In line with the results shown in Fig 2, D, *zc3h12a*^{-/-} CD4⁺ T cells expressed higher IL-5 and IL-13 but not IL-4 than WT CD4⁺ T cells in response to OVA stimulation *in vitro* (Fig E7, E). These data indicate that MCP1 inhibits OVA-specific CD4 T cell responses *in vivo*, through down-regulating allergen-specific Th2 inflammation mediated by IL-5 and IL-13.

To define whether *zc3h12a*^{-/-} Th2 cells are pathogenic, we first expanded OVA-specific T cells by culturing splenocytes from the above OVA-immunized mice with OVA and IL-2. The OVA-specific *zc3h12a*^{-/-} CD4⁺ T cells secreted large amounts of IL-4, IL-5 and IL-13 in response to OVA stimulation, while IFN- γ production was reduced (Fig 7, B). Next, we purified the OVA-specific CD4⁺ T cells and transferred them intravenously into congenic CD45.1⁺ recipient mice. Three days after T cell transfer, recipient mice were challenged with OVA for 5 days and then analyzed. Mice that received OVA-specific *zc3h12a*^{-/-} CD4⁺ T cells displayed an increase in infiltrating immune cells and inflammation in the airways compared to mice received WT OVA-specific CD4 T cells (Fig 7, C). Lung eosinophils (CD11b⁺SiglecF⁺) were also increased in the mice receiving OVA-specific *zc3h12a*^{-/-} CD4 T cells compared with mice transferred with OVA-specific WT CD4 T cells (Fig 7, D and Fig E7, F), indicating that *zc3h12a*^{-/-} CD4 T cells are pathogenic and can induce asthmatic-like airway inflammation. Analyses of Th2 cytokine transcripts revealed that IL-5 and IL-13 but not IL-4 mRNA were increased in the lungs of the mice receiving OVA-specific *zc3h12a*^{-/-} CD4 T cells (Fig 7, E). The purity of donor cells was confirmed by checking CD3 and CD4 expression gated on CD45.1 and CD45.2 (Fig E7, G). In addition, *zc3h12a*^{-/-} donor CD4 T cells displayed enhanced proliferation and effector phenotypes (Fig 7, F), and produced increased IL-5 and IL-4 *in vivo* (Fig 7, G).

Furthermore, we tested the effects of *zc3h12a*^{-/-} CD4 T cells with physiopathologically

relevant house dust mite extracts (HDM). *Zc3h12a*^{-/-} mice and WT littermates were sensitized and challenged with HDM as illustrated in Fig 8, A. Lung inflammation and PAS positive cells were increased in *zc3h12a*^{-/-} mice compared to WT mice (Fig 8, B). We next expanded HDM-specific CD4 T cells with HDM and IL-2, and checked the HDM-specific responses. *Zc3h12a*^{-/-} CD4⁺ T cells produced significantly more IL-5 and IL-13 than WT CD4⁺ T cells in response to HDM stimulation (Fig 8, C). Then, we transferred the HDM-specific CD4⁺ T cells into WT B6 mice and challenged them with HDM. The mice receiving the HDM-specific *zc3h12a*^{-/-} CD4⁺ T cells displayed more severe lung inflammation and more PAS positive cells than the mice transferred with WT cells (Fig 8, D). The expression of IL-5 and IL-13 were also increased in lungs of the mice received KO HDM-specific CD4 T cells (Fig 8, E). Taken together, the results of the HDM model are consistent with the OVA model, indicating that CD4 T cells deficient in MCP1P1 exaggerate type 2 biased immune responses and cause Th2 inflammation in the airway.

DISCUSSION

We provide several lines of evidence identifying MCPIP1 as a major negative regulator for Th2 cell differentiation and effector function, especially IL-5/IL-13-producing Th2 cells. We discovered enhanced IL-5⁺Th2 differentiation and airway inflammation in MCPIP1 deficient mice. Mechanistically, we identified Notch pathway and Gata3 as critical targets of MCPIP1 for regulating IL-5/IL-13-producing Th2 cell development. CD4⁺ T cells are polarized and differentiated into IL-5-producing Th2 cells in the absence of MCPIP1. MCPIP1 also directly targets Th2 cytokines for mRNA decay (data not shown). The preferential differentiation of naïve *zc3h12a*^{-/-} CD4 T cells towards Th2 but not Th17 or Treg cells *in vitro* may be due to targeting several key transcription factors important for Th2 cell differentiation, including Gata3, Gfi, and Irf4. The mRNAs of Rorc, Tbx21 and Foxp3 were not altered significantly in *zc3h12a*^{-/-} CD4 T cells. IL-4 is a key cytokine for Th2 differentiation. T cells deficient in MCPIP1 lead to IL-4 overproduction that might have an autocrine effect on Th2 differentiation. However, we found that neutralizing IL-4 only inhibited IL-4 expression and had no effects on IL-5 expression in *zc3h12a*^{-/-} CD4 T cells, indicating that the development of IL-5-producing *zc3h12a*^{-/-} Th2 cells is independent of IL-4. MCPIP1 also targets ICOS and OX40²⁸ which have been shown to be important for Th2 differentiation^{42, 43}. Most importantly, our data show that MCPIP1 directly targets the Th2 specific transcript factor, Gata3. Naïve T cells express low levels of Gata3 induced by TCR and IL-4R⁴⁴. Zheng and colleagues¹⁸ show that Gata3 was present in naïve CD4 T cells and increased during Th2 development and extinguished during Th1 development. Gata3 can be induced by both IL-4-dependent and independent way⁴⁴. MCPIP1-deficient CD4⁺ T cells express high levels of Gata3 even under non-polarizing conditions, indicating that MCPIP1 is critical for naïve CD4 T cells to maintain their non-polarized phenotypes. When the levels of MCPIP1 are reduced, CD4 T cells are more likely to polarize and differentiate into IL-5-producing Th2 cells, leading to Th2-associated inflammation. Type 2 innate lymphoid cells (ILC2) share many features with *zc3h12a*^{-/-} Th2 cells and also produce IL-13 and IL-5⁴⁵⁻⁴⁷. ILC2 is regulated by Gata3 and plays an important role in allergic airway inflammation⁴⁸⁻⁵⁰. It will be of interest to see whether MCPIP1 regulates ILC2 as well.

The Notch pathway is important in Th2 cell differentiation and activation⁵¹⁻⁵⁵. Notch-mediated Th2 cell differentiation and Th2 immunity³⁸⁻⁴⁰ are quite similar to the effects of MCPIP1 on IL-5/IL-13-producing Th2 cells, namely both are independent of IL-4 and STAT6 (data not shown). Blockade of Notch signaling inhibits Th2 cytokine production only in naïve but not in effector *zc3h12a*^{-/-} CD4 T cells, suggests that Notch signaling only controls the initiation of Th2 cell differentiation mediated by MCPIP1. Since GSI blocks NICD cleavage from the membrane, it suggests that ligand-receptor interaction upstream of the intracellular Notch pathway mediates the effects of MCPIP1 on Th2 cells. Indeed, the expression of Notch ligands, Jag1/2 and DLL4 were increased in MCPIP1-deficient CD4 T cells (Fig 5, A), suggesting that Jag1/2 and DLL4 may involve in activation of surrounding *zc3h12a*^{-/-} CD4 T cells to attain the Th2 phenotype.

