bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

SARS-CoV-2 proteome microarray for mapping COVID-19 antibody interactions at amino acid resolution

Hongye Wang[1,4], Xin, Hou[2,4], Xian Wu[2,4], Te Liang[1], Xiaomei Zhang[1], Dan Wang[1], Fei Teng[3], Jiayu Dai[1], Hu Duan[1], Shubin Guo[3], Yongzhe Li[2,5] and Xiaobo Yu[1,5]

[1]State Key Laboratory of Proteomics, Beijing Proteome Research Center, National Center for Protein Sciences-Beijing (PHOENIX Center), Beijing Institute of Lifeomics, Beijing, 102206, China.

[2]Department of Clinical Laboratory, Peking Union Medical College Hospital, Chinese Academy of Medical Science & Peking Union Medical College, Beijing 100730, China.

[3] Department of Emergency Medicine, Beijing Chao-Yang Hospital, Capital Medical University, & Beijing Key Laboratory of Cardiopulmonary Cerebral Resuscitation, Beijing 100020, China.

[4]These authors contributed equally to this work.

[5]Correspondence to yuxiaobo@mail.ncpsb.org and yongzhelipumch@126.com.

Abstract

COVID-19 has quickly become a worldwide pandemic, which has significantly impacted the economy, education, and social interactions. Understanding the humoral antibody response to SARS-CoV-2 proteins may help identify

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

biomarkers that can be used to detect and treat COVID-19 infection. However, no immuno-proteomics platform exists that can perform such proteome-wide analysis. To address this need, we created a SARS-CoV-2 proteome microarray to analyze antibody interactions at amino acid resolution by spotting peptides 15 amino acids long with 5-amino acid offsets representing full-length SARS-CoV-2 proteins. Moreover, the array processing time is short (1.5 hours), the dynamic range is ~2 orders of magnitude, and the lowest limit of detection is 94 pg/mL. Here, the SARS-CoV-2 proteome array reveals that antibodies commercially available for SARS-CoV-1 proteins can also target SARS-CoV-2 proteins. These readily available reagents could be used immediately in COVID-19 research. Second, IgM and IgG immunogenic epitopes of SARS-CoV-2 proteins were profiled in the serum of ten COVID-19 patients. Such epitope biomarkers provide insight into the immune response to COVID-19 and are potential targets for COVID-19 diagnosis and vaccine development. Finally, serological antibodies that may neutralize viral entry into host cells via the ACE2 receptor were identified. Further investigation into whether these antibodies can inhibit the propagation of SARS-CoV-2 is warranted. Antibody and epitope profiling in response to COVID-19 is possible with our peptide-based SARS-COV-2 proteome microarray. The data gleaned from the array could provide invaluable information to the scientific community to understand, detect, and treat COVID-19.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

Text

The first case of infection caused by the novel coronavirus (SARS-CoV-2) was reported in Wuhan City, China, in December 2019. SARS-CoV-2 has since proven to be highly infectious, with the median incubation period of 4 days [1-3]. Infection of SARS-CoV-2, called COVID-19, results in a range of symptoms, ranging from a mild cough to pneumonia. It is estimated that 17.9% of patients might be asymptomatic[4], which may lead to two or even three transmissions per infected individual [3,5,6].Particular subsets of the population are extremely vulnerable to COVID-19, including the elderly, those with underlying conditions, and immunocompromised individuals. For example, 80% of the deaths attributed to COVID-19 occur among adults $\geq$ 65 years old. On the evening of January 30, 2020, the World Health Organization listed the novel coronavirus outbreak as a public health emergency of international concern [7].As of March 25, the novel coronavirus had spread worldwide[8], with 417,966 confirmed cases and 18,615 deaths in 169 countries. The high transmission rates of SARS-CoV-2, limited diagnostic tests, and no anti-viral treatment options pose huge challenges for the control and treatment of SARS-CoV-2 infected patients [9,10].

SARS-CoV-2 is 82% similar to the original SARS virus attributed to the outbreak in 2003[11]. Generally, a mature SARS-CoV-2 virus has a polyprotein (the open reading frame 1a and 1b, Orf1ab), four structural proteins (envelope, E; membrane, M; nucleocapsid, N; spike, S) and five accessary proteins

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

(Orf3a, Orf6, Orf7a, Orf8, Orf10)[12]. The Orf1ab is involved in viral RNA replication and transcription[12]. The E and M proteins are important in the viral assembly of a coronavirus, and the N protein is necessary for viral RNA synthesis[13]. The S protein is on the surface of the viral particle, enabling the infection of host cells by binding to the host cell receptor, ACE2, via the S-protein's receptor binding domain (RBD) within the S-protein's subunit 1. The RBD of SARS-CoV-2 is very different from the S protein's RBD of SARS-CoV-1; in fact, they only share 73.6% homology[14]. The SARS-CoV-2 RBD can bind faster to the ACE2 receptor than the SARS-CoV-1 RBD, thus resulting in high transmission efficiency of the virus[14]. The accessory proteins may have functions in signaling inhibition, apoptosis induction and cell cycle arrest[12].

