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Protein expression in female salivary glands of pyrethroid-susceptible and resistant strains of *Aedes aegypti* mosquitoes

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Abstract

Background: A group of insecticides called pyrethroids has been used extensively worldwide and development of pyrethroid resistance within mosquito populations, especially in *Aedes aegypti*, has rapidly spread through populations. In this study, SDS-PAGE, 2-DE coupled with NanoLC-MS, and bioinformatics were used to analyze the female salivary gland proteins of pyrethroid-susceptible (PMD) and pyrethroid-resistant (PMD-R and UPK-R) strains of *Ae. aegypti* mosquitoes for the first time.

Results: SDS-PAGE analysis revealed that among the three strains at least nine major proteins were detected but one protein band (20 kDa) was found only in the PMD strain. Two-dimensional gel electrophoresis analysis revealed 19 similarly expressed proteins in the salivary glands of the three strains involved in blood-feeding process, stress response, immunogenic response, and metabolic process and five additional major protein spots differentially expressed in the susceptible and resistant strains. Comparative analysis of the expression volume of each protein spot between the PMD and the PMD-R strains showed three downregulated proteins of the PMD-R mosquitoes. For UPK-R strains, six major proteins were downregulated when compared to the PMD strain. Additionally, four downregulated proteins were found in the UPK-R when compared to the PMD-R strain. These results suggest that pyrethroids might induce alteration of salivary gland proteins in resistant mosquitoes. Network analysis by STITCH database 5.0 showed that SRPN23 interacted with sodium and calcium ions, suggesting that SRPN23 might be involved in insecticide resistance.

Conclusions: Information obtained from this study will be useful for further studies on the roles of differentially expressed salivary gland proteins in resistance to insecticides and viral transmission.

Keywords: *Aedes aegypti*, Insecticide resistance, Pyrethroids, Salivary glands, Mosquito salivary proteins

Background

Aedes aegypti mosquitoes serve as the primary vector of dengue hemorrhagic fever (DHF), yellow fever, chikungunya fever and Zika fever. The diseases are considered as serious public health problems in several countries in tropical and subtropical areas. The diseases cause mortality and morbidity of populations; however, there is a lack of vaccines or specific treatments. Therefore, mosquito

control is an essential method to control the transmission of the diseases. One of the popular methods used to reduce the population of mosquitoes is the application of chemical compounds such as insecticides. However, this method is impeded by the development of resistance within mosquito populations [1, 2].

In Thailand, at least four groups of synthetic compounds, organochlorine (DDT), organophosphates (temephos, fenitrothion, malathion and chlorpyrifos), carbamates (propoxur, pirimiphosmethyl and bendiocarb) and pyrethroids (permethrin, deltamethrin, lambda-cyhalothrin and etofenprox) have been

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extensively used to control mosquito vectors. At present, several mosquitoes have been reported as resistant to insecticides, especially to DDT, pyrethroids (i.e. permethrin and deltamethrin), carbamates (i.e. propoxur) and organophosphates (i.e., temephos and fenitrothion) [3, 4].

The insecticide susceptible Pang Mei Daeng (PMD) and Pang Mei Daeng resistant (PMD-R) strains of *Ae. aegypti* from Chiang Mai, Thailand, have been used in several previous studies [5–9]. The PMD strain is susceptible to pyrethroids, but resistant to DDT, which has mainly been attributed to increased DDTase activity [5]. No *kdr* mutations (S989P, V1016G and F1534C) have been found in this strain [10]. PMD-R (S/S989 + V/V1016 + C/C1534, or SS + VV + CC) is homozygous for C1534, lacks S989P and V1016G *kdr* mutations, and is resistant to both DDT and permethrin but susceptible to deltamethrin [10–12]. Recently, the Upakhut resistant (UPK-R) strain has also been established from mosquitoes collected from Wat Upakhut in the city of Chiang Mai. The UPK-R strain is homozygous for the G1016 *kdr* allele and resistant to DDT, permethrin and deltamethrin. It harbors P/P989 + G/G1016 + F/F1534, or PP + GG + FF [13–15].

An important role of the mosquito salivary glands is to modulate host responses that facilitate transmission of pathogens. Dengue virus is transmitted to a vertebrate host while saliva of infected female mosquitoes is injected into the host. A recent study demonstrated that *Ae. aegypti* salivary gland extract enhances dengue pathogenesis after infection [16]. In previous studies, differential expression of salivary gland proteins between insecticide-susceptible and resistant strains of *Culex quinquefasciatus* [17], *Anopheles gambiae* [18] and *Anopheles stephensi* [19] has been reported. Regarding insecticide resistant mosquitoes in Thailand, no report of differentially expressed salivary gland proteins between susceptible and insecticide-resistant *Ae. aegypti* mosquitoes is available.

In the present study, SDS-PAGE, two-dimensional gel electrophoresis (2-DE) coupled with mass-spectrometry

(NanoLC-MS), and bioinformatics were used to compare salivary gland expression profiles and identify the differentially expressed proteins in the three *Ae. aegypti* strains, PMD, PMD-R and UPK-R. The information on proteins in the salivary glands of these insecticide-resistant mosquitoes might help to explain their impact on vectorial capacity.

Results

Insecticide susceptibility test

To confirm the insecticide susceptibility status of the *Ae. aegypti* PMD, PMD-R, and UPK-R strains, the mosquito strains were tested according to the WHO standard method [20]. The results revealed that the PMD strain was susceptible to both permethrin and deltamethrin, while the PMD-R strain was resistant to permethrin with 13.33% mortality and the UPK-R strain was resistant to permethrin and deltamethrin with zero and 1.67% mortality, respectively (Table 1).

Comparison of female salivary gland protein profiles of *Ae. aegypti* mosquitoes between susceptible and resistant strains by SDS-PAGE

A comparative analysis of the female salivary gland protein profiles among the PMD, PMD-R and UPK-R strains was performed using SDS-PAGE. The SDS-PAGE results showed that nine major proteins were detected in the three strains (Fig. 1). The molecular weights of these protein bands were estimated to be 66, 60, 49, 47, 37, 31, 29, 18 and 16 kilodaltons (kDa), respectively. However, one protein band (20 kDa) was found only in the PMD strain. As a protein band may consist of more than one protein, 2-DE was therefore used for further detailed analysis.

Comparison of female salivary gland protein profiles of *Ae. aegypti* mosquitoes between susceptible and resistant strains by 2-DE

Protein profiles of the female mosquitoes of the PMD, PMD-R and UPK-R strains were also analyzed by 2-DE

Table 1 Insecticide susceptibility status of *Ae. aegypti* PMD, PMD-R and UPK-R strains

Strain	Insecticide	Total no. of tested mosquitoes	Total no. of mosquito deaths	Mortality rate (%)	Status ^a
PMD	Permethrin	300	300	100	S
	Deltamethrin	300	300	100	S
PMD-R	Permethrin	300	40	13.33	R
	Deltamethrin	300	300	100	S
UPK-R	Permethrin	300	0	0	R
	Deltamethrin	300	5	1.67	R

^a Interpretation of mortality rate (WHO, 2013)

Abbreviations: R, resistance; S, susceptibility

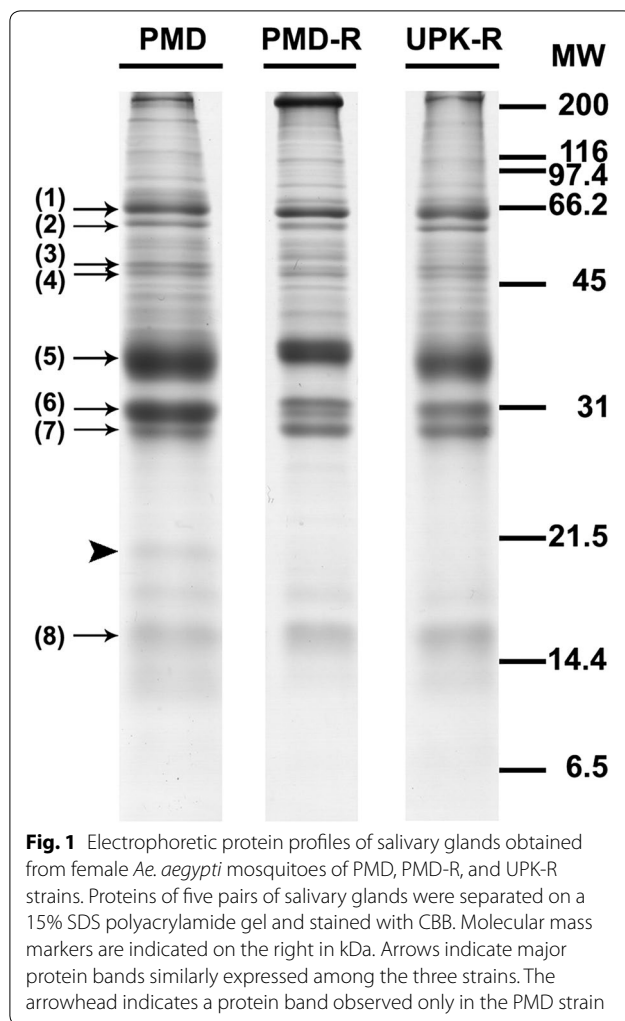


Fig. 1 Electrophoretic protein profiles of salivary glands obtained from female *Ae. aegypti* mosquitoes of PMD, PMD-R, and UPK-R strains. Proteins of five pairs of salivary glands were separated on a 15% SDS polyacrylamide gel and stained with CBB. Molecular mass markers are indicated on the right in kDa. Arrows indicate major protein bands similarly expressed among the three strains. The arrowhead indicates a protein band observed only in the PMD strain

(Fig. 2). Two-dimensional gel electrophoresis (2-DE) was performed on three biological replicates and images (Additional file 1: Figure S1) were analyzed using Image MasterTM Platinum 7.0 software (GE Healthcare, Buckinghamshire, UK). The molecular weight (MW), isoelectric point (pI) and normalized volume of each protein spot from each independent gel image were calculated following instructions for the software supplied by the manufacturer. Then, the average normalized volume (ANV) and standard deviation (SD) of the same protein (same MW and pI) from the three replicated 2-DE gel images were also calculated by the software. In this study, 57, 45 and 52 protein spots were detected in the PMD, PMD-R and UPK-R strains, respectively. Protein spots with ANV ≥ 0.1 were selected as major proteins. Results showed that at least 24 major protein spots were detected in each strain. The molecular weight of these proteins varied between 15 and 75 kDa, with pI ranging between 3.5 and 9.5. It was noted that the A1 spot was