Several studies reported that MCPIP1 protein levels in T cells were decreased in response to anti-CD3/CD28 antibody stimulation^{28, 34}. The degradation and cleavage of MCPIP1 were abolished in the presence of a Malt1 protease inhibitor, as well as in Bcl10 or Malt1 deficient CD4⁺ T cells, indicating that MCPIP1 is cleaved by Malt1²⁸. Inhibition of Malt1 activity leads to reduction of IL-5/IL-13 expression in WT CD4 T cells, but not in *zc3h12a*^{-/-} CD4 T cells, indicating that Malt1/MCPIP1 axis mainly regulates the development of IL-5/IL-13-producing Th2 cells. IL-5 expression is known to be more dependent on the expression levels of Gata3 than IL-4^{56, 57}. In T cell transfer experiments, MCPIP1-deficient Th2 cells were capable of causing enhanced airway inflammation and accumulation of Th2 cytokine-producing cells with a memory phenotype. It was reported that treatment of MCPIP1 deficient mice with antibiotics increased survival and reduced mRNA levels of inflammatory cytokines⁵⁸, suggesting a role of microbiota in MCPIP1-mediated inflammation. MCPIP1 selectively targets the mRNAs encoding Th2 cytokine IL-5 and IL-13 for degradation (data not shown). Why MCPIP1 selectively targets specific cytokines, especially those associated with Th2 cell differentiation and function needs further investigation.

Though the role of Gata3 in regulating Th2 cytokines is well established⁵⁹, the mechanisms that suppress Gata3 expression have not been fully defined. Gata3 can be induced through

TCR, IL-4R/STAT6, IL-2R/STAT5 and Notch signaling during Th2 differentiation. Our work reveals high levels of Gata3 in *zc3h12a*^{-/-} CD4 T cells, and MCPIP1 promotes Gata3 mRNA decay via the RNase domain. We provide strong evidence indicating that MCPIP1 directly regulates Gata3 by binding to its 3'UTR for mRNA destabilization. Blonska et al⁶⁰ showed that CARMA1 up-regulates Gata3 on the transcriptional level independent of NF-κB activation. Given the fact that MCPIP1 is cleaved by Malt1, up-regulation of Gata3 by CARMA1 may involve in cleavage of MCPIP1 by the CARMA1–BCL10–MALT1 complex.

Based on our results, we propose the following model of Th2 cell development regulated by MCPIP1. MCPIP1 inhibits Th2 cell development by binding to the mRNAs encoding multiple Th2 transcription factors, including Gata3, and by inhibiting Th2-associated Notch signaling. Upon T cell activation, MCPIP1 level is reduced and therefore releases its suppression on Th2 transcription factor and Notch pathway, which allows Gata3 to bind to the IL-5 and IL-13 gene loci, resulting in IL-5⁺/IL-13⁺Th2 cell differentiation. Meanwhile, Reduction of MCPIP1 increases the mRNA stability of IL-5 and IL-13. Combination of the increased Th2 differentiation and Th2 cytokine transcripts leads to enhanced Th2 development and Th2 effector function. Therefore, strategies that increase the levels of MCPIP1 will inhibit IL-5⁺/IL-13⁺Th2 cell differentiation and effector function, resulting in suppression of Th2-mediated inflammation.

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SUPPLEMENTAL INFORMATION

Supplemental information includes seven figures and two tables.

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FIGURE LEGENDS

Fig 1. *Zc3h12a*^{-/-} mice develop asthmatic airway inflammation with increased IL-5⁺ Th2 cells. Lung tissue sections were stained with H&E **(A)** and PAS **(B)** (n=4). Cytokines in sera **(C)** and in BAL **(D)** were measured by ELISA (3-4 mice in each group). **(E)** Muc5ac and Gob5 mRNA expression in lung tissues were measured by qRT-PCR (n=3). **(F)** Intracellular cytokine staining in lung infiltrates gated on CD4⁺ T cells and the summary of 3-5 mice per group. *: *p* < 0.05, ***: *p* < 0.001.

Fig 2. Cytokines are overproduced in *zc3h12a*^{-/-} CD4 T cells *in vivo*. Frequency of cytokine⁺ and CD44⁺ splenic cells gated on CD4 after stimulation with P/I **(A and B)** (n=5). **(C)** Summary of cytokine-expressing cells as shown in (B) (three independent experiments with 3-6 mice per group). **(D)** Intracellular cytokine expression in total CD4 T cells after stimulation with anti-CD3/CD28 Abs for 4 days, resting for 2 days and then re-stimulation with P/I (6 mice per group). **(E)** Total CD4⁺ T cells were stimulated by anti-CD3 Ab with or without anti-CD28 Ab for 48 h and supernatants were used to detect cytokines by ELISA (3 mice per group). *: *p* < 0.05, **: *p* < 0.01, ***: *p* < 0.001.

Fig 3. Loss of MCP1P1 leads to enhanced Th2 cytokine production in a cell-intrinsic manner. Naïve WT (CD45.1⁺) and *zc3h12a*^{-/-} (CD45.2⁺) CD4⁺ T cells were co-cultured with anti-CD3/CD28 Abs in the same well for 3 d, rested for 3 d with IL-2, and then treated with P/I for 4 h. Numbers in outlined areas indicate the percentages of cytokine-producing cells **(A)** and a summary of 3-4 mice per group **(B)**. **(C)** The mRNA expression in naïve WT and *zc3h12a*^{-/-} CD4⁺ T cells stimulated with anti-CD3/CD28 Abs for 3 d (n=3). **(D)** The mRNA expression of cytokines in naïve WT and *zc3h12a*^{-/-} CD4⁺ T cells stimulated with anti-CD3/CD28 Abs for 3 days, rested for 3 days, and re-stimulated with P/I for 4 h (n=3). *: *p* < 0.05, **: *p* < 0.01, ***: *p* < 0.001.

Fig 4. Loss of MCP1P1 results in enhanced IL-5⁺ T cell development under unfavorable conditions. Intracellular cytokines were stained in naïve WT and *zc3h12a*^{-/-} CD4⁺ T cells under Th0, Th1, or Th2 polarizing conditions **(A and B)**. The percentages of

cytokine-producing CD4 T cells **(C)** and the mRNA levels **(D)** as in (A and B) were measured and summarized (n=3-4). **(E)** Naïve *zc3h12a*^{-/-} CD4⁺ T cells were stimulated with anti-CD3/CD28 Abs with anti-IL-4 (20 ng/ml), medium, or recombinant IL-4 (10 ng/ml), and stained for Gata3, IL-4 and IL-5 after P/I treatment for 4 h. **(F)** A summary of three independent experiments as in (E) was shown. *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig 5. Notch signal is required for the initiation of IL-5-producing Th2 cells mediated by MCPIP1. **(A)** The mRNA levels of Notch ligands in naïve WT and *zc3h12a*^{-/-} CD4 T cells stimulated with anti-CD3/CD28 Abs were measured by qRT-PCR (n=3). **(B)** Naïve *zc3h12a*^{-/-} CD4 T cells were treated with GSI and anti-CD3/CD28 Abs for 3 days, and then stained for Gata3, IL-4 and IL-5 (n=3). **(C)** A summary of three independent experiments as in (B) was shown. **(D)** Total *zc3h12a*^{-/-} CD4 T cells were treated with GSI and anti-CD3/CD28 Abs for 3 days and then stained for Gata3, IL-4 and IL-5. **(E)** Total CD4 T cells of WT (CD45.2⁻) and *zc3h12a*^{-/-} (CD45.2⁺) mice were cultured in the same well at 1:1 ratio with anti-CD3/CD28 Abs and GSI for 3 d, rested for 3 d, and then stimulated by P/I for 4 h. Cytokine-producing cells were measured by FACS gated on either CD45.2⁺ (*zc3h12a*^{-/-}) or CD45.2⁻ cells (WT). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig 6. MCPIP1 promotes Gata3 mRNA decay. **(A)** Gata3 was measured in naïve WT and *zc3h12a*^{-/-} CD4⁺ T cells stimulated with anti-CD3/CD28 Abs. **(B)** IL-5 in *zc3h12a*^{-/-} CD4⁺ T cells was measured in groups based on Gata3 levels: high (up); intermediate (middle); and low (low) and summarized. **(C)** *Zc3h12a*^{-/-} CD4 T cells were transduced with Gata3/lentivirus or scramble/lentivirus and the percentages of Gata3⁺ and IL-5⁺ were measured by FACS. **(D)** Gata3⁺ and GFP⁺ CEM-MCPIP1-Tet-on cells were measured after Dox treatment (100 ng/ml). **(E)** Gata3 and MCPIP1 mRNA levels in CEM-Tet-on cells were measured after treatment with different amounts of Dox. **(F)** CD4⁺ T cells were stimulated with anti-CD3/CD28 Abs for 4 h, and then added ActD and DRB. Remaining Gata3 mRNA was measured at different times. **(G)** CEM-Tet-on cells were treated with and without Dox (100 ng/ml) for 24 h and then added ActD and DRB. The remaining Gata3 mRNA was measured as in (F). Gata3 3'UTR luciferase reporters or empty vector (Control) were transiently transfected into