B cells defend the body against viruses, such as the SARS-CoV-2 virus, by producing antibodies, which bind to viral particles to mark them for destruction by other cells in the immune system [15,16]. However, there still don't have an immuno-proteomics platform to perform proteome-wide analysis of humoral antibody response to SARS-CoV-2 proteins yet. Here we describe a SARS-CoV-2 proteome peptide microarray that enables high throughput antibody screening of COVID-19 patients to all SARS-CoV-2 protein sequences at amino acid resolution.

To produce the SARS-CoV-2 proteome microarray (Figure 1a), we first extracted the reference sequences of ten proteins encoded by the

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

SARS-CoV-2 coronavirus genome (Wuhan-Hu-1 strain) from the NCBI database (Accession no. MN908947.3). Using these reference sequences, we prepared a peptide library containing 966 peptides representing SARS-CoV-2 proteins, in which each peptide was 15 amino acids long with a 5 amino acid overlap. All peptides were labeled with a C-terminal biotin group and printed onto a 3D-modified microscope slide using biotin-streptavidin chemistry. Full-length SARS-COV-2 N protein, full-length E, and five S truncated proteins were also printed. (Supplementary Table 1).

We next determined the optimal lengths of time to block the array, incubate with serum samples, and incubate with the detection antibody using serum spiked with anti-SARS antibodies. Optimal signal-to-noise ratios were obtained with blocking for 1 min, serum incubation for 30 min, and detection antibody incubation for 30 min (Supplemental Figures 1 – 3). Serum screening using the SARS-CoV-2 proteome microarray can be performed in 1.5 hours while keeping a good dynamic range (~2 orders) and sensitivity (94 pg/mL) (Figure 1b). This represents a significant decrease in time compared to the standard ~ 18 hours using protein microarrays [17]. The r correlation within an array and between different arrays were 0.9992 and 0.9978, respectively, demonstrating the high reproducibility the SARS-COV-2 proteome microarrays (Figure 1c).

Since the SARS-CoV-1 and SARS-CoV-2 genomes are highly similar, we tested rabbit monoclonal and polyclonal anti-SARS-CoV-1 N protein antibodies

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

on the SARS-COV-2 proteome microarray (Figure 1d, Supplementary Figure 4). The monoclonal Ab displayed high specificity to two epitopes (RRGPE and PAADL) on the SARS-COV-2 N protein with a Z-score higher than 3. The functions of these epitopes are currently unknown. Minor cross-reactivity was observed on the epitope (SVLLF) of the E protein. The polyclonal antibody bound to eleven epitopes (E1-E11) on the N protein with crossreactivity to six epitopes on M, S, ORF8, and ORF1ab proteins. The cross-reactive epitopes on M, S, ORF8, and ORF1ab proteins are different than those present in the N-protein (Figure 1d), and it is unclear why the antibodies have crossreactivity to these proteins. The results were validated using full-length N- and S-proteins (Supplementary Figure 5). These results demonstrate that the antibodies prepared previously to SARS-CoV-1 proteins may also detect SARS-COV-2 proteins [18]. SARS-CoV-1 antibodies could provide a quick alternative to fighting COVID-19 since the generation of antibodies to SARS-COV-2 proteins will take three to six months.

To demonstrate the utility of the SARS-COV-2 proteome microarray in antibody profiling, we screened IgM and IgG antibodies in the serum of ten COVID-19 patients and constructed a landscape of humoral response to the SARS-COV-2 proteome (Figure 2). All IgG and IgA antibody epitopes were identified with a Z-score higher than 3 in at least one of COVID-19 patients (Table 1). Many antibodies targeted peptides from seven SARS-COV-2 proteins (M, N, S, Orf1ab, Orf3a, Orf7a, and Orf8). Notably, four

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

immunodominant epitopes with antibodies in more than 80% COVID-19 patients were present in N (residue 206-210, SPARM), S (residue 816-820, SFIED) and Orf3a (residue 136-140, KNPLL; residue 176-180, SPISE) proteins. However, antibodies to E, Orf6 and Orf10 were not detected.