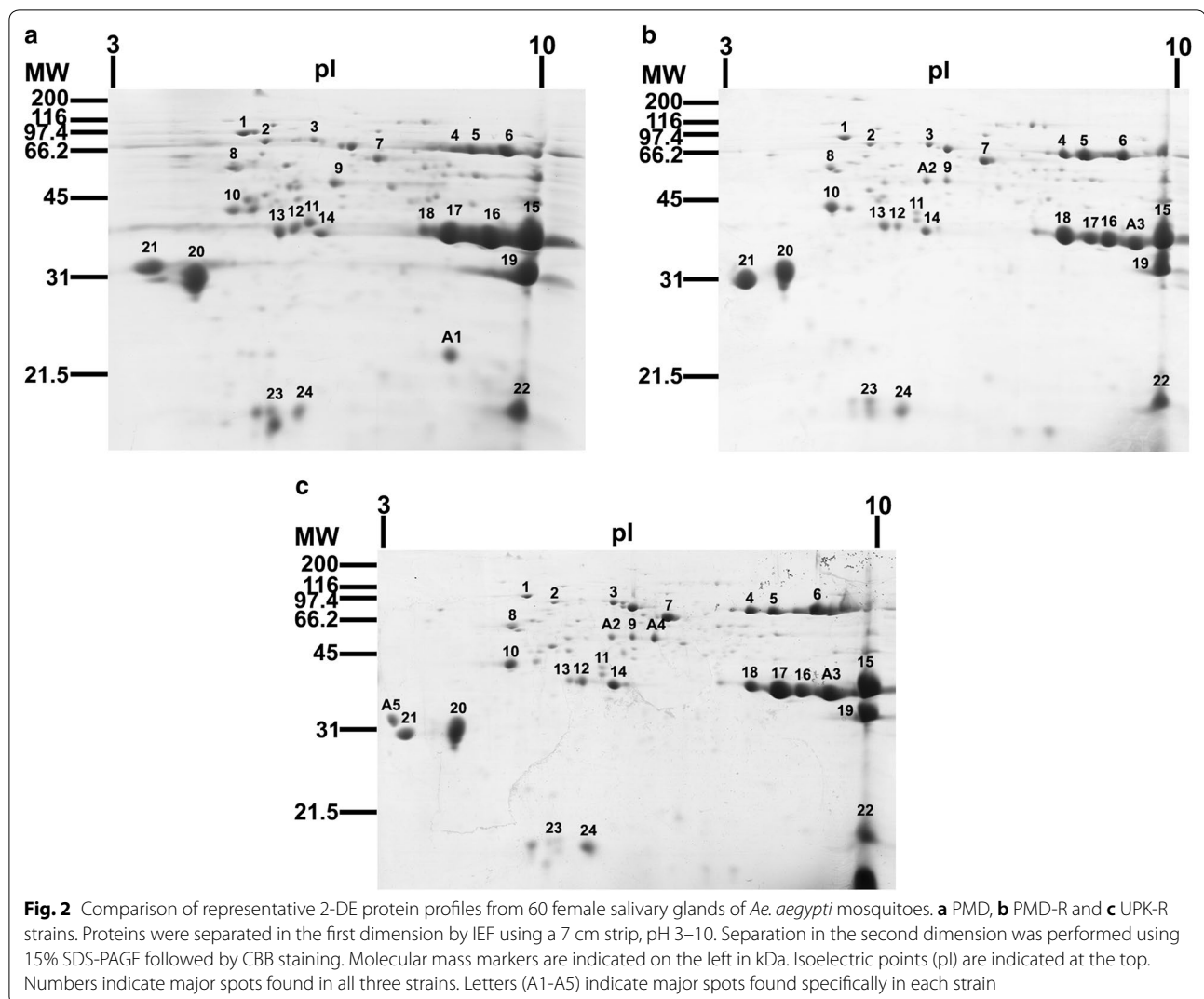
specific to the PMD strain (Fig. 2a) whereas the A2 and A3 spots were found in both the PMD-R (Fig. 2b) and UPK-R (Fig. 2c) strains. The A4 and A5 spots were found only in the UPK-R strain.

Identification of female salivary gland proteins of *Ae. aegypti* PMD, PMD-R and UPK-R strains by NanoLC-MS analysis

In order to identify the detected salivary gland proteins, a total of 154 protein spots comprised of 57, 45 and 52 protein spots from the PMD, PMD-R and UPK-R strains, respectively, were subjected to NanoLC-MS analysis. For the major protein spots, the relevant spot from two independent gels was used for NanoLC-MS analysis to confirm the identification. The results reported here refer to the identification by Mascot against the non-redundant NCBI nr_Metazoa database. Of these, 24 major protein spots were identified as similar proteins in all strains and only the best hit of the proteins from *Ae. aegypti* is reported in Table 2. The mass list (list of peptides) found for each major spot analyzed on the three mosquito strains is shown in Additional file 2: Table S1. These proteins included AAEL017349-PA, which was spot number 1 (SN1), heat-shock cognate 70 (SN2), putative secreted protein (SN3), apyrase (SN4 and 5), AAEL006333-PA/5'-nucleotidase (SN6), AAEL005672-PA/putative adenosine deaminase (SN7), AAEL000641-PA (SN8), salivary anti-FXa serpin (SN9), putative purine hydrolase (SN10), angiopoietin-like protein (SN11), putative 34 kDa secreted protein (SN12 and 13), putative 34 kDa family secreted salivary protein (SN14), D7 protein family (SN15, 16, 17 and 18), AAEL000793-PA/putative secreted protein (SN19), AAEL010228-PA/putative 19.6 kDa secreted protein (SN20), 30 kDa salivary gland allergen variant 3 (SN21), short form D7Cclu23 salivary protein (SN22), putative C-type lectin/AAEL000533-PA (SN23) and short salivary D7/putative 16.9 kDa secreted protein (SN24). For the differentially expressed protein spots, the A1, A2 and A4 spots were identified as a short D7 protein, a salivary anti-FXa serpin and a salivary serpin, respectively (Table 2). Unfortunately, the A3 and A5 spots could not be identified in our study.

Differential protein expression in salivary glands between susceptible and resistant strains

Quantification of the ANV of each major protein of the PMD, PMD-R and UPK-R strains was carried out. The fold expression values of the major proteins were calculated and compared between PMD and PMD-R (Fig. 3 and Additional file 3: Table S2), PMD and UPK-R (Fig. 4 and Additional file 4: Table S3), and PMD-R and UPK-R (Fig. 5 and Additional file 5: Table S4). For PMD-R compared to PMD, three downregulated proteins were identified, namely



salivary anti-FXa serpin (SN9), angiopoietin-like protein (SN11) and short form D7Cclu23 salivary protein (SN22) (Fig. 3). For UPK-R compared to PMD, six downregulated proteins were identified, namely salivary anti-FXa serpin (SN9), angiopoietin-like protein (SN11), putative 34 kDa secreted protein (SN13), 30 kDa salivary gland allergen variant 3 (SN21), short form D7Cclu23 salivary protein (SN22) and putative C-type lectin (SN23) (Fig. 4). For PMD-R compared to PMD, four downregulated proteins were identified, namely putative 34 kDa secreted protein (SN13), D7 protein (SN18), 30 kDa salivary gland allergen variant 3 (SN21), and putative C-type lectin (SN23) (Fig. 5).

Network analysis of identified salivary gland proteins of *Ae. aegypti* PMD, PMD-R and UPK-R strains by STITCH database 5.0

The STITCH database 5.0 was used to analyze potential functional associations of the identified *Ae. aegypti*

salivary gland proteins with other proteins, chemicals and insecticides. In this study, the STITCH network of identified protein-insecticide chemical interaction was presented as a network view. Based on this analysis, the interaction of the 24 major proteins of the three strains was similar. The network view showed that eight major proteins, AAEL017349-PA (SN1), apyrase (SN4 and 5), AAEL00633/5'-nucleotidase (SN6), AAEL005672-PA/putative adenosine deaminase (SN7), AAEL000641-PA (SN8), salivary anti-FXa serpin (SN9), putative purine hydrolase (SN10) and putative C-type lectin/AAEL000533-PA (SN23) were found to possibly interact with other proteins (Fig. 6). The confidence scores of interaction between the eight major proteins and other proteins and chemicals are shown in Additional file 6: Table S5.

Although A2 and A4 proteins had different accession numbers in the NCBI protein database, they had

Table 2 A list of identified female salivary gland proteins of *Ae. aegypti* PMD, PMD-R, and UPK-R strains by NanoLC-MS analysis

SN ^a (MW/pI)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pI	Peptide sequence(s)	Gene name ^f
1 (75/53)	PMD	XP_011493320	AAEL017349-PA	65	36	2/3	72.4/5.1	RLTPEDIERM	AAEL017349-PA
					29			KVADDDKATMEEAIDEK.I + Oxidation (M)	
	PMD-R	XP_011493320	AAEL017349-PA	65	36	2/3	72.4/5.1	RLTPEDIERM	AAEL017349-PA
2 (73/56)	UPK-R	XP_011493320	AAEL017349-PA	271	66	4/7	72.4/5.1	KVADDDKATMEEAIDEK.I + Oxidation (M)	
					29			KIVITNDQNR.L	AAEL017349-PA
					71			KFEELNMDLFRS + Oxidation (M)	
3 (72/62)	PMD	ABF18332	Heat-shock cognate 70	40	40	1/1	71.4/5.3	KDVDEVLVGGSTR.I	
	PMD-R	ABF18332	Heat-shock cognate 70	49	49	1/1	71.4/5.3	KVFAPEEISAMV/LGKM + Oxidation (M)	na ^g
	UPK-R	ABF18332	Heat-shock cognate 70	163	80	2/3	71.4/5.3	KVEIANDQGNR.T	na
					83			KVEIANDQGNR.T	na
					84			KDAGTISGLNLR.I	na
					84			RFEELNADLFRS	
4 (69/85)	PMD	AAL76020	Putative secreted protein	84	84	1/1	66.6/5.8	KTASNDCTPFR.L	AAEL000748
	PMD-R	AAL76020	Putative secreted protein	84	84	1/1	66.6/5.8	KTASNDCTPFR.L	AAEL000748
	UPK-R	AAL76020	Putative secreted protein	130	62	2/4	66.6/5.8	KDLEGITDELGKV	AAEL000748
5 (68/85)	PMD	AAC37218	Apyrase	231	65	3/6	63.2/8.5	KDIATLEQNVTLHK.D	
					82			RTGPLDSDVFKN	APY
					84			KVTLSNAVEAVRR	
	PMD-R	AAC37218	Apyrase	449	66	6/12	63.2/8.5	KVEAIGSTWGETK.I	APY
					61			RTGNCLQVSLR.I	
					76			KVTLSNAVEAVRR	
					73			KVEAIGSTWGETK.I	
					96			KKIEVMDYTNPKS + Oxidation (M)	
	UPK-R	AAC37218	Apyrase	231	80	3/5	63.2/8.5	KDKVEGYPYPTIVESKN	APY
					81			KIGIIGVLYDKT	
6 (67/85)					72			RIMDISKPIRS	
					78			KVTLSNAVEAVRR	

Table 2 (continued)

SN ^a (MW/pI)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pI	Peptide sequence(s)	Gene name ^f
5 (68/89)	PMD	AAC37218	Apyrase	137	59	2/4	63.2/8.5	R ⁺ TNCLQVSGLR.I	APY
					75			K ⁺ VEAIGSTVVGGETK.I	
				531	66	7/12	63.2/8.5	R ⁺ TGPLDSDVFK.N	
	PMD-R	AAC37218	Apyrase		71			K ⁺ IGIGLYDK.T	APY
					69			R ⁺ TNCLQVSGLR.I	
					87			K ⁺ VTLSNAVEAVRR	
					68			R ⁺ TGPLDSDVFK.N	
					79			K ⁺ VEAIGSTVVGGETK.I	
					93			K ⁺ DKVEGYPPTIVESK.N	
6 (68/95)	UPK-R	AAC37218	Apyrase	641	61	9/17	63.2/8.5	R ⁺ TGPLDSDVFK.N	APY
					81			K ⁺ IGIGLYDK.T	
					64			R ⁺ TNCLQVSGLR.I	
					70			R ⁺ VIDSKPIRS	
					79			K ⁺ VTLSNAVEAVRR	
					59			K ⁺ GLAPYLAELNKE	
					60			K ⁺ VEAIGSTVVGGETK.I	
					80			K ⁺ KIEVMDYTNPKS + Oxidation (M)	
					63			K ⁺ DKVEGYPPTIVESK.N	
				139	80	2/3	63.9/8.7	R ⁺ ISLAAGAITRG	
					61			K ⁺ LELSNDCRL	
				139	80	2/3	63.9/8.7	R ⁺ ISLAAGAITRG	
					61			K ⁺ LELSNDCRL	
				96	20	3/6	61.8/8.9	R ⁺ VTVLGSTITK	
					38			K ⁺ VGESKLELSNDCRL	
7 (66/72)	PMD	XP_001651237	AAEL005672-PA	593	39	11/23	59.9/6.4	R ⁺ GQILTALPFNSNANR.V	AAEL005672
					37			R ⁺ LKLEEMVK.G + Oxidation (M)	
					48			R ⁺ HIFEVLDR.F	
					45			R ⁺ FATDDEFLK.I	
					35			K ⁺ SNHPEFIGAKF	
					51			K ⁺ EWSLVSEIRQ	
					51			R ⁺ SLLEFAPALLK.L	
					52			K ⁺ GDFGVSHGPOFKC	
					70			R ⁺ GVLPDVYDLDGK.I	
					87			K ⁺ TDQNLDIAVLGSKR	
					62			K ⁺ QFYDDHVQYLEFRG	
					54			K ⁺ QLALNSIEYSAMNSEK.T + Oxidation (M)	

Table 2 (continued)