CEM-Tet-On cells expressing WT MCP1P1 (**H**) or expressing mutant MCP1P1 (**I**), and then added Dox. Luciferase activity was determined in cell lysates (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig 7. *Zc3h12a*^{-/-} CD4 T cells cause increased Th2 cell-mediated disease. (**A** and **B**) *Zc3h12a*^{-/-} mice and WT littermates were sensitized and then challenged by OVA and sacrificed on day 26. Cells isolated from DLN were stimulated with different amounts of OVA for 72 h and cytokines in the supernatants were measured by ELISA (n=3). (**C**) CD45.1 recipient mice were transferred with OVA-specific CD4 T cells as in (B) and then challenged daily with OVA for 5 times. H&E-stained lung sections of the recipient mice were shown and inflammation was scored. (**D**) The percentages of eosinophil and neutrophil in BAL of the CD45.1⁺ recipient mice after T cell transfer were gated on SiglecF and Ly6G (n=3). (**E**) mRNA levels in lungs of the recipient mice were measured by qRT-PCR (n=3). (**F**) The percentages of CD44⁺CD45.1⁻ (donor) CD4 T cells were measured in the CD45.1⁺ recipient mice. (**G**) Intracellular IL-5 and IL-4 in the donor cells were stained as in (F) gated on CD44 (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig 8. HDM-induced Th2 inflammation is increased in *zc3h12a*^{-/-} mice. (**A**) *Zc3h12a*^{-/-} mice and WT littermates were sensitized and then challenged by HDM. (**B**) Mice were sacrificed on day 26. Lung sections were stained with H&E and PAS and inflammation was scored (n=3). (**C**) Cells isolated from DLN as in (B) were stimulated with HDM for 72 h. IL-5 and IL-13 in supernatants were measured by ELISA (n=3). (**D**) WT B6 mice were transferred with HDM-specific CD4 T cells as in (C) and then sensitized with HDM. Representative H&E- and PAS-stained lung sections were shown from the recipients received HDM-treated WT or *zc3h12a*^{-/-} cells and inflammation was scored (five mice each group). (**E**) Cells from lungs of the recipient mice as in (D) were stimulated with P/I for 4 h and then RNA was extracted to measure mRNAs by qRT-PCR (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig.1.

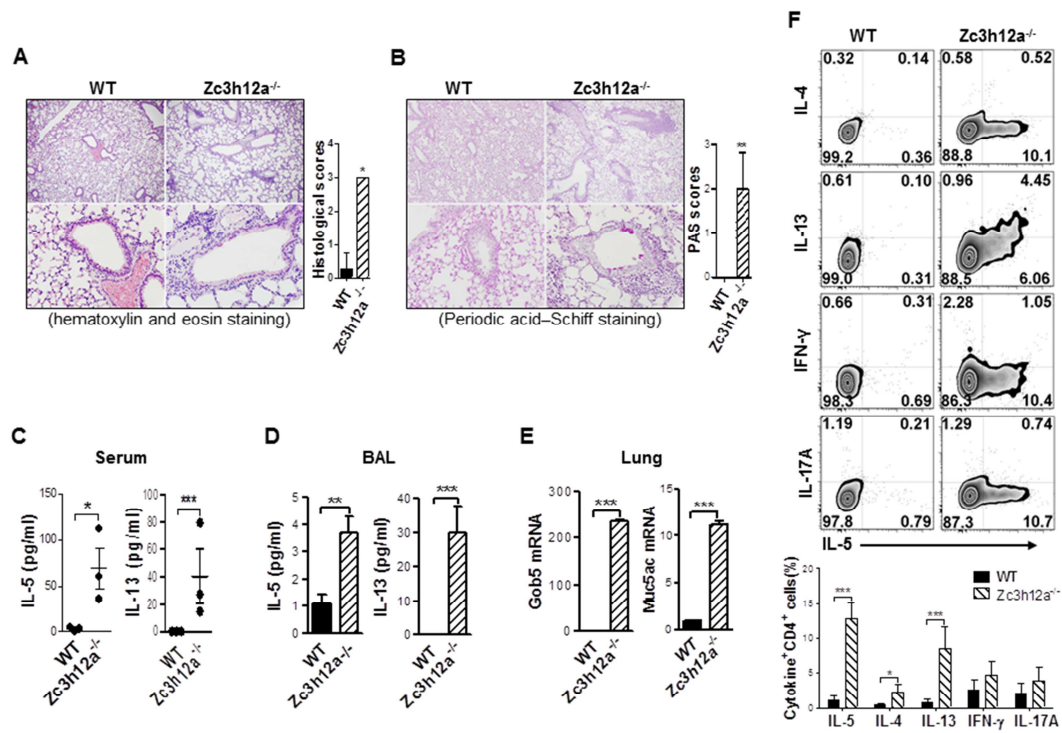


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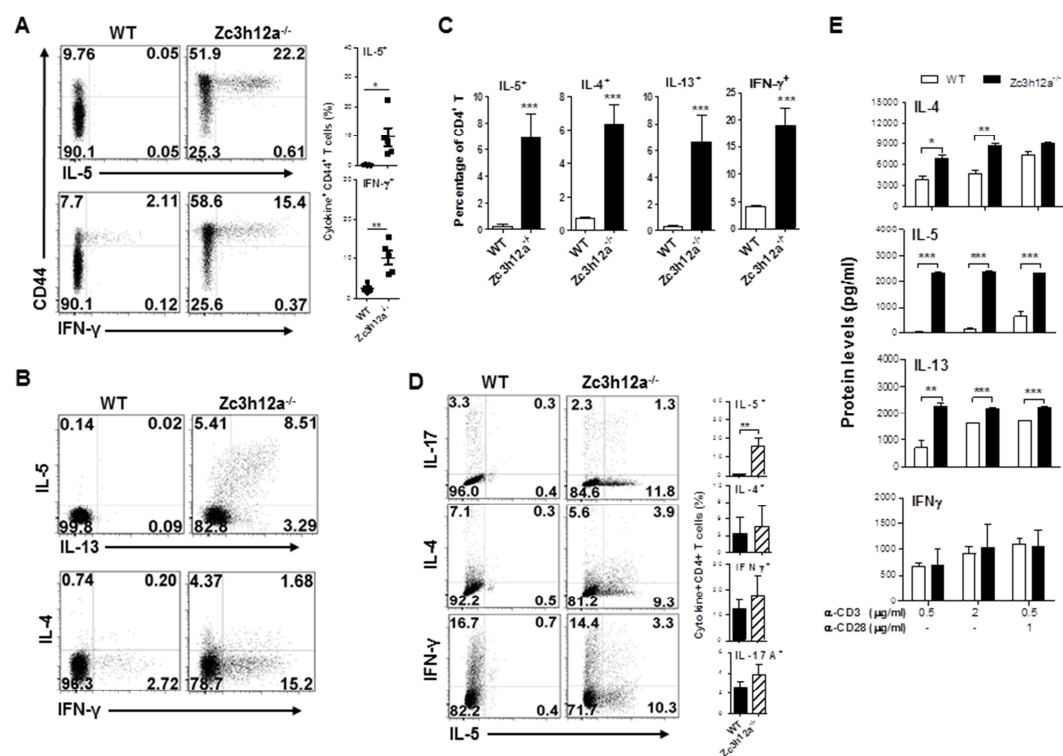


Fig. 4.