The identification of B-cell and T-cell epitopes for SARS-COV-2 proteins is critical for the development of effective diagnostic tests and vaccine, especially for structural N and S proteins, which had been predicted by bioinformatics[19,20]. For example, there are several RT-PCR tests for detecting COVID-19 that target the S or N protein genome[21]. ELISA and lateral flow devices are also available that measure IgM and IgG levels to N or S proteins[22]. In this work, using overlapping peptides representing the full-length S protein, human IgM and human IgG antibodies were found to target three and six epitopes, respectively (Figure 2, Table 1). Likewise for the N-protein, IgM antibodies targeted two epitopes and IgG antibodies bound to eight epitopes (Figure 2, Table 1). The structural analysis show that all epitope peptides in RNA binding domain of N protein are located at the loop that are easily accessible to the antibodies (Figure 3a). Six epitopes were identified in S protein structure, in which three epitopes are located at the surface and three epitopes are located inside of the protein (Figure 3b). From the predicted B-cell epitopes[19,20], two epitopes (residue 806-820, LPDPSKPSKRSFIED; residue 456-460, FRKSN)) on S protein, one epitope on N protein (residue 166-170, TLPKG), and one epitope (residue 6-10, GTITV) on M protein were experimentally confirmed in

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

this study. The B-cell epitopes that are identified for SARS-COV-2 proteome using microarray will facilitate the understanding of B-cell immunity, identify biomarkers and vaccine candidates for COVID-19 treatment, which should be investigated in a large clinical cohort in future[20].

The SARS-COV-2 S-protein's RBD (residue 438-498) directly engages the ACE2 receptor and might be an ideal target for developing neutralizing antibodies[23]. However, the identification of neutralizing antibodies to competitively inhibit the binding of SARS-COV-2 virus to the host ACE2 receptor has proved challenging. In this work, we analyzed the immunological response to seven peptide sequences within the RBD. Some IgM antibodies from patient "P52" and IgG antibodies from patients "P10" and "P45" bind to the same epitope (residue 456-460, FRKSN)(Figure 4a and 4b). Structural analysis of the RBD-ACE2 complex shows that the epitope is located in the RBD loop engaged with the ACE2 receptor[24] (Figure 4c), thus supporting our data. This epitope may serve as an antigen to stimulate neutralizing antibodies to the RBD-ACE2 interaction and increase CD4+/CD8+ T-cell responses[19,25].

There are three limitations to the SARS-COV-2 proteome microarrays. First, some antibodies may recognize post-translational modifications or conformational (rather than linear) epitopes. To address this issue, we included full-length N, S and E proteins on our microarrays as a comparison. Second, more than one hundred new SARS-COV-2 strains have been identified

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

(https://www.gisaid.org/) since the preparation of our proteome microarray. These new strains could be included in the next version of the SARS-COV-2 proteome microarrays. Third, our SARS-COV-2 proteome microarray is for research use only at this time; it is not approved to diagnose or manage the treatment of patients at this time.

Altogether, our data demonstrate that a peptide-based SARS-COV-2 proteome microarray can map the humoral antibody response to COVID-19. The data could be used to monitor the immune response and identify immunogenic epitopes to develop effective therapeutic treatment of COVID-19. Scientists who wish to acquire these arrays to help fight this COVID-19 pandemic are encouraged to contact us.

References

1 Huang, C. *et al.* Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*, doi:10.1016/S0140-6736(20)30183-5 (2020).

2 Li, Q. *et al.* Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus-Infected Pneumonia. *N Engl J Med*, doi:10.1056/NEJMoa2001316 (2020).

3 Guan, W. J. *et al.* Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med*, doi:10.1056/NEJMoa2002032 (2020).

4 Mizumoto., K., Kagaya., K., Zarebski;, A. & Chowell., G. Estimating the Asymptomatic Proportion of 2019 Novel Coronavirus onboard the Princess Cruises Ship, 2020 *medRxiv*, doi:10.1101/2020.02.20.20025866 (2020).

5 Mahase, E. China coronavirus: mild but infectious cases may make it hard to control outbreak, report warns. *BMJ* **368**, m325, doi:10.1136/bmj.m325 (2020).

6 Phan, L. T. *et al.* Importation and Human-to-Human Transmission of a Novel Coronavirus in Vietnam. *N Engl J Med*, doi:10.1056/NEJMc2001272 (2020).

7 Wang, C., Horby, P. W., Hayden, F. G. & Gao, G. F. A novel coronavirus outbreak of global health concern. *Lancet*, doi:10.1016/S0140-6736(20)30185-9 (2020).

8 Holshue, M. L. *et al.* First Case of 2019 Novel Coronavirus in the United States. *N Engl J Med*, doi:10.1056/NEJMoa2001191 (2020).

9 Du Toit, A. Outbreak of a novel coronavirus. *Nat Rev Microbiol*,

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

doi:10.1038/s41579-020-0332-0 (2020).