SN ^a (MW/pl)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pl	Peptide sequence(s)	Gene name ^f
8 (62/5.1)	PMD-R	AAL76033	Putative adenosine deaminase	100	22	3/4	60.6/6.5	KFYAPGRF	AAEL005672
					36			KSSHPEFIGAK.F	
					45			RFATDDEFKJ	
	UPK-R	AAL76033	Putative adenosine deaminase	219	70	3/7	60.6/6.5	RGVLPDVYDLGKI	
9 (59/66)					87			KTDQNLDVILGSKR	AAEL005672
					62			KQFYDDHVQYLEFRG	
	PMD	XP_001649774	AAEL000641-PA	163	77	2/4	56.3/5.0	KMDATANELEHTKI + Oxidation (M)	
					86			KVDATEEQELSEKH	
	PMD-R	XP_001649774	AAEL000641-PA	47	47	1/2	56.3/5.0	RNGTPIETGGRE	
	UPK-R	XP_001649774	AAEL000641-PA	82	82	1/2	56.3/5.0	KVDATEEQELSEKH	
	PMD	ABF18028	Salivary anti-FXa serpin	121	59	2/5	48.3/6.1	RMQVINDALKN + Oxidation (M)	
					62			KTPPEAAMGLEDKQ + Oxidation (M)	
10 (49/5.1)	PMD-R	ABF18028	Salivary anti-FXa serpin	58	58	1/2	48.3/6.1	R.TAMAVAIGSEK.V + Oxidation (M)	SRPN23
					59	4/11	48.3/6.1	R.TAMAVAIGSEK.V + Oxidation (M)	
	UPK-R	ABF18028	Salivary anti-FXa serpin	307	77			RMQVINDALKN + Oxidation (M)	
					106			RIPQFGLQTTVPGRQ	
11 (48/6.0)					64			KVFEQGQDVALGEIVQKM	AAEL006485
	PMD	AAL76010	Putative purine hydrolase	48	48	1/2	38.2/5.0	RQMVEDLNR- + Oxidation (M)	
	PMD-R	AAL76010	Putative purine hydrolase	76	42	2/4	38.2/5.0	RLITPAPR.R	
					34			RQMVEDLNR- + Oxidation (M)	
	UPK-R	AAL76010	Putative purine hydrolase	73	73	1/3	38.2/5.0	KSEIAGIYILGNNRN	
	PMD	ABF18025	Angiopietin-like protein variant, partial	68	22	2/6	33.5/5.7	KFWGPEEQR.Y	
					45			KFSDVSNTPKL.L	
	PMD-R	AAL76032	Angiopietin-like protein	68	22	2/6	33.5/5.7	KFWGPEEQR.Y	
12 (45/5.8)					45			KFSDVSNTPKL.L	AAEL000749
	UPK-R	ABF18025	Angiopietin-like protein variant, partial	153	37	4/15	33.5/5.7	KFWGPEEQR.Y	
					32			KFSDVSNTPKL.L	
					35			KSSGLTIPIGSEPR.V	
					49			RFTQMFSSQFYRH + Oxidation (M)	
	PMD	ABF18018	Putative 34 kDa secreted protein	64	64	1/3	36.4/5.2	KLQEEIEEQTK.Q	
	PMD-R	ABF18018	Putative 34 kDa secreted protein	64	64	1/3	36.4/5.2	KLQEEIEEQTK.Q	
	UPK-R	ABF18018	Putative 34 kDa secreted protein	132	65	2/6	36.4/5.2	KSVATEIVQMRD + Oxidation (M)	
					68			KQNFEDQVNVK.S	AAEL003601

Table 2 (continued)

SN ^a (MW/pl)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pl	Peptide sequence(s)	Gene name ^f
13 (46/5.7)	PMD	AAL76018	Putative 34 kDa secreted protein	54	54	1/4	36.4/5.2	RDSAISTDQVDQLKQ	AAEL003601
	PMD-R	AAL76018	Putative 34 kDa secreted protein	155	74	2/7	36.4/5.2	RDSAISTDQVDQLKQ	AAEL003601
	UPK-R	AAL76018	Putative 34 kDa secreted protein	176	40	4/12	36.4/5.2	KQNFEDQVNVK.S R.IYLGALRF K.SVATEIVQMRD + Oxidation (M) K.VTELEQQIAKQ K.QNFEDQVNVK.S K.LQEEIEETK.Q K.QCNLSEDDLT.L	AAEL003601
14 (43/6.2)	PMD	ABF18017	Putative 34 kDa family secreted salivary protein	98	49	2/6	36.0/6.1	K.QCNLSEDDLT.L	AAEL003600
	PMD-R	ABF18017	Putative 34 kDa family secreted salivary protein	206	73	3/12	36.0/6.1	K.QCNLSEDDLT.L K.TQGVSNMEVQLLR.E + Oxidation (M) R.QANQDTSKAEQEMVEK.I + Oxidation (M)	AAEL003600
	UPK-R	ABF18017	Putative 34 kDa family secreted salivary protein	256	66	3/8	36.0/6.1	K.VLNTILDQVNVK.L R.KVLNTILDQVNVK.L K.TQGVSNMEVQLLR.E	AAEL003600
15 (40/9.5)	PMD	AAL16049	Long form D7Bclu1 salivary protein	450	64	6/18	39.2/9.4	K.NELDVDEIARD R.SKDYFAALTGK.L K.SSSCSEVFEAFK.K R.KQVDDIDKIQCS.- K.SSSCSEVFEAFK.KV K.LEPSDDQATQCYTKC K.IGFYEPGEKRF R.SKDYFAALTGK.L K.YQMGSQGVFER.H + Oxidation (M) K.SSSCSEVFEAFK.K R.KQVDDIDKIQCS.- K.VTNDKFAFDYRE K.LEPSDDQATQCYTKC R.YMTSKNELDVDEIARD + Oxidation (M)	AAEL006417
	PMD-R	XP_001657778	AAEL006417-PA	614	58	8/29	39.2/9.4	K.IGFYEPGEKRF R.SKDYFAALTGK.L K.YQMGSQGVFER.H + Oxidation (M) K.SSSCSEVFEAFK.K R.KQVDDIDKIQCS.- K.VTNDKFAFDYRE K.LEPSDDQATQCYTKC R.YMTSKNELDVDEIARD + Oxidation (M)	AAEL006417
	UPK-R	AAL16049	Long form D7Bclu1 salivary protein	385	63	5/14	39.2/9.4	K.AALQNWLGWKL R.SKDYFAALTGK.L K.SSSCSEVFEAFK.K K.SSSCSEVFEAFK.KV K.LEPSDDQATQCYTKC	AAEL006417

Table 2 (continued))

SN ^a (MW/pl)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pl	Peptide sequence(s)	Gene name ^f
16 (39/9.5)	PMD	AAA29347	D7 protein	437	72	6/21	37.6/8.4	KSKVEAYANAVKQ	D7
					71			KFDASVIOEQFK A	
					58			RQYTVLDDALFKE	
					75			KDQLDVEEVKRD	
					85			KCLVESSVKDDFKE	
	PMD-R	AAA29347	D7 protein	363	83	5/17	37.6/8.4	RCMEDNILEDGANRL + Oxidation (M)	D7
					66			KSKVEAYANAVKQ	
					60			KDDFKEAFDYRE	
					69			KFDASVIOEQFK A	
					72			KQSYFFECENK.Y	
	UPK-R	AAA29347	D7 protein	390	102	6/21	37.6/8.4	KQLPSTNNDCAAVFKA	D7
					61			RTGLYDPVAQKF	
					68			KSKVEAYANAVKQ	
					71			KFDASVIOEQFK A	
					67			RQYTVLDDALFKE	
	PMD	AAA29348	D7 protein	153	63	3/9	37.5/8.5	KDQLDVEEVKRD	D7
					70			KQLPSTNNDCAAVFKA	
					55			RTGLYDPVAQKF	
					75			KINNELDAEEVKRD	
					23			KIKSYFFECENK.Y	
	PMD-R	AAA29348	D7 protein	191	65	3/10	37.5/8.5	KFDASVIOEQFK A	D7
					71			RQYTVLDDALFKE	
					59			KINNELDAEEVKRD	
					67			RQYTVLDDALFKE	
					84			KDQLDVEEVKRD	
	UPK-R	AAA29347	D7 protein	223	76	3/10	37.6/8.4	KCLVESSVKDDFKE	D7
					75			KINNELDAEEVKRD	
					63			KDDFKEAFDYRE	
					73			KFDASVIOEQFK A	
					74			KIKSYFFECENK.Y	
18 (42/8.9)	PMD	AAA29348	D7 protein	75	75	1/3	37.5/8.5	KINNELDAEEVKRD	D7
					208			KDDFKEAFDYRE	
					74			KFDASVIOEQFK A	
					74			KIKSYFFECENK.Y	
					74			KFDASVIOEQFK A	
	PMD-R	AAA29348	D7 protein	211	67	3/9	37.5/8.5	RQYTVLDDALFKE	D7
					74			KIKSYFFECENK.Y	

Table 2 (continued)

SN ^a (MW/pl)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pl	Peptide sequence(s)	Gene name ^f
19 (33/9.5)	PMD	XP_001651000	AAEL000793-PA	199	87	2/16	29.6/9.1	RMP TLVWDELAHIA SFNARK + Oxidation (M)	AAEL000793
					112			RGRPHVACNAPTQFGSACGQEPK.F	
	PMD-R	AAL76012	Putative secreted protein	199	87	2/16	29.6/9.1	RMP TLVWDELAHIA SFNARK + Oxidation (M)	AAEL000793
					112			RGRPHVACNAPTQFGSACGQEPK.F	
20 (33/4.6)	UPK-R	AAL76009	Putative secreted protein	38	38	1/3	30.2/9.0	KDYCSSVFRK	AAEL003053
	PMD	AAX54871	Putative 19.6 kDa secreted protein	31	31	1/4	21.8/4.8	KVIEQLDQIK.V	AAEL010228
					62	2/12	21.8/4.8	KDSNAYQCSQDR.S	AAEL010228
					104			RNPV DVG SAGDFS.KJ	
21 (31/4.1)	UPK-R	XP_001660690	AAEL010228-PA	173	66	2/12	21.8/4.8	KDSNAYQCSQDR.S	AAEL010228
					107			RNPV DVG SAGDFS.KJ	
	PMD	ABF18125	30 kDa salivary gland allergen variant 3	124	64	2/6	27.9/4.2	RVPVWEAIGRI	AAEL010235
					61			KNDPADTYR.Q	
	PMD-R	ABF18125	30 kDa salivary gland allergen variant 3	452	64	6/18	27.9/4.2	RVPVWEAIGRI	AAEL010235
					61			RQWALLDKDTK.V	
					86			RSALNNDLQSEVR.V	
					75			KSEYQCSSEDSFAAAK.S	
					80			KKSEYQCSSEDSFAAAK.S	
					88			KKSEYQCSSEDSFAAAK.S	
	UPK-R	ABF18125	30 kDa salivary gland allergen variant 3	301	61	4/17	27.9/4.2	KNDPADTYR.Q	AAEL010235
					62			RQWALLDKDTK.V	
22 (15/9.5)	PMD	AAL16050	Short form D7Cclu23 salivary protein	32	32	1/6	18.2/9.0	RSEYQCSSEDSFAAAK.S	AAEL006423
					71			RFFTEKPFCK.-	
	PMD-R	AAL16050	Short form D7Cclu23 salivary protein	163	47	4/22	18.2/9.1	KINGWAYFRC	AAEL006423
					46			RFFTEKPFCK.-	
23 (15/5.6)	UPK-R	AAL16050	Short form D7Cclu23 salivary protein	32	32	1/6	18.2/9.1	RINFLCHTANFKL	AAEL006423
					16			KTEMGDKINGWAYFRC + Oxidation (M)	
					32			RFFTEKPFCK.-	
					33	1/6	18.1/5.3	KQQQVEELAQ.RV	CTL16
	PMD	AAL76029	Putative C-type lectin	33	33	1/6	18.1/5.3	RLAVLDSEKQ.Q	CTL16
	PMD-R	XP_001647974	AAEL000533-PA	87	46	2/12	18.1/5.3	KQQQVEELAQ.RV	
					40			RLAVLDSEKQ.Q	
	UPK-R	AAL76029	Putative C-type lectin	93	51	2/12	18.1/5.3	KQQQVEELAQ.RV	CTL16
					42			KQQQVEELAQ.RV	