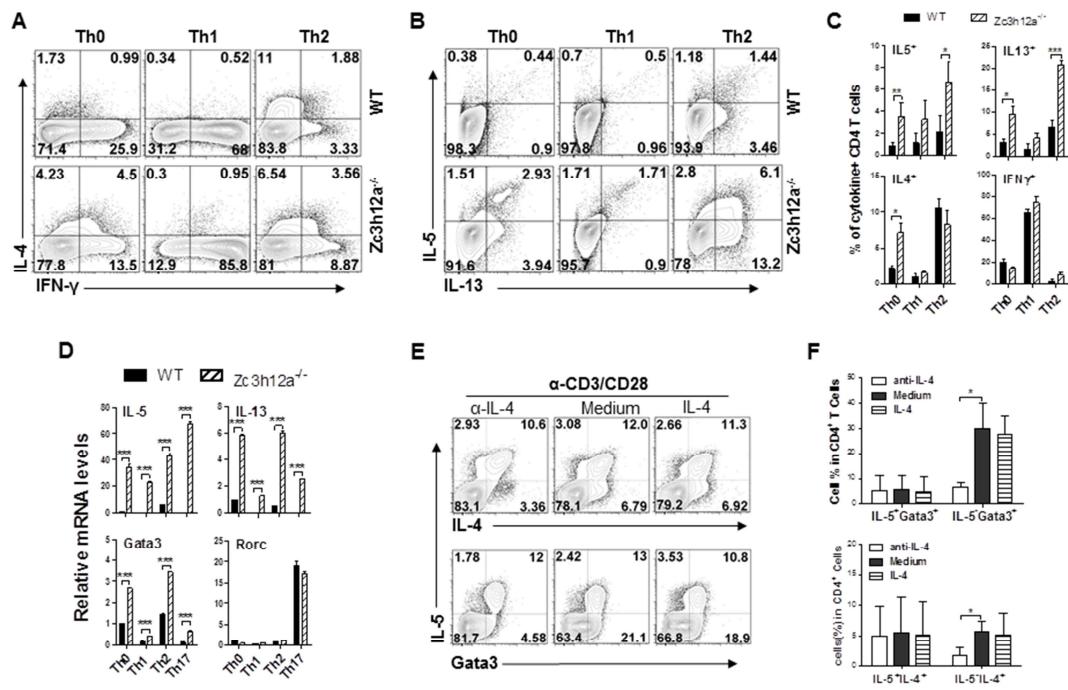


Fig.5.

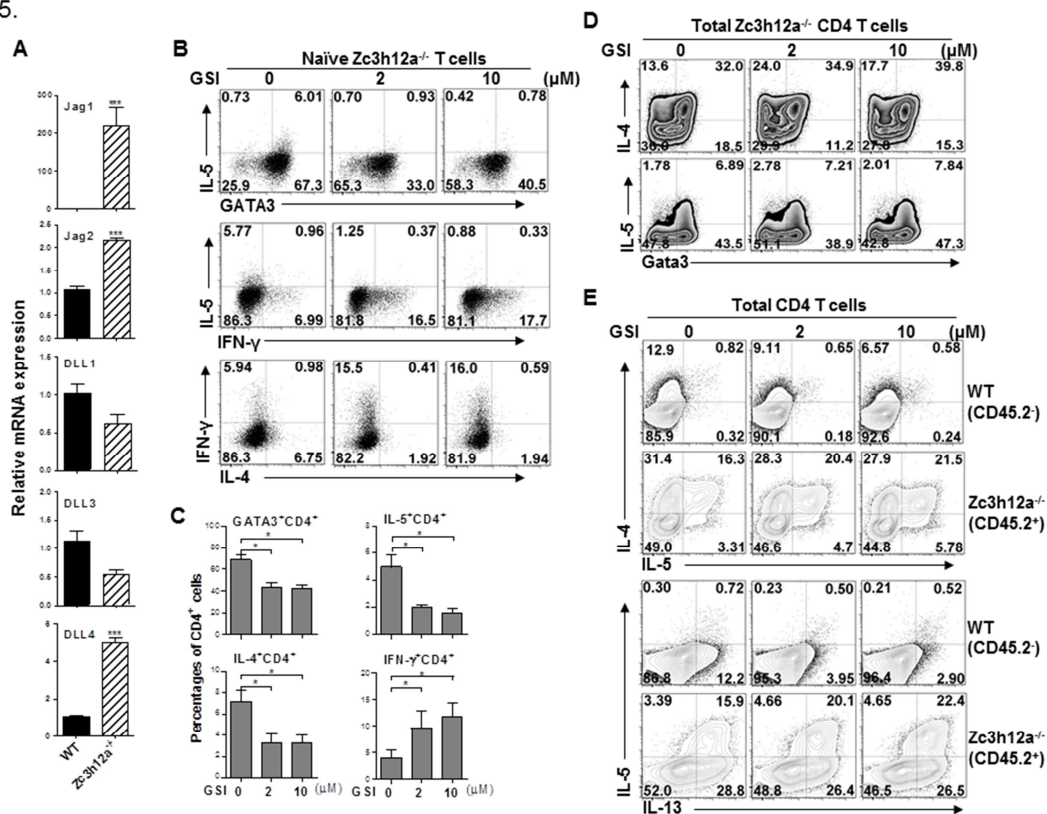


Fig. 6.

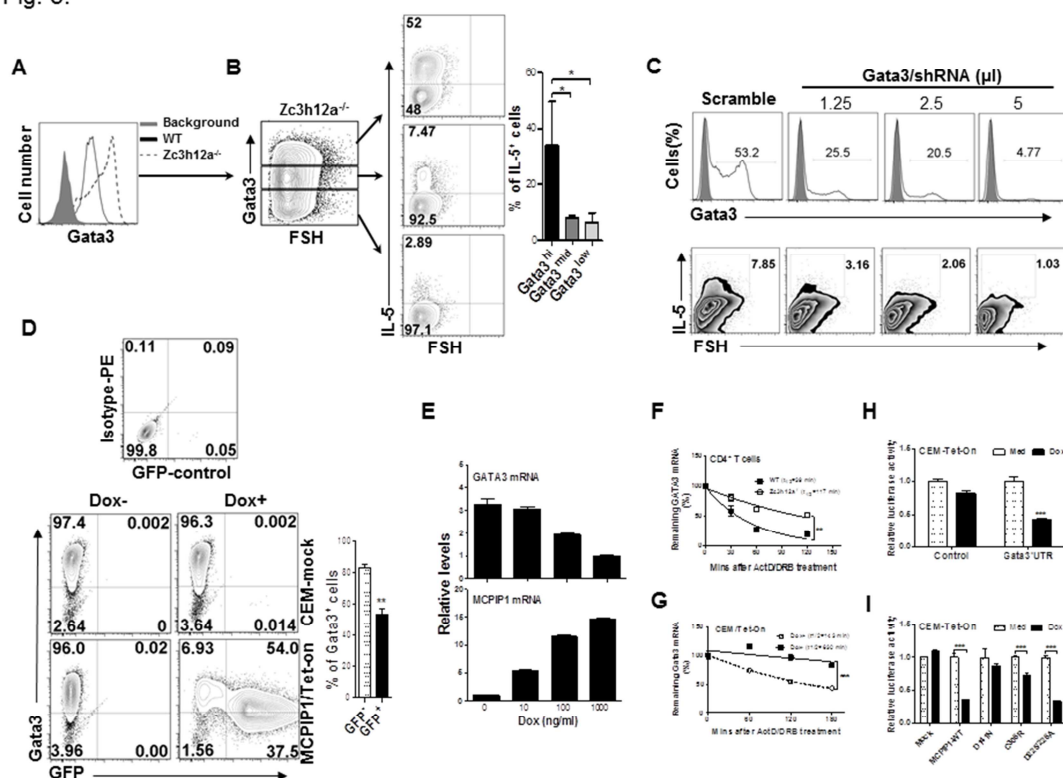


Fig. 7.