10 Chan, J. F. *et al.* A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. *Lancet*, doi:10.1016/S0140-6736(20)30154-9 (2020).

11 Chan, J. F. *et al.* Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan. *Emerg Microbes Infect* **9**, 221-236, doi:10.1080/22221751.2020.1719902 (2020).

12 Narayanan, K., Huang, C. & Makino, S. SARS coronavirus accessory proteins. *Virus Res* **133**, 113-121, doi:10.1016/j.virusres.2007.10.009 (2008).

13 Schoeman, D. & Fielding, B. C. Coronavirus envelope protein: current knowledge. *Virol J* **16**, 69, doi:10.1186/s12985-019-1182-0 (2019).

14 Tai., W. *et al.* Characterization of the receptor-binding domain (RBD) of 2019 novel coronavirus: implication for development of RBD protein as a viral attachment inhibitor and vaccine. *Cell Mol Immunol*, doi:10.1038/s41423-020-0400-4 (2020).

15 Yu, X. *et al.* Exploration of panviral proteome: high-throughput cloning and functional implications in virus-host interactions. *Theranostics* **4**, 808-822, doi:10.7150/thno.8255 (2014).

16 Zhu, H. *et al.* Severe acute respiratory syndrome diagnostics using a coronavirus protein microarray. *Proc Natl Acad Sci U S A* **103**, 4011-4016, doi:10.1073/pnas.0510921103 (2006).

17 Yu, X. *et al.* Multiplexed Nucleic Acid Programmable Protein Arrays. *Theranostics* **7**, 4057-4070, doi:10.7150/thno.20151 (2017).

18 Colwill, K., Renewable Protein Binder Working, G. & Graslund, S. A roadmap to generate renewable protein binders to the human proteome. *Nat Methods* **8**, 551-558, doi:10.1038/nmeth.1607 (2011).

19 Baruah, V. & Bose, S. Immunoinformatics-aided identification of T cell and B cell epitopes in the surface glycoprotein of 2019-nCoV. *J Med Virol* **92**, 495-500, doi:10.1002/jmv.25698 (2020).

20 Grifoni, A. *et al.* A Sequence Homology and Bioinformatic Approach Can Predict Candidate Targets for Immune Responses to SARS-CoV-2. *Cell Host Microbe*, doi:10.1016/j.chom.2020.03.002 (2020).

21 Diao., B. *et al.* Diagnosis of Acute Respiratory Syndrome Coronavirus 2 Infection by Detection of Nucleocapsid Protein. *medRxiv*, doi:10.1101/2020.03.07.20032524 (2020).

22 OKBA., N. *et al.* SARS-CoV-2 specific antibody responses in COVID-19 patients. *medRxiv*, doi:10.1101/2020.03.18.20038059 (2020).

23 Wang., C., W., L., Drabek., D., Okba., N. & Haperen., R. A human monoclonal antibody blocking SARS-CoV-2 infection. *BioRxiv*, doi:10.1101/2020.03.11.987958 (2020).

24 Yan., R., Zhang., Y., Li., Y., Xia., L. & Zhou., Q. Structure of dimeric full-length human ACE2 in complex with B0AT1. *BioRxiv*, doi:10.1101/2020.02.17.951848 (2020).

25 Jiang, S., He, Y. & Liu, S. SARS vaccine development. *Emerg Infect Dis* **11**, 1016-1020, doi:10.3201/1107.050219 (2005).

Contributions

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

X. H., Y. L. X. W. provided the clinical samples. H.W., X. W., X. H., X. Z., T. L., J. D., T. F., S. G., H. D. and X.Y. executed microarray experiments. D. W., X. Z., T. L. and X.Y. executed the bioinformatics and statistical analysis. X.Y., and Y. L. conceived the idea, designed experiments, analyzed the data and wrote the manuscript.

Acknowledgement

This work was supported by the State Key Laboratory of Proteomics (SKLP-C202001，SKLP-O201703 and SKLP-K201505), the Beijing Municipal Education Commission, National Natural Science Foundation of China (81671618，81871302, 81673040, 31870823), the National Program on Key Basic Research Project (2018YFA0507503, 2017YFC0906703 and 2018ZX09733003) and the CAMS Initiative for Innovative Medicine (2017-I2M-3-001 and 2017-I2M-B&R-01）. We also thank Dr. Brianne Petritis for her critical review and editing of this manuscript.

Competing interests

None declared.

Supplementary information

The supplementary information includes the materials and methods, 1 supplementary table and 5 supplementary figures.

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

Tables

Table 1. The epitopes identified in the serum of COVID-19 patients using SARS-COV-2 proteome microarrays.