Table 2 (continued)

SN ^a (MW/pI)	Strain	Accession number ^b	Protein description	Prot. score ^c	PS ^d	No. P/ %C ^e	Database MW/pI	Peptide sequence(s)	Gene name ^f
24 (15/5.9)	PMD	AAL76035	Putative 16.9 kDa secreted protein	135	23	3/20	17.4/5.5	K:TELDEVTKE	SG14.5sp
					32			R:NLQVDGNMPK.V + Oxidation (M)	
	PMD-R	ABF18031	Short salivary D7 protein, partial	80	80	1/8	17.5/5.7	K:VAQDMVPYGFNIK.T + Oxidation (M)	na
	UPK-R	ABF18031	Short salivary D7 protein, partial	75	75	1/8	17.5/5.7	K:VAQDMVPYGFNIK.T + Oxidation (M)	na
A1 (20/8.6)	PMD	ABF18082	Short D7 protein	44	44	1/5	18.6/8.2	R:QNQIEELNKL	SG16.2sp
A2 (56/6.2)	PMD-R	ABF18028	Salivary anti-FXa serpin	229	70	3/8	48.3/6.1	R:TAMAVAIGSEK.V + Oxidation (M)	SRPN23
					69			K:MEMSIAADGEKQ + Oxidation (M)	
	UPK-R	ABF18028	Salivary anti-FXa serpin	95	90	1/3	48.3/6.1	K:TPPEAAMGLEDKQ + Oxidation (M)	SRPN23
A4 (59/6.2)	UPK-R	ABF18509	Salivary serpin	235	62	3/8	48.2/6.3	R:IPQFGLQTTVPGRQ	SRPN23
					68			R:MGQVINDALK.N + Oxidation (M)	
					105			K:TPPEAAMGLEDKQ + Oxidation (M)	
								R:IPQFGLQTTVPGRQ	

^a Spot number refers to those shown in Fig. 2 (Observed MW/pI)^b Accession number of the best hit of proteins from *Ae. aegypti*, available at <http://www.ncbi.nlm.nih.gov>^c Prot. Score = Protein score, Mowse score ≥ 30 ^d PS = Peptide score^e No. P/%C = Number of peptide/% coverage^f Name of the gene that codes for the protein sequences, available at <http://www.uniprot.org/uploadlists>^g na, not available

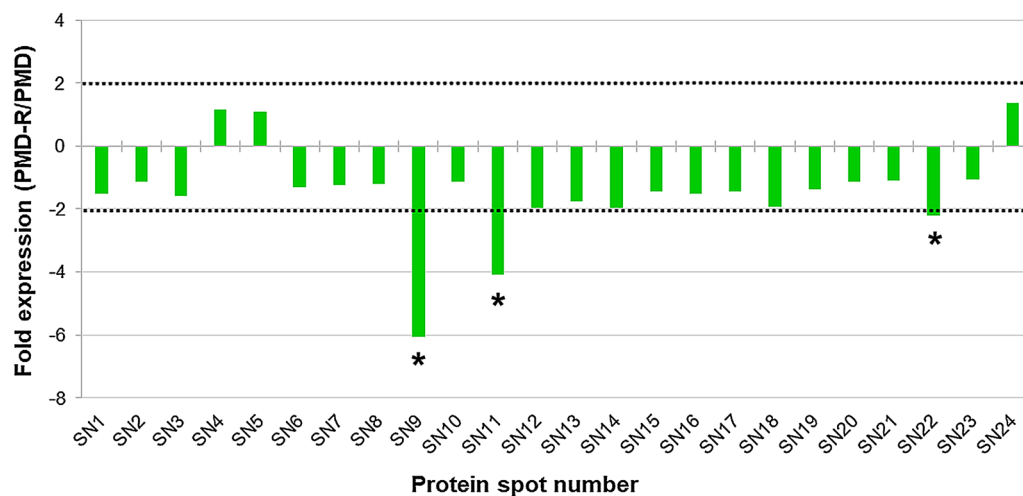


Fig. 3 Differential protein expression in salivary glands between PMD and PMD-R strains. Differences in protein expression are represented as expression ratio or fold expression (PMD-R/PMD). Horizontal dot lines indicate the 2-fold difference in expression level in either direction (2 for a higher expression in the PMD-R strain and -2 for a lower expression in the PMD-R strain). Proteins showing significantly different more than or less than 2-fold expression (Student's t-test, $P \leq 0.05$) are indicated by an asterisk (*). The ANV of each major protein and their fold expression values are shown in Additional file 3: Table S2

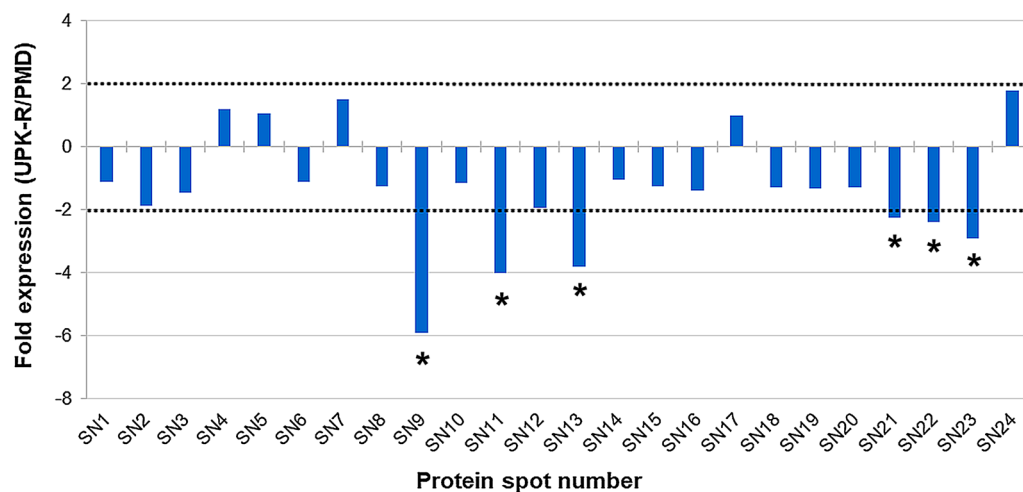


Fig. 4 Differential protein expression in salivary glands between PMD and UPK-R strains. Differences in protein expression are represented as expression ratio or fold expression (UPK-R/PMD). Horizontal dot lines indicate the 2-fold difference in expression level in either direction (2 for a higher expression in the UPK-R strain and -2 for a lower expression in the UPK-R strain). Proteins showing significantly different more than or less than 2-fold expression (Student's t-test, $P \leq 0.05$) are indicated by an asterisk (*). The ANV of each major protein and their fold expression values are shown in Additional file 4: Table S3

the same gene name (SRPN23) in UniProtKB. Therefore, both A2 and A4 are likely to be the same protein, SRPN23. The network view of the SRPN23 showed that the protein was found interact with sodium and calcium ions (Fig. 7). The confidence scores of interaction between the SRPN23 and sodium and calcium ions are shown in Additional file 7: Table S6.

Discussion

The present results revealed variation in protein expression profiles and abundance of specific proteins of *Ae. aegypti* salivary glands between insecticide-susceptible and resistant strains. In addition, we were able to identify the specific protein involved. Proteins identified from *Ae. aegypti* salivary glands in this study have been shown to

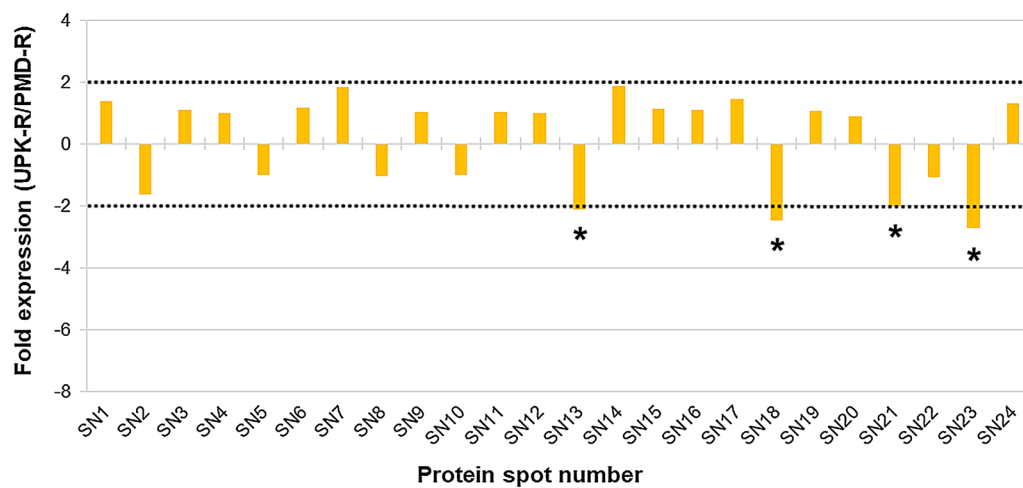


Fig. 5 Differential protein expression in salivary glands between PMD-R and UPK-R strains. Differences in protein expression are represented as expression ratio or fold expression (UPK-R/PMD-R). Horizontal dot lines indicate the 2-fold difference in expression level in either direction (2 for a higher expression in the UPK-R strain and -2 for a lower expression in the UPK-R strain). Proteins showing significantly different more than or less than 2-fold expression (Student's t-test, $P \leq 0.05$) are indicated by an asterisk (*). The ANV of each major protein and their fold expression values are shown in Additional file 5: Table S4

be associated with the facilitation of blood-feeding [21–26]. These proteins include apyrase/5'-nucleotidase (SN4, 5 and 6), putative adenosine deaminase (SN7), salivary anti-FXa serpin (SN9), D7 protein family (SN15, 16, 17, 18, 22 and 24) and 30 kDa salivary gland allergen variant 3 (SN21). Apyrase hydrolyzes ATP and ADP to adenosine by inhibiting ADP-dependent platelet aggregation [23]. Ribeiro et al. [27] proposed that adenosine deaminase activity may help blood-feeding in *Ae. aegypti* by producing inosine and by removing adenosine. Salivary anti-FXa serpin has anti-coagulant properties that inhibits coagulation factor Xa [25]. D7 protein family in *Ae. aegypti* mosquitoes exists in two forms, the long form (~37 kDa) and the short form (~16–18 kDa). The long form D7 salivary proteins function as scavengers of biogenic amines [25]. The short form D7 salivary proteins bind to biogenic amines such as epinephrine, histamine and serotonin, thereby inhibiting platelet aggregation, vasoconstriction and inflammation [22]. Aegyptin is a platelet inhibitor identified in *Ae. aegypti*. It binds to collagens, thereby preventing collagen interaction with Von Willebrand factor, integrin $\alpha 2\beta 1$ and glycoprotein VI [26]. A protein involved in heat-shock response, heat-shock cognate 70 (SN2), was also identified. Zhao et al. [28] demonstrated that expression level of heat-shock cognate 70 mRNA of female *Ae. aegypti* increases (five- to six-fold) after 42 °C treatment for one hour. Studies by Wasinpiyamongkol et al. [29] and Oktarianti et al. [30] demonstrated that heat-shock cognate 70 (SN2), apyrase/5'-nucleotidase (SN4, 5 and 6), putative adenosine deaminase (SN7), salivary anti-FXa serpin (SN9), angiopoietin-like protein

variant, partial (SN11), putative 34 kDa family secreted salivary protein (SN14) and D7 protein family (SN15, 16, 17 and 18) are immunogens.

Differentially expressed protein profiles of the salivary gland proteins of *Ae. aegypti* PMD, PMD-R and UPK-R strains were examined based on the annotated spots in the 2-DE gels and the fold expression values. The results showed that five additional major protein spots (A1–A5) were differentially expressed in the *Ae. aegypti* sialome between the susceptible and resistant strains. A short D7 protein (A1) was specific to the PMD strain whereas a salivary anti-FXa serpin (A2) and A3 were only detected in both the PMD-R and UPK-R strains. A salivary serpin (A4) and A5 were found only in the UPK-R strain. As the A3 and A5 spots were found only in the resistant strains but could not be identified, their DNA and amino acid sequence information is required for further study on their function.