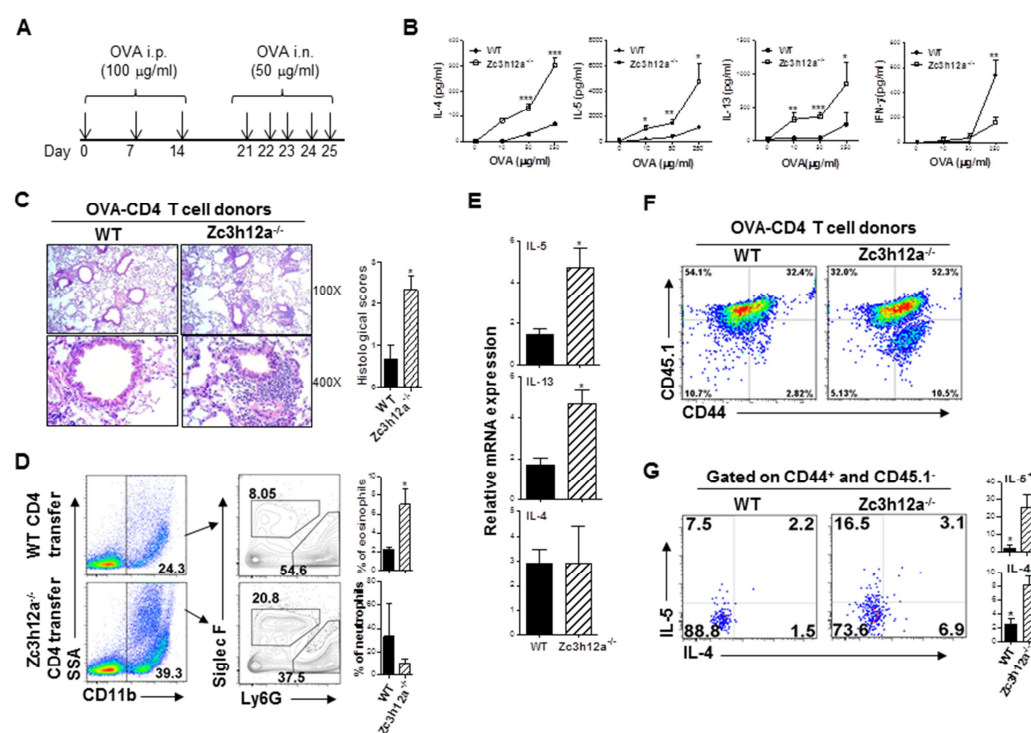
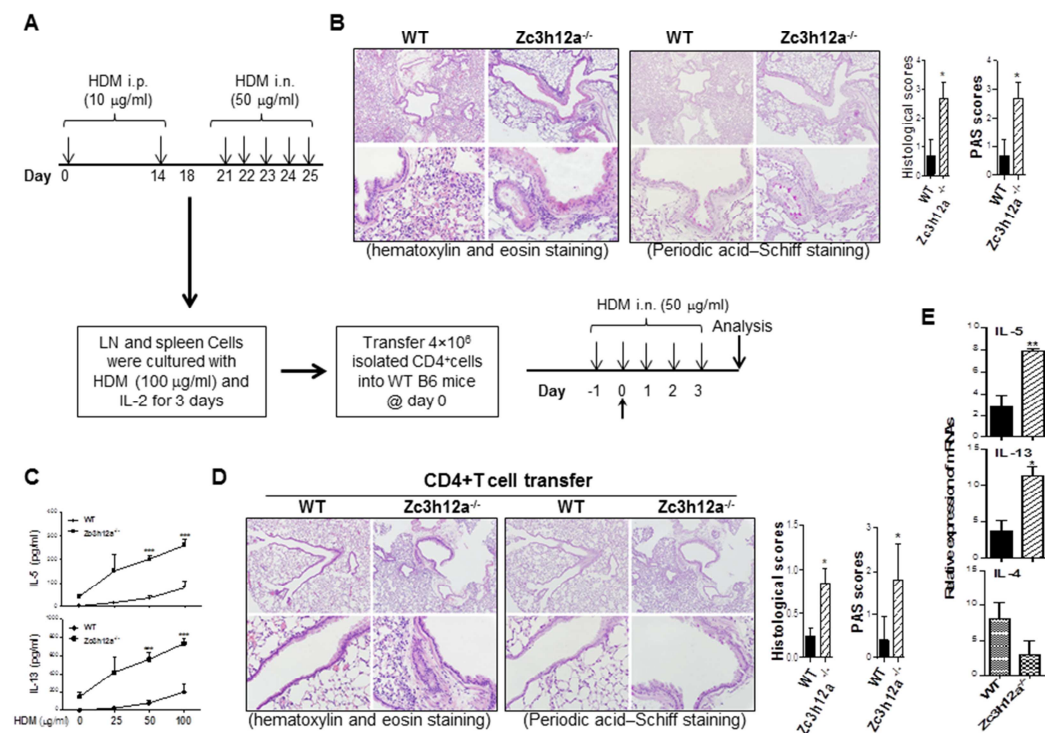


Fig. 8.



SUPPLEMENTARY METHODS

Antibodies and flow cytometry

For surface staining, cells were stained at 4 °C in the presence of Fc Block (2.4G2; BD Biosciences) in flow cytometry buffer (DPBS plus 2% FCS). After staining surface markers, cells were fixed and permeabilized for intracellular cytokine and transcription factor staining according to the manufacturer's instructions (BD Biosciences). Antibodies used for surface and intracellular staining were listed in the supplementary Table 2.

T cell lentiviral transduction

The lentiviral vector Gata3-shRNA and scramble-shRNA were purchased from Sigma. Lentivirus was produced in HEK293T cells. CD4⁺ T cells were stimulated for 24 h with plate-bound anti-CD3 Ab (0.5 µg/ml) and then transduced with virus-containing medium supplemented with polybrene (8 µg/ml). 20 hours later, culture medium was replaced with complete medium with anti-CD3 Ab (0.5 µg/ml) for 2 days, rested for 2 days and then stimulated with PMA and Ionomycin for 4 hours for intracellular staining.

T cell stimulation

Mouse CD4⁺ T cells were negatively purified and activated by plate-coated anti-CD3 (1 µg/ml) and anti-CD28 (1 µg/ml) Abs for 72 h in complete cell medium (RPMI1640) supplemented with 10% heat-inactivated FBS, 2 mM glutamine, penicillin-streptomycin, nonessential amino acids, sodium pyruvate, 10 mM HEPES and 50 µM 2-mercaptoethanol, then expanded for another 3 day in fresh medium containing 1 ng/ml recombinant mouse IL-2. In some experiments, cells cultured for 3 days and supernatants were collected for measuring cytokines by ELISA assay.

For isolation of infiltrating cells from lungs, lungs were minced and digested in 5 ml of RPMI1640 media plus 10% FBS with 250 µg/ml collagenase IV (Roche) and 30 U/ml DNase I (Sigma-Aldrich) for 60 min at 37 °C with shaking. Cells were passed through a 70-µm strainer and spin down. The cell pellet was resuspended in 40% Percoll, carefully layered onto 80% Percoll and centrifuged at 650 g, without brake, RT for 30 min, then collected the

layer in the interface between the two Percoll concentrations. Cells were washed with PBS before red blood cells were lysed with ACK lysis buffer. Cells were counted and 2×10^6 cells were activated with PMA and ionomycin for analysis of intracellular cytokines and surface markers by flow cytometry.