Protein name	Epitope*		Total number
	IgG	IgM	
M	16-LLEQW-20	6-GTITV-10	5
	106-TRSMW-110	176-LSYYK-180	
	196-YSRYR200	196-YSRYR-200	
S	26-PAYTN-30	816-SFIED-820	8
	186-FKNLR-190	886-WTFGA-890	
	356-KRISN-360	1046-GYHLM-1050	
	456-FRKSN-460		
	806-LPDPSKPSKRSFIED-820		
	1196-SLIDL-1200		
N	66-FPRGQ-70	206-SPARM-210	8
	96-GGDGK-100	386-QKKQQ-390	
	166-TLPKG-170		
	206-SPARM-210		
	226-RLNQL-230		
	256-KKPRQ-260		
	316-GMSRI-320		
	366-TEPKK DKKKKADETQALPQRQKKQQTVTLPAAD L-400		
Orf1ab	166-SSGVT-170	296-FMGRI-300	32
	306-VASPN-310	336-FVKAT-340	
	386-EYHNESGLKTILRKG-400	1496-TPEEH-1500	
	546-SIFSR-550	1636-HTTDPSFLGRYMSAL-1650	
	1046-VEEAK-1050	2656-KLSHQ-2660	
	1106-SGHNL-1110	4616-QTTPG-4620	
	1346-LKKCK-1350	5976-YRRLI-5980	
	2186-TNSRI-2190	6536-VIWDY-6540	

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

	3206-RYLAL-3210		
	3836-DAFKL-3840		
	4076-DYNTY-4080		
	4226-KYLYF-4230		
	4346-KGKYV-4350		
	4516-MADLV-4520		
	4676-DRYFK-4680		
	4716-TSFGP-4720		
	5136-EFYAY-5140		
	5346-RPFLC-5350		
	5746-FNSVC-5750		
	5836-ISPYN-5840		
	6206-AVHEC-6210		
	6366-QLPFF-6370		
	6716-ELEDF-6720		
	6926-ISDMY-6930		
Orf3a	66-LKKRWQ-70	136-KNPLL-140	4
	136-KNPLL-140	176-TSPIS-180	
	176-TSPIS-180	216-STQLS-220	
	216-STQLS-220		
Orf7a	116-LKRKT-120	26-GTTVL-30	3
		66-ACPDG-70	
		116-LKRKT-120	
Orf8	36-PCPIHFYSKWYIRVGARKSA PLIEL-60	36-PCPIHFYSKWYIRVGARKSAPLIEL-60	1

*Bound by serological antibodies identified with a Z-score higher than 3 in at least one of COVID-19 patients.

Figure and legends

Figure 1. SARS-COV-2 proteome microarray fabrication and application in antibody characterization. (a) The schematic illustration of SARS-COV-2 proteome microarray fabrication and biomedical applications; (b) Dynamic

bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

range of serum antibody detection using SARS-COV-2 proteome microarray. The lowest of detection limit (LOD) was calculated using the signal of the buffer control plus two standard deviations. (c) Reproducibility of serum antibody detection using the SARS-COV-2 proteome microarray. (c) Epitope binding of the anti-SARS-CoV-1 N protein antibody using the SARS-COV-2 proteome microarray. The specific antibody binding to the target epitope is selected with a Z-score higher than 3 as a threshold. The false-colored rainbow color from blue to red corresponds to the Z-score from low to high, respectively.