Based on the fold expression values of the major salivary gland proteins, three downregulated proteins (> 2 fold change in expression) were identified in the PMD-R, compared to the PMD including salivary anti-FXa serpin (SN9), angiopoietin-like protein (SN11) and short form D7Cclu23 salivary protein (SN22). Comparison of differentially expressed protein profiles between the PMD and UPK-R strains showed six downregulated proteins in UPK-R compared to PMD including salivary anti-FXa serpin (SN9), angiopoietin-like protein (SN11), putative 34 kDa secreted protein (SN13), 30 kDa salivary gland allergen variant 3 (SN21), short form D7Cclu23 salivary protein (SN22) and putative C-type lectin (SN23).

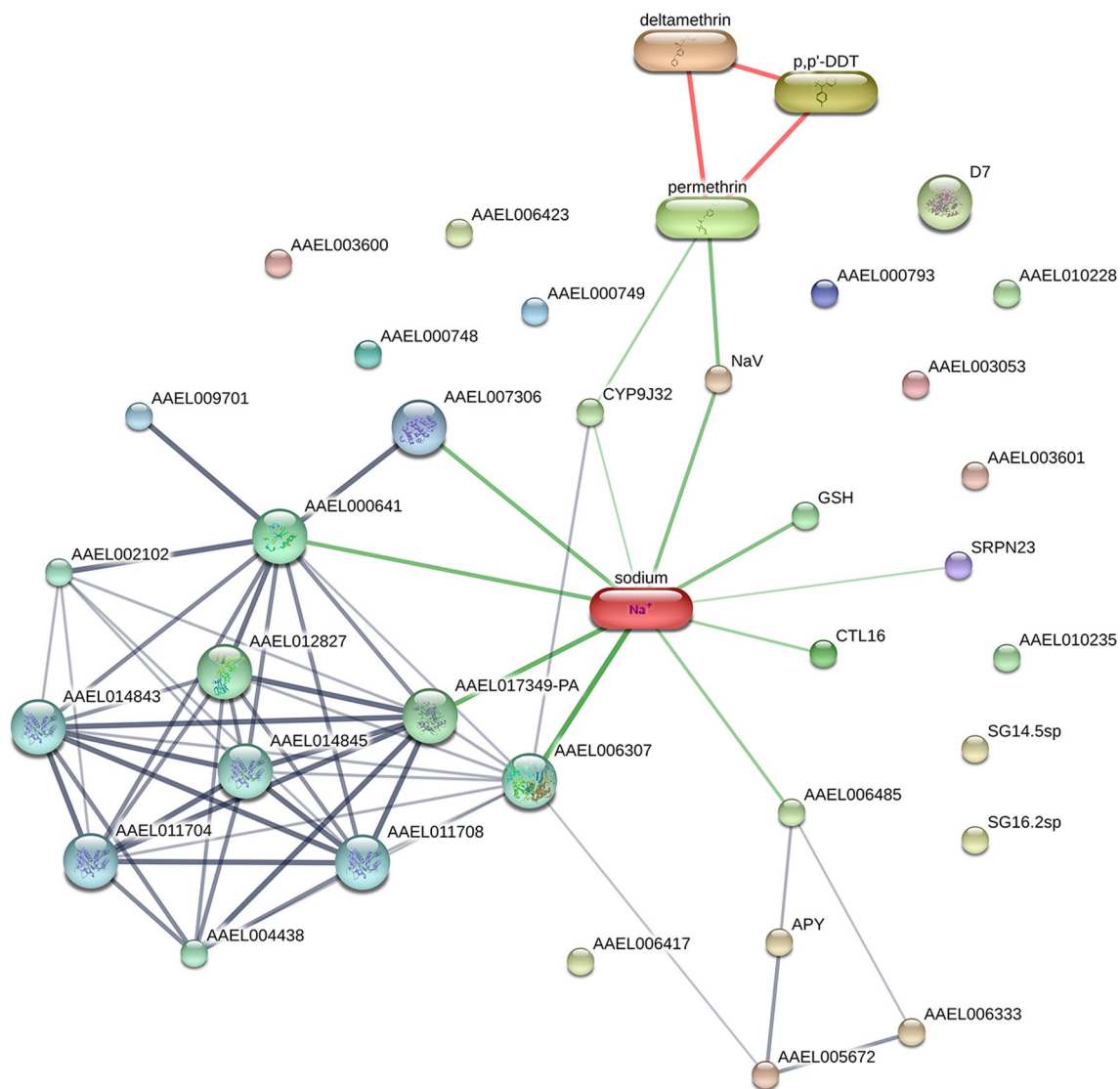


Fig. 6 Representative functional associations of salivary gland proteins of adult female *Ae. aegypti* between PMD, PMD-R, and UPK-R strains with other proteins, chemicals and insecticide using the STITCH database 5.0. The predicted functional interaction networks are shown in the network view where the stronger associations are represented by thicker lines. Protein-protein interactions are shown in grey, chemical-protein interactions in green and interactions between chemicals in red. Gene names corresponding to the proteins are described in Table 2

Comparison between the PMD-R and UPK-R strains revealed four downregulated proteins in the UPK-R strain, compared to the PMD-R strain: putative 34 kDa secreted protein (SN13), D7 protein (SN18), 30 kDa salivary gland allergen variant 3 (SN21), and putative C-type lectin (SN23). The results indicated that the pyrethroid resistant strains expressed lower amounts of the differentially expressed major salivary gland proteins than the PMD strain. Furthermore, the UPK-R strain had fold expression values of the differentially expressed proteins that were lower than the PMD-R strain, suggesting that

pyrethroids might induce change or alteration of salivary gland proteins and/or their expression in the resistant mosquitoes. Since the change in expression of salivary proteins may impact on the overall fitness of the resistant mosquitoes, these results might help explain blood-feeding and other fitness problems in *Ae. aegypti* female populations with high insecticide resistance levels [31–33].

Djegbe et al. [17] demonstrated four salivary proteins differentially expressed in susceptible (SLAB) and resistant (SR) strains of *Cx. quinquefasciatus* including three metabolic enzymes (endoplasmic, triosephosphate

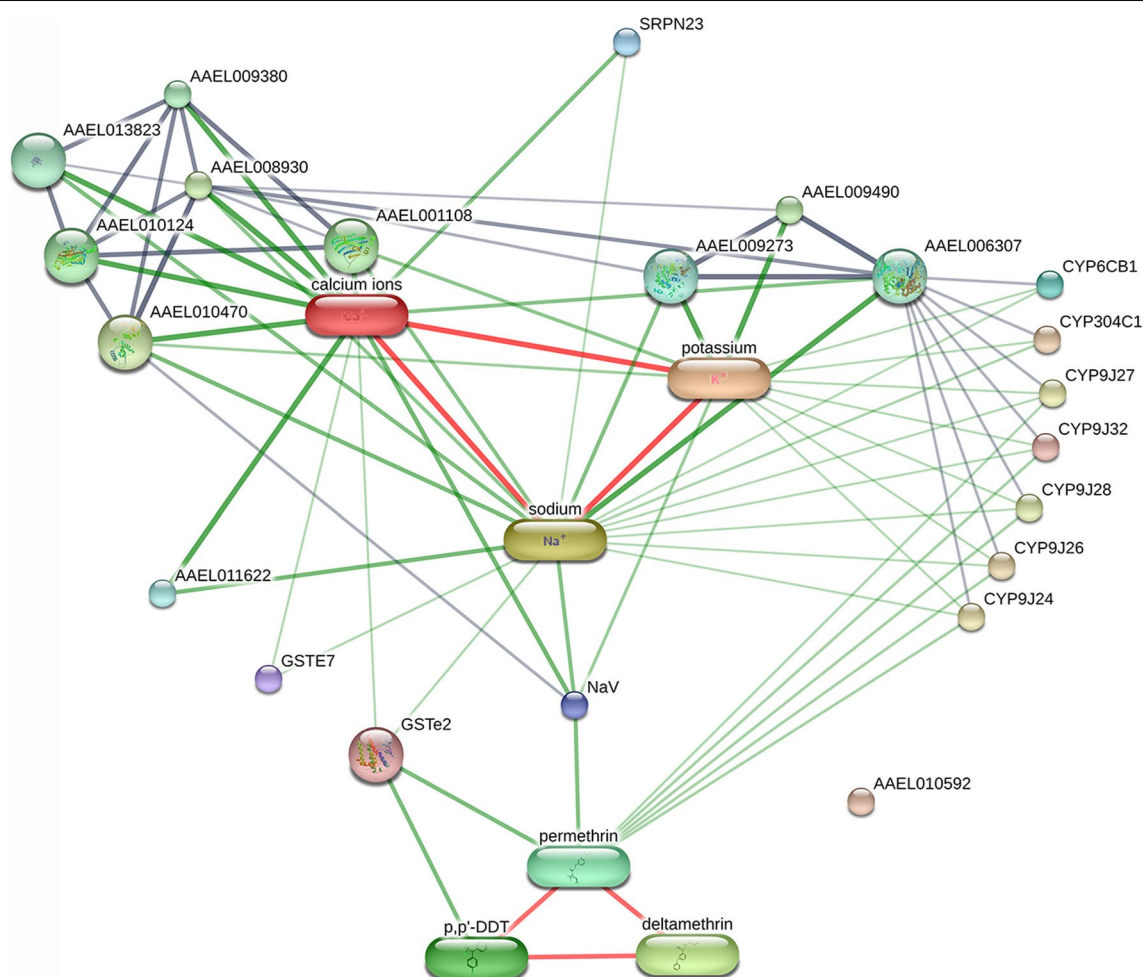


Fig. 7 Representative functional association of the SRPN23 with pyrethroid insecticide using the STITCH database 5.0. The predicted functional interaction networks are shown in the network view where the stronger associations are represented by thicker lines. Protein-protein interactions are shown in grey, chemical-protein interactions in green and interactions between chemicals in red. Gene names corresponding to the protein are described in Table 2

isomerase and heat-shock protein 83) and D7 long form. In susceptible and *ace-1^R* resistant strains of *An. gambiae* mosquitoes, five proteins regulated in the resistant AceRKis salivary gland extracts have been identified. Two of them, Saglin and TRIO have shown high differences between the susceptible and *ace-1^R* resistant strains. Both Saglin and TRIO are involved in protection against oxidation, blood-feeding process and pathogen invasion [18]. Vijay et al. [19] reported differentially expressed proteins and enzymes in salivary glands of deltamethrin/DDT resistant strains of *An. stephensi* that may have an impact on insecticide resistance and xenobiotic detoxification, such as short chain dehydrogenase reductase, cytochrome 450 and phosphodiesterase.

A search within UniProtKB (<http://www.uniprot.org/uploadlists/>) revealed that peptides produced from spots SN9, A2 and A4 all had identity to SRPN23. Given that

these spots were found to have variable pIs, it is likely that these spots represent post-translationally modified isoforms of SRPN23. Interestingly, two additional spots, A2 and A4 (SRPN23 isoforms), were expressed only in the PMD-R and UPK-R strains, suggesting that pyrethroids might induce alteration of serpin isoforms in the salivary glands of the resistant mosquitoes by an effect of post-translation modifications (PTM). Insecticide resistance could probably be associated with specific isoform of SRPN23 protein rather than changes in expression of the protein. The potential functional associations of the identified *Ae. aegypti* salivary gland proteins with other proteins, chemicals and insecticides were analyzed by STITCH database 5.0. The results showed that SRPN23 interacted with sodium and calcium ions. These results could warrant investigation of a role of SRPN23 in insecticide resistance because both pyrethroid resistant

strains had additional SRPN23 isoforms. Furthermore, testing for the effect of exposure to insecticides in the three strains of *Ae. aegypti* mosquitoes on blood-feeding behavior and metabolic mechanisms should be performed in the future.

Although salivary gland proteins of *Ae. aegypti* mosquito have been studied by several research groups, for example by Valenzuela et al. [21], Ribeiro et al. [22] and Wasinpiyamongkol et al. [29, 34], the status of insecticide resistance of the mosquitoes used in their studies, i.e. *Ae. aegypti* Liverpool/black eye or Bangkok strains, is unknown. In this study, results revealed that the salivary gland proteins of the PMD, PMD-R and UPK-R strains were differentially expressed. Therefore, comparative analyses of the expression of salivary gland proteins of different strains of *Ae. aegypti*, especially the Liverpool/black eye and Rockefeller, 1016 Ile/Ile kdr homozygous strains [31], should be carried out to investigate the expression of salivary proteins due to insecticide resistance.