Tet-On System

Wild-type or mutant MCP1P1 (D141N, C306R, D225/226A) expressing CEM Tet-on cells were generated as described previously¹. The cells were cultured for 24 hours with Dox (100 ng/ml) and then treated with PMA (10 ng/ml) and Ionomycin (0.5 μ g/ml) for 4 hours. Total RNA was isolated and used to measure gene expression. For western blot, CEM Tet-On cells were treated with Dox (100 ng/ml) for 48 h and then lysed.

Luciferase assay

CEM-Tet-on cells were transfected with luciferase reporter plasmid containing the 3' UTR of Gata3² using Nucleofector (Lonza). MCP1P1 expression was induced by Dox (100 ng/ml). Cells were lysed after 24 hours and luciferase activity in lysates was determined with Dual-Luciferase Reporter Assay system (Promega).

Quantitative RT-PCR

Quantitative PCR was performed with SYBR green (Invitrogen, Life Technologies) using an Applied Biosystems StepOne PCR system. Results were analyzed with $\Delta\Delta$ CT method with glyceraldehyde-3-phosphate dehydrogenase (GAPDH) as an endogenous reference control. Primer sequences are listed in supplementary table1.

ELISA

Cell culture supernatants were assayed by ELISA for mouse IL-4, IFN- γ (BD Biosciences), IL-5 and IL-13 (eBioscience), according to the manufacturers' instructions.

RNA Immunoprecipitation

Th2-polarized cell lysates were prepared from WT mice after 6 d of *in vitro* polarization. Cell lysates were prepared in NT-2 buffer (50 mmol/L Tris-HCl, pH 7.4, 150 mmol/L NaCl,

1 mmol/L MgCl_2 and 0.05% NP-40) supplemented with 1 mmol/L dithiothreitol, 0.25 U/mL RNase out and $1\times$ protease inhibitor and incubated on ice for 10 min. Cell lysates were obtained after high-speed centrifugation at 15,000 g for 15 min. Equal amounts of lysates were used for immunoprecipitation with MCPIP1 antibody and IgG1 control. Anti-MCPIP1 Ab or IgG1 were pre-coated onto protein A Sepharose beads and incubated with cell lysates. Protein A Sepharose beads were then washed with NT-2 buffer and incubated with 20 U RNase-free DNase I (15 min, 30°C) and further incubated in 100 μL NT-2 buffer containing 0.1% SDS and 0.5 mg/mL proteinase K (30 min, 55°C). The total RNAs were isolated by adding Trizol reagent (Invitrogen) to the beads and quantitative PCR (qPCR) was performed to measure GAPDH and GATA3.

Histological analysis

Peribronchiolar and perivascular inflammation were determined by a semiquantitatively graded scale as follows: 0, no inflammation; 1, mild inflammation; 2, moderate inflammation; 3, severe inflammation. Goblet-cell hyperplasia was assessed as follows: 0, no or rare PAS⁺ cells; 1, <10% PAS⁺ cells observed at a magnification of $\times 10$; 2, 10–25% PAS⁺ cells observed at a magnification of $\times 10$; and 3, >25% PAS⁺ cells observed at a magnification of $\times 4$. A composite score was determined by multiplication of the graded score and the frequency of airways or vessels with a given pathological finding. All slides were examined in a random order and scored by a blinded pathologist.

References

1. Liu S, Qiu C, Miao R, et al. MCPIP1 restricts HIV infection and is rapidly degraded in activated CD4⁺ T cells. *Proc Natl Acad Sci U S A*. Nov 19 2013;110(47):19083-19088.
2. Yang Y, Ahn YH, Gibbons DL, et al. The Notch ligand Jagged2 promotes lung adenocarcinoma metastasis through a miR-200-dependent pathway in mice. *J Clin Invest*. Apr 2011;121(4):1373-1385.

FIGURE LEGENDS

Fig E1. Cell subsets in BAL of *zc3h12a*^{-/-} mice. Total number of cells in the BAL of WT and *Zc3h12a*^{-/-} mice were counted and summarized (3-4 mice per group). *: $p < 0.05$.

Fig E2. Effector CD4 T cells are increased in spleen of *zc3h12a*^{-/-} mice. **(A)** The expression of CD62L and CD44 in CD4⁺ T cells of WT and *zc3h12a*^{-/-} mice were measured by flow cytometry and summarized (n=3). **(B)** Percentages of CD8 and CD4 T cells in spleens of *zc3h12a*^{-/-} mice and WT littermates were gated on CD3 (n=3). **(C)** The levels of IL-5 and IL-13 in culture supernatants of splenic CD4⁺ T cells were measured by ELISA after stimulation with anti-CD3/CD28 Abs (1 μ g/ml) for different times (n=3). **(D)** Splenic cells were stimulated with P/I for 4 h and the percentages of IL-5⁺ and IFN- γ ⁺ CD4 cells were measured by flow cytometry gated on CD44⁺ (n=4). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig E3. Gene expression profiles of *zc3h12a*^{-/-} CD4 T cells. The mRNA levels of Th2-related genes were measured by qRT-PCR in naïve WT littermate and *zc3h12a*^{-/-} CD4 T cells stimulated with anti-CD3/CD28 Abs for 3 days (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig E4. IL-5 and IL-13 expression depend on MCPIP1 in CD4 T cells. **(A-C)** WT littermate and *zc3h12a*^{-/-} CD4⁺ T cells were pretreated with or without Malt1-specific inhibitor MI-2 (1 μ M) for 30 min and then stimulated with anti-CD3/CD28 Abs for 4 hrs. The mRNA expression of cytokines was measured by qRT-PCR (n=3). *: $p < 0.05$, **: $p < 0.01$.

Fig E5. Notch signal is required for the initiation of IL-5-producing Th2 cells mediated by MCPIP1. **(A)** The percentages of Gata3⁺*zc3h12a*^{-/-} CD4⁺T cells were analyzed after pretreatment with GSI and then with anti-CD3/CD28 Abs (n=3). **(B)** Total CD4⁺ T cells of WT and *zc3h12a*^{-/-} were pretreated with GSI and then cultured with anti-CD3/CD28 Abs. Cytokine-producing cells were gated on CD4 and summarized (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig E6. MCPIP1 inhibits the expression of Th2-associated transcription factors. **(A)** The

percentages of $Gata3^{+}Zc3h12a^{-/-}$ $CD4^{+}$ as in Fig. 6C were summarized from three independent experiments. **(B)** The expression of GFP/MCPIP1 fusion protein in CEM-Tet-On cells after adding Dox. **(C)** CEM-Tet-On cells were treated with Dox and P/I, followed by measuring Gata3 and IL-13 mRNA expression by qRT-PCR (n=3). **(D)** $CD4^{+}$ T cells from WT and $zc3h12a^{-/-}$ mice were stimulated with anti-CD3/CD28 Abs and ActD/DRB. Then total RNAs were harvested for measuring mRNA levels by qRT-PCR. **(E)** The levels of Gata3 and MCPIP1 mRNA in WT $CD4^{+}$ Th2 cells were measured by qRT-PCR. **(F)** Cytoplasmic extracts were collected from the Th2 cells and immunoprecipitated with anti-MCPIP1 or isotype matched Ab, followed by RNA extraction. Gata3 mRNA was detected by PCR (n=3). **(G)** A schematic diagram for putative functional conserved domains of MCPIP1. **(H)** The expression of Gata3 mRNA in CEM Tet-On cells after adding Dox and stimulating with P/I for 3 h (n=3). **(I)** The MFI of Gata3 in CEM-Tet-On cells after adding Dox and then stimulating with P/I. **(J)** Stable CEM-Tet-On cells were treated with Dox and P/I, and then treated with ActD and DRB for different times. The remaining Gata3 mRNA was measured by qRT-PCR (n=3). *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Fig E7. MCPIP1 deficiency enhances Th2 cell-mediated disease in an allergen-induced asthma model. WT and $zc3h12a^{-/-}$ mice were sensitized and challenged with OVA as in Fig. 7A. H&E and PAS staining of lung tissue sections were examined on day 26 **(A)** and inflammation was scored **(B)** (n=3). **(C)** The percentages of $CD44^{high}$ cells were gated on $CD4^{+}$ T cells in spleens of the mice as in (A) and summarized (n=3). **(D)** Splenocytes from the mice in (A) were stimulated with P/I and the percentages of $IL-5^{+}$ and $IL-4^{+}$ $CD4^{+}$ T cells were measured and analyzed (n=4). **(E)** Splenocytes from the mice in (A) were re-stimulated with OVA (200 μ g/ml) and IL-2 (10 ng/ml) for 3 days, and stimulated with P/I. The percentages of cytokine-producing $CD4^{+}$ T cells were measured by FACS. **(F)** Cell subtypes in BAL of the CD45.1 recipient mice as in Fig. 7C were counted and summarized (n=4). **(G)** Splenocytes from the CD45.1⁺ recipient mice as in (D) were used to detect donor cells ($CD45.2^{+}$) and host cells ($CD45.1^{+}$) gated on $CD3^{+}CD4^{+}$ and $CD3^{+}CD4^{-}$ cells. *: $p < 0.05$, **: $p < 0.01$, ***: $p < 0.001$.