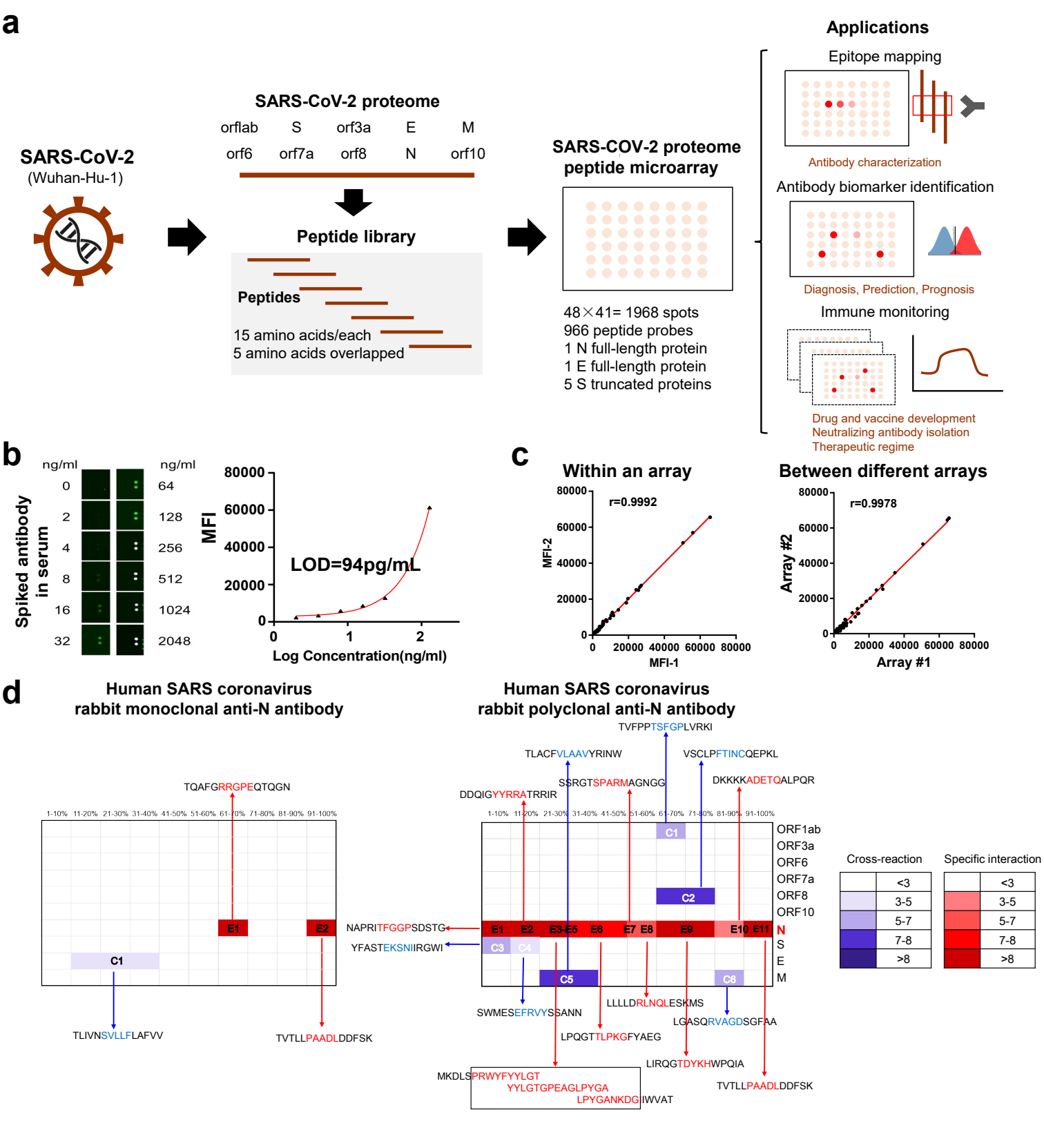
Figure 2. Landscape of humoral antibody response to SARS-COV-2 proteome (a) and (b) are the distribution of human IgM and IgG antibodies to SARS-COV-2 individual proteins, respectively. The x-axis represents the sequence of amino acids of SARS-COV-2 proteins. The y-axis represents the serum samples from COVID-19 patients. The false-colored rainbow color from blue to red corresponds to the signals of antibody binding from low to high, respectively.