In summary, to our knowledge this study reports for the first time the proteins expressed differentially in the salivary glands of insecticide-resistant *Ae. aegypti* mosquitoes between pyrethroid-susceptible (PMD) and resistant (PMD-R and UPK-R) strains. Salivary anti-FXa serpin, angiopoietin-like protein and short form D7Cclu23 salivary protein were downregulated in PMD-R when compared to PMD. Six downregulated proteins were detected in UPK-R when compared to PMD, including salivary anti-FXa serpin, angiopoietin-like protein, putative 34 kDa secreted protein, 30 kDa salivary gland allergen variant 3, short form D7Cclu23 salivary protein and putative C-type lectin. Four downregulated proteins were found in the UPK-R when compared to PMD-R, including putative 34 kDa secreted protein, D7 protein, 30 kDa salivary gland allergen variant 3, and putative C-type lectin. Network analysis by STITCH database 5.0 showed that the SRPN23 interacted with sodium and calcium ions.

Conclusions

This study provides new information on differentially expressed salivary gland proteins in pyrethroid-resistant *Ae. aegypti* mosquitoes. The findings emphasize the requirement for further studies regarding roles of these salivary proteins in viral infection, development and transmission in the resistant strains that might be useful for the development of control strategies for virus transmission by mosquitoes.

Methods

Mosquito strains and rearing

Colonies of *Ae. aegypti*, PMD (susceptible to pyrethroids but resistant to DDT), PMD-R (resistant to DDT and

permethrin) and UPK-R (resistant to DDT, permethrin and deltamethrin) strains were successfully maintained in an insectary of the Department of Parasitology, Faculty of Medicine, Chiang Mai University, Thailand and utilized in this study. *Ae. aegypti* PMD and PMD-R strains were established from field-caught mosquitoes from Ban Pang Mai Daeng, Mae Taeng District, Chiang Mai Province and maintained in an insectary at the Department of Parasitology, Faculty of Medicine, Chiang Mai University, Thailand since 1997 [5]. The UPK-R strain was established from wild-caught mosquitoes from Wat Upakhut, Chiang Mai city and maintained in an insectary at the Department of Parasitology, Faculty of Medicine, Chiang Mai University, Thailand since 2006 [14]. The permethrin resistance level of UPK-R and PMD-R strains, as determined by larval bioassays, is higher than the PMD strain by 325-fold and 25-fold, respectively ([10, 11], P. Somboon unpublished data). The deltamethrin resistance level of the UPK-R and PMD-R strains, as determined by larval bioassays, is higher than the susceptible PMD strain by 53-fold and 13-fold, respectively [14]. For each strain, a total of 300–400 eggs were placed into a 25 × 35 × 6 cm plastic tray filled with 3 l of distilled water and allowed to hatch. After hatching, the larvae were fed on finely ground dog food and the water was changed three times per week. Pupae were collected and transferred to plastic cups containing distilled water and then placed into a 30 × 30 × 30 cm mosquito cage. After emergence, the mosquitoes were maintained in the insectary at 27 ± 2 °C with a relative humidity of 70 ± 10% and a light-dark photocycle of 12:12 h. The mosquitoes were provided with 10% sucrose solution. They were allowed to feed on adult albino rats, *Rattus norvegicus*, to produce eggs. To avoid contamination of the mosquito strains, each strain was separately maintained in a different room of our insectary in the Department of Parasitology. Sugar-fed females aged five to seven days post-emergence were used in this study.

Insecticide susceptibility test

The mosquitoes from PMD-R and UPK-R colonies had been exposed regularly to the standard WHO permethrin (0.75%) and deltamethrin (0.05%) papers, respectively, to maintain their insecticide resistant status. Insecticide susceptibility tests were performed on all mosquito strains according to the protocol described by WHO [20]. Briefly, 25 one-day-old unfed female mosquitoes were exposed to insecticide impregnated paper in an exposure tube for 1 h and transferred to the holding tube. The mortality rate was counted after 24 h post-exposure. Control mosquitoes were exposed to paper without insecticide for 1 h and the mortality rate was counted after 24 h post-exposure. Two replicates of the control

group and four replicates of each test were performed each time. The mosquitoes that were able to fly and rest on the paper were recorded as survived or resistant. Mortality of 5–20% was accepted in the control groups and Abbott's formula was used to correct the mortality rate in the test groups [35]. If the mortality in the control group was over 20%, all tests were discarded.

Salivary gland dissection and protein quantification

The salivary glands of females of the three strains were dissected using the method described by Jariyapan et al. [36]. Mosquitoes of each strain were anaesthetized on ice before dissection. Salivary glands were dissected in phosphate-buffered saline (PBS), washed with PBS on a new slide to clean them from other contaminated tissues, and collected in a microcentrifuge tube at -80 °C until use. The protein content was determined using a Micro BCA Protein Assay Kit (Pierce, Rockford, IL, USA).

Sds-page

SDS-gel loading buffer (50 mM Tris-HCl, pH 6.8, 100 mM DTT, 2% SDS, 0.1% bromophenol blue, 10% glycerol), 1:2 (v/v), was added into each salivary gland sample of the three strains. The samples were heated for 5 min before loading on 15% SDS polyacrylamide gels. Molecular weight markers (Bio-Rad, Hercules, California, USA) were applied in each gel [37, 38].

Two-dimensional gel electrophoresis

Two-dimensional gel electrophoresis was performed using a 2D system from GE Healthcare (Buckinghamshire, UK) as described in our previous studies by Sor-suwan et al. [37] and Phattanawiboon et al. [38]. Briefly, 60 pairs of female salivary glands (≈ 80 μ g) were used as a sample for 2-DE analysis. 2-DE samples were collected from different cohorts of each mosquito strain. Samples from each strain were subjected to triplicate runs. A 2-D Clean-Up kit (GE Healthcare) was used to desalt the salivary gland samples. For the first dimension, each sample was diluted in a 125 μ l rehydration solution (8 M urea, 50 mM DTT, 0.2% 3/10 Bio-lyte Ampholyte, 4% CHAPS, 0.002% bromophenol blue). This solution was applied onto an IPG strip (7 cm, pI 3–10, GE Healthcare). The IPG strip was submitted to isoelectric focusing on an Ettan IPGphor III (GE Healthcare) which was operated as described in Phattanawiboon et al. [38]. After that, the focused IPG strip was incubated in 10 ml SDS equilibration buffer (6 M urea, 0.05 M Tris, pH 8.8, 2% SDS, 30% glycerol, 0.002% bromophenol blue) containing 100 mg DTT for 15 min and then in 5 ml SDS equilibration buffer containing 125 mg iodoacetamide for 15 min. The equilibrated strip was applied to the surface of vertical 15% SDS polyacrylamide gels, sealed with 0.1% agarose. The

Mini-PROTEAN Tetra Electrophoresis System (Bio-Rad) was used to separate proteins in the second dimension. Molecular weight markers (broad range from Bio-Rad) were loaded on each gel.

Coomassie Brilliant Blue (CBB) staining and gel image analysis

After electrophoresis, the gels were stained with CBB and spots in each gel and their expression volume were analyzed as described in our previous studies by Phattanawiboon et al. [38].

Statistical analysis

Image Master 2D Platinum 7.0 software (GE Healthcare) was used to measure the density of all protein spots and quantify the ANV for each protein spot. Statistical analysis (Student's t-test, $P \leq 0.05$) was performed using SPSS version 22.0 software (SPSS, Chicago, IL, USA) with a cut-off of 2-fold up- or downregulated to compare the ANV of each protein spot between two strains, i.e. PMD and PMD-R, PMD and UPK-R, and PMD-R and UPK-R strains.

In-gel digestion

Protein spots of interest were cut from the 2-DE gels using sterile surgical blades ensuring that technique was aseptic. Each spot sample was placed separately in a sterile microcentrifuge tube. The samples were subjected to in-gel digestion using methods described in our previous studies by Sor-suwan et al. [37] and Phattanawiboon et al. [38]. The gel plugs were dehydrated with 100% acetonitrile (ACN), reduced with 10 mM DTT in 10 mM ammonium bicarbonate at room temperature for 1 h, and alkylated at room temperature for 1 h in the dark in the presence of 100 mM iodoacetamide (IAA) in 10 mM ammonium bicarbonate. After alkylation, the gel pieces were dehydrated twice with 100% ACN for 5 min. The gels were digested in 10 μ l of trypsin solution (10 ng/ μ l trypsin in 50% ACN/10 mM ammonium bicarbonate at room temperature for 20 min, and then 20 μ l of 30% ACN) were added. After that the gel samples were incubated at 37 °C for a few hours or overnight. Products of digested proteins from the gels were extracted by adding 40 μ l of 50% ACN in 0.1% formic acid (FA) and shaking for 10 min at room temperature. Extracted peptides were collected in a new tube. The pool-extracted peptides were dried by a vacuum centrifuge and kept at -80 °C for further analysis.

NanoLC-MS analysis and protein identification

NanoLC-MS analysis of the pool-extracted peptides was performed as a method described in Phattanawiboon et al. [38]. The pool-extracted samples were injected

into an Ultimate 3000 LC System (Dionex, Sunnyvale, California, USA) coupled to an ESI-Ion Trap MS (HCT Ultra PTM Discovery System, Bruker, Germany) with electrospray at a flow rate of 300 nl/min to a nanocolumn (Acclaim PepMap 100 C18, 3 μ m, 100 A, 75 μ m id $\times$ 150 mm). A solvent gradient (solvent A: 0.1% formic acid in water; solvent B: 80% 0.1% formic acid in 80% acetonitrile) was run for 40 min. Mascot from Matrix Science Ltd. (London, UK) was used to search all of the tandem mass spectra [39]. The resulting sequence data was searched against the National Center for Biotechnology Information nonredundant (NCBI nr) protein database. The parameters for searching were as follows: enzyme of specificity strict trypsin; three missed cleavages; fixed modification (carbamidomethyl); variable modification (Methionine oxidation); peptide tolerance, variable from 50 ppm to 100 ppm; fragment mass tolerance of $\pm$ 0.5 Da; peptide change of 1+, 2+, and 3+; and monoisotopic. Proteins identified with a statistically significant Mowse score ($\geq$ 30) was reported. Gene ontology analysis was performed using UniProtKB (<http://www.uniprot.org/uploadlists/>) for molecular function, biological processes and cellular component. The mapping of protein-chemical interactions was analyzed according to STITCH 5.0 database (<http://stitch.embl.de/>) and KEGG (Kyoto Encyclopedia of Genes and Genomes) PATHWAY data.

Additional files

Additional file 1: Figure S1. Three independent biological replicated 2-DE gel images of each strain.

Additional file 2: Table S1. The mass list (list of peptides) found for each major spot analyzed on the three mosquito strains.

Additional file 3: Table S2. Differential protein expression in salivary glands between PMD and PMD-R strains.

Additional file 4: Table S3. Differential protein expression in salivary glands between PMD and UPK-R strains.

Additional file 5: Table S4. Differential protein expression in salivary glands between PMD-R and UPK-R strains.

Additional file 6: Table S5. The confidence scores of interaction between the eight major proteins and other proteins and chemicals.

Additional file 7: Table S6. The confidence scores of interaction between the SRPN23 and sodium and calcium ions.

Abbreviations

ACN: acetonitrile; ANV: average normalized volume; BSA: bovine serum albumin; CBB: Coomassie Brilliant Blue; CHAPS: 3-[(3-cholamidopropyl)dimethylammonio]-1-propanesulfonate; CHIKV: chikungunya virus; CRISPR-Cas9: clustered regularly interspaced short palindromic repeats-associated protein 9; DDT: dichlorodiphenyltrichloroethane; DENV: dengue virus; DHF: dengue hemorrhagic fever; DTT: dithiothreitol; FA: formic acid; IAA: iodoacetamide; IEF: isoelectric focusing; IPG: immobilized pH gradient; kDa: kilodalton; kdr: knockdown resistance; KEGG: Kyoto Encyclopedia of Genes and Genomes; MW: molecular weight; NanoLC-MS: nano liquid chromatography-mass

spectrometry; Na₂SO₄: sodium sulfate; NCBI nr: National Center for Biotechnology Information nonredundant; PBS: phosphate-buffered saline; pI: isoelectric points; PMD: Pang Mai Dang susceptible strain; PMD-R: Pang Mai Dang resistant strain; PTM: post-translation modifications; RNAi: RNA interference; SDS: sodium dodecyl sulfate; SDS-PAGE: sodium dodecyl sulphate polyacrylamide gel electrophoresis; SN: spot number; STITCH: Search Tool for Interacting Chemicals; UPK-R: Upakut resistant strain; WHO: World Health Organization; 2-DE: two-dimensional gel electrophoresis.