Table E1.**Primer sequences**

Gene symbol	Forward primer (5'---3')	Reverse primer (5'—3')
mIrf4	5'-GCAGCCTTCAGGGCTCGTCG	5'-GCAGCCTTCAGGGCTCGTCG
mIl4	5'-GGTCTCAACCCCCAGCTAGT	5'-GCCGATGATCTCTCTCAAGTGAT
mIl5	5'-CTCTGTTGACAAGCAATGAGACG	5'-TCTTCAGTATGTCTAGCCCCCT
mIl13	5'-CCTGGCTCTTGCTTGCCT	5'-GGTCTTGTGTGATGTTGCTCA
mIfng	5'-ATGAACGCTACACACTGCATC	5'-CCATCCTTTTGCCAGTTCCTC
mIl17a	5'-CTCCAGAAGGCCCTCAGACTAC	5'-GGGTCTTCATTGCGGTGG
mTgf-b	5'-TAAGAGGTCACCCGCGTGCT	5'-AAAGACAGCCACTCAGGCGTA
mCcl24	5'-ATTCTGTGACCATCCCTCAT	5'-TGTATGTGCCTCTGAACCCAC
mMuc5ac	5'-CTCCTCTCGGTGACAGAGTCT	5'-GTGGTTTGACACTGACTTCCC
mGob5	5'-CGTGGTCATTGGCGATGACG	5'-CTGTCTTCCTCTTGATCCTCCA
mSocs3	5'-ATGGTCACCCACAGCAAGTTT	5'-TCCAGTAGAATCCGCTCTCCT
mGfi	5'-GATGAGCTTTGCACACTGGA	5'-AGGAACGCAGCTTTGACTGT
mRorc	5'-AAGATCTGCAGCTTTTCCACA	5'-TTTGGAAGTGGCTTTCCATC
mGata3	5'-CTCGGCCATTTCGTACATGGAA	5'-GGATACCTCTGCACCGTAGC
mStat6	5'-CTCTGTGGGGCCTAATTTCCA	5'-CATCTGAACCGACCAGGAACT
mTbx21,	5'-TTTCCAAGAGACCCAGTTCATTG	5'-ATGCGTACATGGACTCAAAGTT
mFoxp3	5'-CCCATCCCCAGGAGTCTTG	5'-ACCATGACTAGGGGCACTGTA
mIcosl	5'-TAAAGTGTCCTGTTTTGTGTCC	5'-ATTGCACCGACTTCAGTCTCT
mCd40	5'-TGTCATCTGTGAAAAGGTGGTC	5'-ACTGGAGCAGCGGTGTTATG
mOx40l	5'-GGGATGCTTCTGTGCTTCATCT	5'-TGTTCTGCACCTCCATAGTTTGA
mCcr3	5'-TCAACTTGGCAATTTCTGACCT	5'-CAGCATGGACGATAGCCAGG
mCxcr4	5'-AGCTAAGCAGCATGAACAAC	5'-AACGCTGCTGTAGAGGTTGACACT
mCcl5	5'-TCGTGCCCACGTCAAGGAGTA	5'-TAGAGCAAGCAATGACAGGGA
mJag1	5'-CCTCGGGTCAGTTTGAGCTG	5'-CCTTGAGGCACACTTTGAAGTA
mJag2	5'-CAATGACACCACTCCAGATGAG	5'-GGCCAAAGAAGTCGTTGCG
mDll1	5'-CAGGACCTTCTTTTCGCGTATG	5'-AAGGGGAATCGGATGGGGTT
mDll3	5'-CTGGTGTCTTCGAGCTACAAAT	5'-TGCTCCGTATAGACCGGGAC

mDl14	5'-TTCCAGGCAACCTTCTCCGA	5'- ACTGCCGCTATTCTTGTCCC
mGapdh	5'- AACTTTGGCATTGTGGAAGG	5'- ACACATTGGGGGTAGGAACA
hGata3	5'-GCCCCCTCATTAAGCCCAAG	5'-TTGTGGTGGTCTGACAGTTCG
hZc3h12a	5'-GGCAGTGGTTTCTTACGAAGGA	5'-CCCATCACAGACCAGCACAT
hIl13	5'-CCTCATGGCGCTTTTGTGAC	5'-TCTGGTTCTGGGTGATGTTGA
pLA-up	5'-ATGCCTTCCTGATCCTATTGG	
pLA-low		5'-CTTTCATGAGCAATGGTCGCA
pLaz-low		5'-GTGCGGGCCTCTTCGCTATTAC

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154 **Table E2.**155 **Antibody list**

Antibody	clone	Company
anti-CD3-APC	145-2C11	BD Biosciences
anti-CD4-PE	H129.19	BD Biosciences
anti-CD8-PE	53-6.7	BD Biosciences
anti-CD4-PerCP	RM4-5	BD Biosciences
anti-CD62L-PE	MEL-14	BD Biosciences
anti-CD44-APC-cy7	IM7	BD Biosciences
anti-CD44-FITC	IM7	BD Biosciences
anti-CD11b-FITC	M1/70	BD Biosciences
anti-CD45.1-AF700	A20	BD Biosciences
Anti-CD45-pecy7	104	BD Biosciences
anti-Ly6G-APC	1A8	BD Biosciences
anti-Siglec F-PE	E50-2440	BD Biosciences
Siglec F-PE	E50-2440	BD Biosciences
anti-IFN- γ -APC	XMG1.2	BD Biosciences
anti-IL-17A-PE	TC11-18H10.1	BD Biosciences
anti-IL-17A-AF488	TC11-18H10.1	BD Biosciences
anti-IL4-AF488	11B11	BD Biosciences
anti-IFN- γ -AF700	XMG1.2	BD Biosciences
anti-IL-5-APC	TRFK5	BD Biosciences
anti-Gata3-PE	C50-823	BD Biosciences
anti-IL-13-PercPeF710	eBio13A	eBioscience
anti-CD11b-FITC	M1/70	Biolegend
anti-CD45-PE-Cy7	N418	Biolegend

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Figure E1.