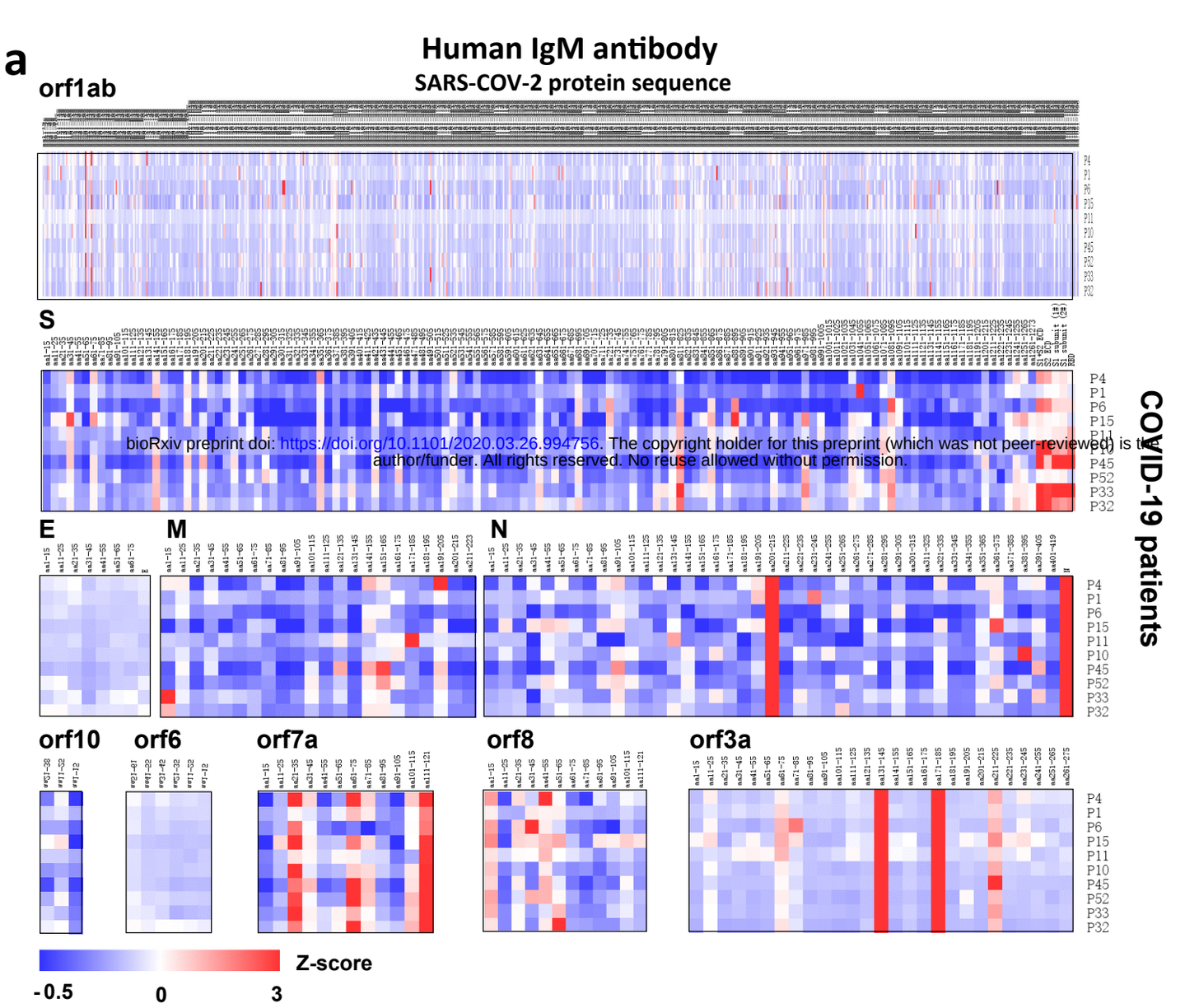
Figure 3. Structural analysis of immunogenic epitopes SARS-COV-2 proteins. (a) and (b) are the structural analysis of nucleocapsid RNA binding domain (PDB ID: 6VYO) and spike trimer protein (PDB ID: 6VXX). The epitope is labeled with yellow and indicated with red arrow.

Figure 4. Identification of potential neutralizing antibody targets in the serum of COVID-19 patients using the SARS-COV-2 proteome microarray.

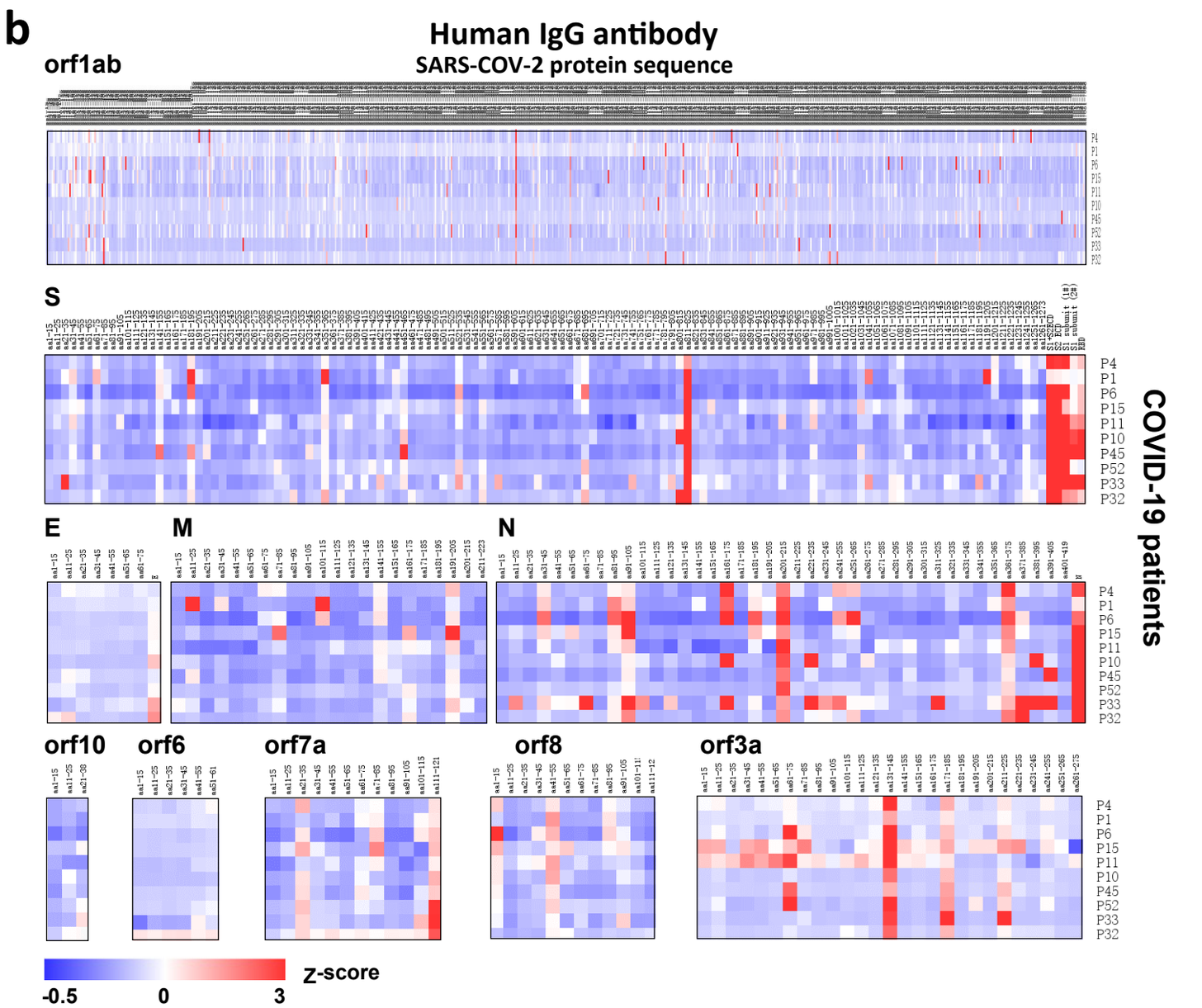
bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