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Availability of data and materials

The data supporting the conclusions of this article are included within the article and its additional files.

Authors' contributions

NJ conceived and designed the experiments. CM, SS and SK performed the experiments. NJ, CM, SR, PT and PS analyzed the data. NJ wrote the paper. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The protocols were approved by the Animal Ethics Committee of the Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand (Approval Protocol Number: PAR-2558-03170, no. 05/2015).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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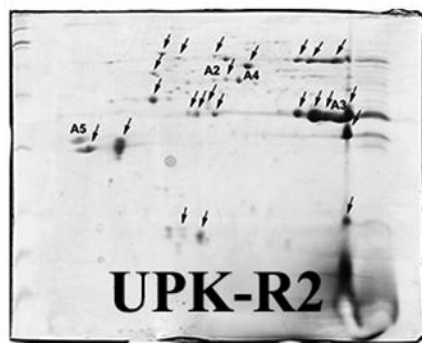
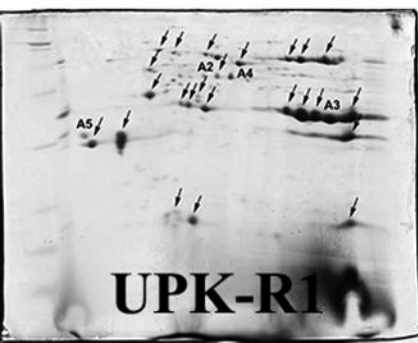
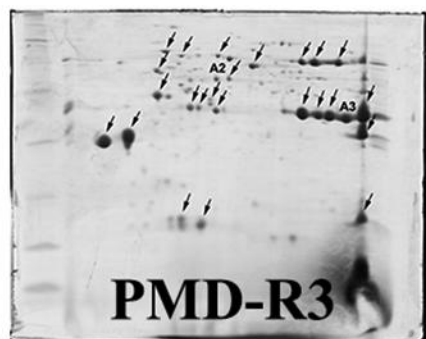
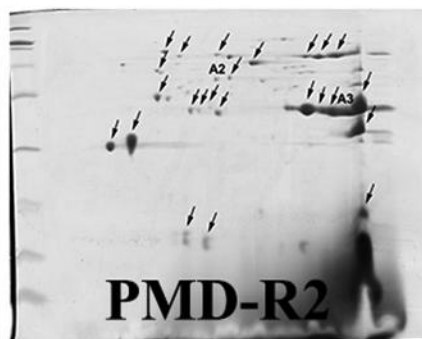
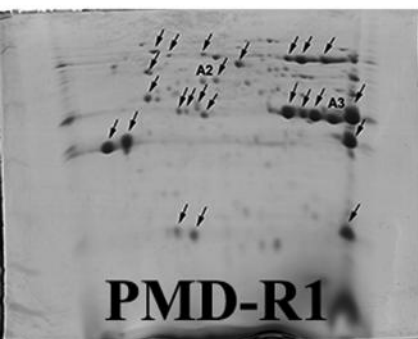
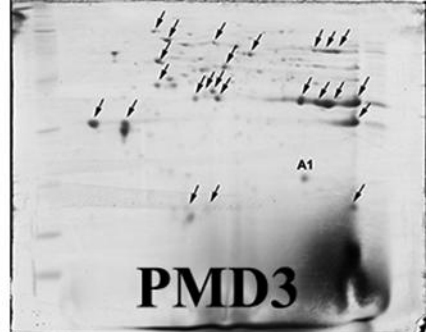
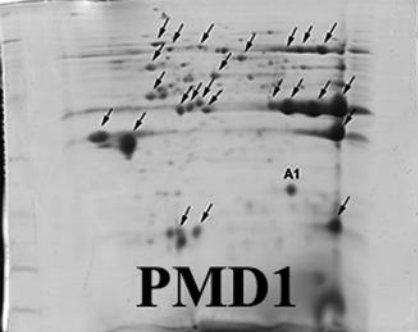
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References

- Amelia-Yap ZH, Chen CD, Sofian-Azirun M, Low VL. Pyrethroid resistance in the dengue vector *Aedes aegypti* in Southeast Asia: present situation and prospects for management. *Parasit Vectors*. 2018;11:332.
- Moyes CL, Vontas J, Martins AJ, Ng LC, Koo SY, Dufour I, et al. Contemporary status of insecticide resistance in the major *Aedes* vectors of arboviruses infecting humans. *PLoS Negl Trop Dis*. 2017;11:e0005625.

3. Chareonviriyaphap T, Bangs MJ, Suwonkerd W, Kongmee M, Corbel V, Ngoen-Klan R. Review of insecticide resistance and behavioral avoidance of vectors of human diseases in Thailand. *Parasit Vectors*. 2013;6:280.
4. Thongwat D, Bunchu N. Susceptibility to temephos, permethrin and deltamethrin of *Aedes aegypti* (Diptera: Culicidae) from Muang district, Phitsanulok Province, Thailand. *Asian Pac J Trop Med*. 2015;8:14–8.
5. Prapanthadara L, Promtet N, Koottathep S, Somboon P, Suwonkerd W, McCarroll L, et al. Mechanisms of DDT and permethrin resistance in *Aedes aegypti* from Chiang Mai, Thailand. *Dengue Bull*. 2002;26:185–9.
6. Prapanthadara L, Reunkum W, Leelapat P, Suwan W, Yanola J, Somboon P. Glutathione S-transferase isoenzymes and the DDTase activity in two DDT-resistant strains of *Aedes aegypti*. *Dengue Bull*. 2005;29:183–91.
7. Somboon P, Prapanthadara LA, Suwonkerd W. Insecticide susceptibility tests of *Anopheles minimus s.l.*, *Aedes aegypti*, *Aedes albopictus*, and *Culex quinquefasciatus* in northern Thailand. *Southeast Asian J Trop Med Public Health*. 2003;34:87–93.
8. Lumjuan N, McCarroll L, Prapanthadara LA, Hemingway J, Ranson H. Elevated activity of an Epsilon class glutathione transferase confers DDT resistance in the dengue vector, *Aedes aegypti*. *Insect Biochem Mol Biol*. 2005;35:861–71.
9. Sathantriphop S, Paeporn P, Supaphathom K. Detection of insecticides resistance status in *Culex quinquefasciatus* and *Aedes aegypti* to four major groups of insecticides. *Trop Biomed*. 2006;23:97–101.
10. Yanola J, Somboon P, Walton C, Nachaiwieng W, Prapanthadara L. A novel F1552/C1552 point mutation in the *Aedes aegypti* voltage-gated sodium channel gene associated with permethrin resistance. *Pestic Biochem Physiol*. 2010;96:127–31.
11. Somwang P, Yanola J, Suwan W, Walton C, Lumjuan N, Prapanthadara LA, et al. Enzymes-based resistant mechanism in pyrethroid resistant and susceptible *Aedes aegypti* strains from northern Thailand. *Parasitol Res*. 2011;109:531–7.
12. Plernsub S, Stenhouse SA, Tippawangkosol P, Lumjuan N, Yanola J, Choochote W, et al. Relative developmental and reproductive fitness associated with F1534C homozygous knockdown resistant gene in *Aedes aegypti* from Thailand. *Trop Biomed*. 2013;30:621–30.
13. Stenhouse SA, Plernsub S, Yanola J, Lumjuan N, Dantrakool A, Choochote W, Somboon P. Detection of the V1016G mutation in the voltage-gated sodium channel gene of *Aedes aegypti* (Diptera: Culicidae) by allele specific PCR assay, and its distribution and effect on deltamethrin resistance in Thailand. *Parasit Vectors*. 2013;6:253.
14. Plernsub S, Saingamsook J, Yanola J, Lumjuan N, Tippawangkosol P, Sukontason K, et al. Additive effect of knockdown resistance mutations, S989P, V1016G and F1534C, in a heterozygous genotype conferring pyrethroid resistance in *Aedes aegypti* in Thailand. *Parasit Vectors*. 2016;9:417.
15. Plernsub S, Saingamsook J, Yanola J, Lumjuan N, Tippawangkosol P, Walton C, et al. Temporal frequency of knockdown resistance mutations, F1534C and V1016G, in *Aedes aegypti* in Chiang Mai city, Thailand and the impact of the mutations on the efficiency of thermal fogging spray with pyrethroids. *Acta Trop*. 2016;162:125–32.
16. Schmid MA, Glasner DR, Shah S, Michlmayr D, Kramer LD, Harris E. Mosquito saliva increases endothelial permeability in the skin, immune cell migration, and dengue pathogenesis during antibody dependent enhancement. *PLoS Pathog*. 2016;12:e1005676.
17. Djegbe I, Cornelie S, Rossignol M, Demettré E, Seveno M, Remoue F, et al. Differential expression of salivary proteins between susceptible and insecticide-resistant mosquitoes of *Culex quinquefasciatus*. *PLoS One*. 2011;6:e17496.
18. Cornelie S, Rossignol M, Seveno M, Demettré E, Mouchet F, Djegbe I, et al. Salivary gland proteome analysis reveals modulation of anopheline unique proteins in insensitive acetylcholinesterase resistant *Anopheles gambiae* mosquitoes. *PLoS One*. 2014;9:e103816.
19. Vijay S, Rawal R, Kadian K, Raghavendra K, Sharma A. Annotated differentially expressed salivary proteins of susceptible and insecticide-resistant mosquitoes of *Anopheles stephensi*. *PLoS One*. 2015;10:e0119666.
20. World Health Organization. Test procedures for insecticide resistance monitoring in malaria vector mosquitoes, 2nd ed. 2016. <http://www.who.int/iris/handle/10665/250677>. Accessed 6 Oct 2018.
21. Valenzuela JG, Pham VM, Garfield MK, Francischetti IM, Ribeiro JM. Toward a description of the salivome of the adult female mosquito *Aedes aegypti*. *Insect Biochem Mol Biol*. 2002;32:1101–22.
22. Ribeiro JM, Arcà B, Lombardo F, Calvo E, Phan VM, Chandra PK, et al. An annotated catalogue of salivary gland transcripts in the adult female mosquito, *Aedes aegypti*. *BMC Genomics*. 2007;8:6.
23. Champagne DE, Smartt CT, Ribeiro JM, James AA. The salivary gland-specific apyrase of the mosquito *Aedes aegypti* is a member of the 5'-nucleotidase family. *Proc Natl Acad Sci USA*. 1995;92:694–8.
24. Stark KR, James AA. Isolation and characterization of the gene encoding a novel factor X a directed anticoagulant from the yellow fever mosquito, *Aedes aegypti*. *J Biol Chem*. 1998;273:20802–9.
25. Calvo E, Mans BJ, Andersen JF, Ribeiro JM. Function and evolution of a mosquito salivary protein family. *J Biol Chem*. 2006;281:1935–42.
26. Calvo E, Tokumasu F, Marinotti O, Villeval JL, Ribeiro JM, Francischetti IM. Aegyptin, a novel mosquito salivary gland protein, specifically binds to collagen and prevents its interaction with platelet glycoprotein VI, integrin $\alpha 2 \beta 1$, and von Willebrand factor. *J Biol Chem*. 2007;282:26928–38.
27. Ribeiro JM, Charlab R, Valenzuela JG. The salivary adenosine deaminase activity of the mosquitoes *Culex quinquefasciatus* and *Aedes aegypti*. *J Exp Biol*. 2001;204:2001–10.
28. Zhao L, Pridgeon JW, Becnel JJ, Clark GG, Linthicum KJ. Identification of genes differentially expressed during heat shock treatment in *Aedes aegypti*. *J Med Entomol*. 2009;46:490–5.
29. Wasinpiyamongkol L, Patramool S, Luplertlop N, Surasombatpattana P, Doucoure S, Mouchet F, et al. Blood-feeding and immunogenic *Aedes aegypti* saliva proteins. *Proteomics*. 2010;10:1906–16.
30. Oktarianti R, Senjarini K, Hayano T, Fatchiyah F, Aulanni'am. Proteomic analysis of immunogenic proteins from salivary glands of *Aedes aegypti*. *J Infect Public Health*. 2015;8:575–82.
31. Brito LP, Linss JG, Lima-Camara TN, Belinato TA, Peixoto AA, Lima JB, et al. Assessing the effects of *Aedes aegypti* kdr mutations on pyrethroid resistance and its fitness cost. *PLoS One*. 2013;8:e60878.
32. Belinato TA, Martins AJ, Valle D. Fitness evaluation of two Brazilian *Aedes aegypti* field populations with distinct levels of resistance to the organophosphate temephos. *Mem Inst Oswaldo Cruz*. 2012;107:916–22.
33. Martins AJ, Ribeiro CD, Bellinato DF, Peixoto AA, Valle D, Lima JB. Effect of insecticide resistance on development, longevity and reproduction of field or laboratory selected *Aedes aegypti* populations. *PLoS One*. 2012;7:e31889.
34. Wasinpiyamongkol L, Patramool S, Thongrungrat S, Maneekan P, Sangmukdan S, Missé D, et al. Protein expression in the salivary glands of dengue-infected *Aedes aegypti* mosquitoes and blood feeding success. *Southeast Asian J Trop Med Public Health*. 2012;43:1346–57.
35. Jay AR, Marjorie AH. Confidence intervals for the Abbott's formula correction of bioassay data for control response. *J Econ Entomol*. 1989;82:331–5.
36. Jariyapan N, Baimai V, Poovorawan Y, Roytrakul S, Saeung A, Thongsahuan S, et al. Analysis of female salivary gland proteins of the *Anopheles barbirostris* complex (Diptera: Culicidae) in Thailand. *Parasitol Res*. 2010;107:509–16.
37. Sor-suwan S, Jariyapan N, Roytrakul S, Paemanee A, Phumee A, Phattanawiboon B, et al. Identification of salivary gland proteins depleted after blood feeding in the malaria vector *Anopheles campestris*-like mosquitoes (Diptera: Culicidae). *PLoS One*. 2014;9:e90809.
38. Phattanawiboon B, Jariyapan N, Mano C, Roytrakul S, Paemanee A, Sor-suwan S, et al. Salivary gland proteome during adult development and after blood feeding of female *Anopheles dissidens* mosquitoes (Diptera: Culicidae). *PLoS One*. 2016;11:e0163810.
39. Perkins DN, Pappin DJ, Creasy DM, Cottrell JS. Probability-based protein identification by searching sequence databases using mass spectrometry data. *Electrophoresis*. 1999;20:3551–67.