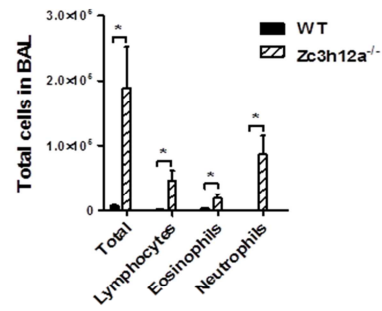


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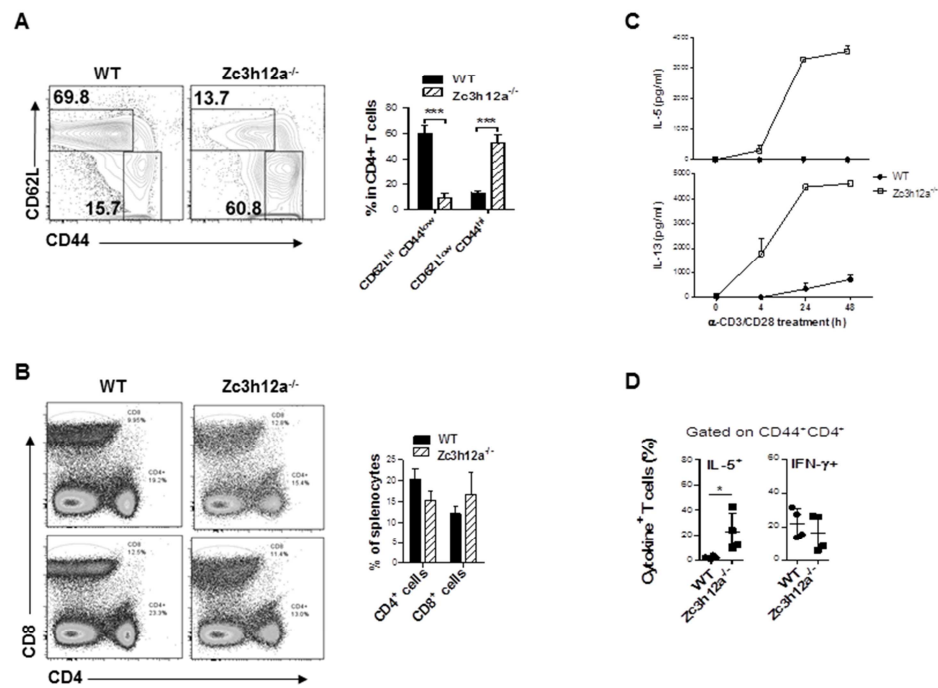


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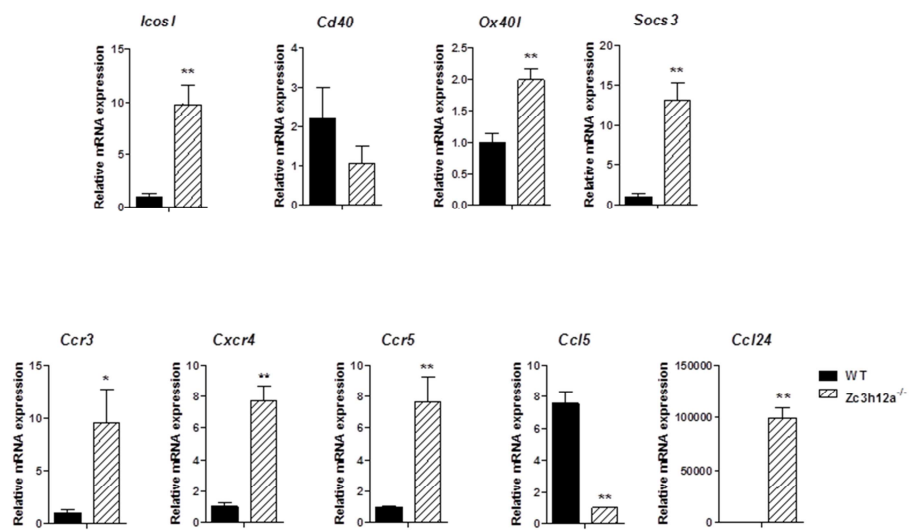


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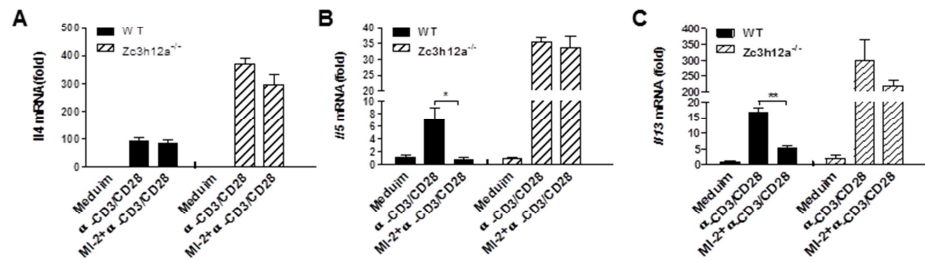


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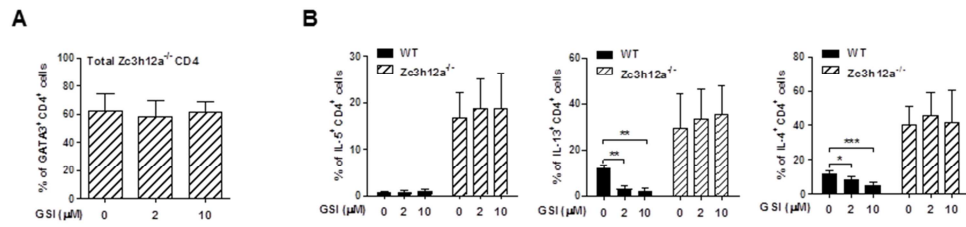


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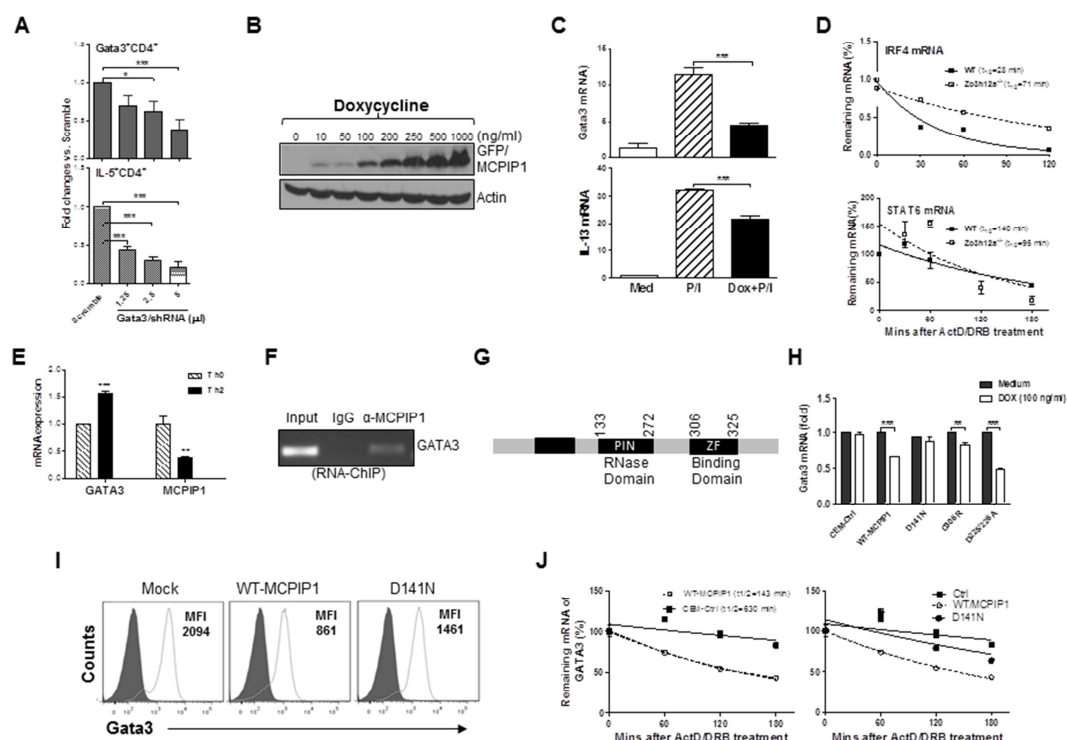


Figure E7.