Additional file 3: Table S2. Differential protein expression in salivary glands between PMD and PMD-R strains

SN ^a	MW/pI ^b	ANV±SD ^c		X-Fold Average
		PMD	PMD-R	
Up-regulated proteins				
4	69/8.5	0.59±0.01	0.75±0.00	1.17
5	68/8.9	1.24±0.02	1.35±0.04	1.09
24	15/5.9	0.25±0.00	0.43±0.00	1.37
Down-regulated proteins				
1	75/5.3	0.33±0.01	0.22±0.00	1.52
2	73/5.6	0.22±0.01	0.19±0.00	1.15
3	72/6.2	0.34±0.01	0.21±0.01	1.59
6	68/9.5	2.56±0.03	1.98±0.03	1.29
7	66/7.2	0.77±0.03	0.63±0.00	1.23
8	62/5.1	0.49±0.02	0.41±0.00	1.19
9	59/6.6	0.81±0.01	0.13±0.03	6.06 ^d
10	49/5.1	0.86±0.03	0.77±0.05	1.13
11	48/6.0	0.58±0.01	0.12±0.02	4.07 ^d
12	45/5.8	0.56±0.01	0.40±0.00	1.95
13	46/5.7	0.73±0.00	0.47±0.00	1.77
14	43/6.2	0.90±0.01	0.46±0.00	1.96
15	40/9.5	9.70±0.03	6.66±0.00	1.46
16	39/9.5	7.23±0.05	4.72±0.00	1.53
17	40/8.9	7.10±0.01	4.93±0.00	1.44
18	42/8.5	3.35±0.01	7.68±0.00	1.94
19	33/9.5	6.32±0.02	4.55±0.00	1.39
20	31/4.1	5.04±0.02	4.03±0.00	1.13
21	33/4.6	4.17±0.00	3.56±0.09	1.11
22	15/9.5	5.82±0.01	2.65±0.00	2.20 ^d
23	15/5.6	0.43±0.00	0.40±0.03	1.06

^aSpot number refers to those shown in Fig. 2

^bObserved molecular mass and isoelectric point

^cAverage Normalized Volume ± Standard Deviation

^dStudent's t-test, P≤0.05 with a cut-off of 2-fold

Additional file 4: Table S3. Differential protein expression in salivary glands between PMD and UPK-R strains

SN ^a	MW/pI ^b	ANV±SD ^c		X-Fold Average
		PMD	UPK-R	
Up-regulated proteins				
4	69/8.5	0.59±0.01	1.05±0.02	1.18
5	68/8.9	1.24±0.02	1.33±0.01	1.07
7	66/7.2	0.77±0.03	1.15±0.01	1.49
17	40/8.9	7.10±0.01	7.11±0.01	1.00
24	15/5.9	0.25±0.00	0.66±0.03	1.79
Down-regulated proteins				
1	75/5.3	0.33±0.01	0.29±0.05	1.12
2	73/5.6	0.22±0.01	0.12±0.01	1.87
3	72/6.2	0.34±0.01	0.23±0.03	1.46
6	68/9.5	2.56±0.03	2.32±0.01	1.11
8	62/5.1	0.49±0.02	0.40±0.03	1.24
9	59/6.6	0.81±0.01	0.14±0.03	5.92 ^d
10	49/5.1	0.86±0.03	0.75±0.03	1.15
11	48/6.0	0.58±0.01	0.12±0.02	4.02 ^d
12	45/5.8	0.56±0.01	0.44±0.03	1.94
13	46/5.7	0.73±0.00	0.22±0.00	3.80 ^d
14	43/6.2	0.90±0.01	0.85±0.02	1.06
15	40/9.5	9.70±0.03	7.62±0.02	1.27
16	39/9.5	7.23±0.05	5.12±0.03	1.41
18	42/8.5	3.35±0.01	2.62±0.00	1.28
19	33/9.5	6.32±0.02	4.83±0.03	1.31
20	31/4.1	5.04±0.02	4.09±0.04	1.28
21	33/4.6	4.17±0.00	2.54±0.01	2.25 ^d
22	15/9.5	5.82±0.01	2.25±0.02	2.38 ^d
23	15/5.6	0.43±0.00	0.15±0.03	2.90 ^d

^aSpot number refers to those shown in Fig. 2

^bObserved molecular mass and isoelectric point

^cAverage Normalized Volume ± Standard Deviation

^dStudent's t-test, P≤0.05 with a cut-off of 2-fold

Additional file 5: Table S4. Differential protein expression in salivary glands between PMD-R and UPK-R strains

SN ^a	MW/pI ^b	ANV±SD ^c		X-Fold Average
		PMD-R	UPK-R	
Up-regulated proteins				
1	75/5.3	0.22±0.00	0.29±0.05	1.36
3	72/6.2	0.21±0.01	0.23±0.03	1.09
4	69/8.5	0.75±0.00	1.05±0.02	1.00
6	68/9.5	1.98±0.03	2.32±0.01	1.17
7	66/7.2	0.63±0.00	1.15±0.01	1.83
9	59/6.6	0.13±0.03	0.14±0.03	1.02
11	48/6.0	0.12±0.02	0.12±0.02	1.01
12	45/5.8	0.40±0.00	0.44±0.03	1.00
14	43/6.2	0.46±0.00	0.85±0.02	1.85
15	40/9.5	6.66±0.00	7.62±0.02	1.14
16	39/9.5	4.72±0.00	5.12±0.03	1.09
17	40/8.9	4.93±0.00	7.11±0.01	1.44
19	33/9.5	4.55±0.00	4.83±0.03	1.06
20	31/4.1	4.034±0.00	4.09±0.04	0.88
24	15/5.9	0.43±0.00	0.66±0.03	1.31
Down-regulated proteins				
2	73/5.6	0.19±0.00	0.12±0.01	1.63
5	68/8.9	1.35±0.04	1.33±0.01	1.02
8	62/5.1	0.41±0.00	0.40±0.03	1.04
10	49/5.1	0.77±0.05	0.75±0.03	1.02
13	46/5.7	0.47±0.00	0.22±0.00	2.14 ^d
18	42/8.5	7.68±0.00	2.62±0.00	2.48 ^d
21	33/4.6	3.56±0.09	2.54±0.01	2.03 ^d
22	15/9.5	2.65±0.00	2.25±0.02	1.08
23	15/5.6	0.40±0.03	0.15±0.03	2.73 ^d

^aSpot number refers to those shown in Fig. 2

^bObserved molecular mass and isoelectric point

^cAverage Normalized Volume ± Standard Deviation

^dStudent's t-test, P≤0.05 with a cut-off of 2-fold

Additional file 6: Table S5. The confidence scores of interaction between the eight major proteins and other proteins and chemicals

Gene name (description of identified proteins) ^a	Gene name or chemical (description of proteins or chemicals) ^b	Confidence score ^c
AAEL017349-PA (AAEL017349-PA)	AAEL012827 (endoplasmin)	0.996
	AAEL004438 (conserved hypothetical protein)	0.994
	AAEL011704 (heat shock protein)	0.987
	AAEL011708 (heat shock protein)	0.987
	AAEL014843 (heat shock protein)	0.987
	AAEL014845 (heat shock protein)	0.987
	Sodium (sodium ion)	0.876
	AAEL000641 (protein disulfide isomerase)	0.757
	AAEL006307 (acetyl-coA carboxylase)	0.537
	AAEL002102 (conserved hypothetical protein)	0.520
APY (apyrase precursor/apyrase)	AAEL005672 (adenosine deaminase)	0.700
	AAEL006485 (inosine-uridine preferring nucleoside hydrolase)	0.546
AAEL006333 (salivary apyrase, putative/5'nucleotidase/AAEL006333-PA)	AAEL005672 (adenosine deaminase)	0.737
	AAEL006485 (inosine-uridine preferring nucleoside hydrolase)	0.432
AAEL005672 (adenosine deaminase, putative)	AAEL006333 (salivary apyrase, putative)	0.737
	APY (apyrase precursor)	0.700
	AAEL006307 (acetyl-coA carboxylase)	0.416
AAEL000641 (protein disulfide isomerase/AAEL000641-PA)	AAEL002102 (conserved hypothetical protein)	0.990
	AAEL007306 (alpha-actinin)	0.980
	AAEL009701 (conserved hypothetical protein)	0.980
	AAEL012827 (endoplasmin)	0.952
	AAEL011704 (heat shock protein)	0.803
	AAEL011708 (heat shock protein)	0.803
	AAEL014843 (heat shock protein)	0.803
	AAEL014845 (heat shock protein)	0.803
	Sodium (sodium ion)	0.772
	AAEL017349-PA (AAEL017349-PA)	0.757
SRPN23 (salivary anti FXa serpin/salivary serpin)	AAEL006307 (acetyl-coA carboxylase)	0.489
	Sodium (sodium ion)	0.412
AAEL006485 (inosine-uridine preferring nucleoside hydrolase/putative purine hydrolase)	Sodium (sodium ion)	0.577
	APY (apyrase precursor)	0.546
	AAEL006333 (salivary apyrase, putative)	0.432
CTL16 (C-Type lectin (CTL), putative/AAEL000533-PA)	Sodium (sodium ion)	0.535

^aThe eight major proteins identified in this study

^bProteins or chemicals that were predicted by STITCH database 5.0 that interacted with the identified proteins

^cThe range of confidence scores by STITCH database 5.0 is 0-1 and 1 is the highest possible confidence