(a) Z-score of serum antibody binding to the peptides with the S-proteins RBD (amino acid residues 431-505). (b) Identification of antibody binding epitope (FRKSN) through sequence alignment. (c) Schematic illustration of the epitope on the RBD (FRKSN) recognized by potential neutralizing antibody in S-protein-ACE2 protein complex (PDB ID: 6M17).


a
SARS-CoV-2
(Wuhan-Hu-1)
SARS-CoV-2 proteome
orflab S orf3a E M
orf6 orf7a orf8 N orf10
Peptide library
Peptides
15 amino acids/each
5 amino acids overlapped
SARS-COV-2 proteome peptide microarray
48×41= 1968 spots
966 peptide probes
1 N full-length protein
1 E full-length protein
5 S truncated proteins
Applications
Epitope mapping
Antibody characterization
Antibody biomarker identification
Diagnosis, Prediction, Prognosis
Immune monitoring
Drug and vaccine development
Neutralizing antibody isolation
Therapeutic regime
b
ng/ml
Spiked antibody in serum
0
2
4
8
16
32
64
128
256
512
1024
2048
MFI
LOD=94pg/mL
Log Concentration(ng/ml)
c
Within an array
r=0.9992
MFI-2
MFI-1
Between different arrays
r=0.9978
Array #2
Array #1
d
Human SARS coronavirus rabbit monoclonal anti-N antibody
TQAFGRRGPEQTQGN
1-10% 11-20% 21-30% 31-40% 41-50% 51-60% 61-70% 71-80% 81-90% 91-100%
E1
E2
C1
TLIVNSVLLFLAFVV
TVTLLPAADLDDFSK
Human SARS coronavirus rabbit polyclonal anti-N antibody
TVFPPTSFGPLVRKI
TLACFVLAAVYRINW
VSCLPFTINCQEPKL
SSRGTSPARMAGNGG
DKKKKADETQALPQR
DDQIGYYRRATRRIR
ORF1ab
ORF3a
ORF6
ORF7a
ORF8
ORF10
N
S
E
M
NAPRITFGGPSDSTG
YFASTEKSNIIRGWI
E3-E5
E6
E7
E8
E9
E10
E11
C2
C3
C4
C5
C6
SWMESEFRVYSSANN
LLLLDRLNQLESKMS
LGASQRVAGDSGFAA
LPQGTTLPKGFYAEG
LIRQGTDYKHWPQIA
MKDLSPRWYFYYLGT
YYLGTGPEAGLPYGA
LPYGANKDGIWVAT
TVTLLPAADLDDFSK
Cross-reaction
Specific interaction
<3
3-5
5-7
7-8
>8



bioRxiv preprint doi: https://doi.org/10.1101/2020.03.26.994756. The copyright holder for this preprint (which was not peer-reviewed) is the author/funder. All rights reserved. No reuse allowed without permission.

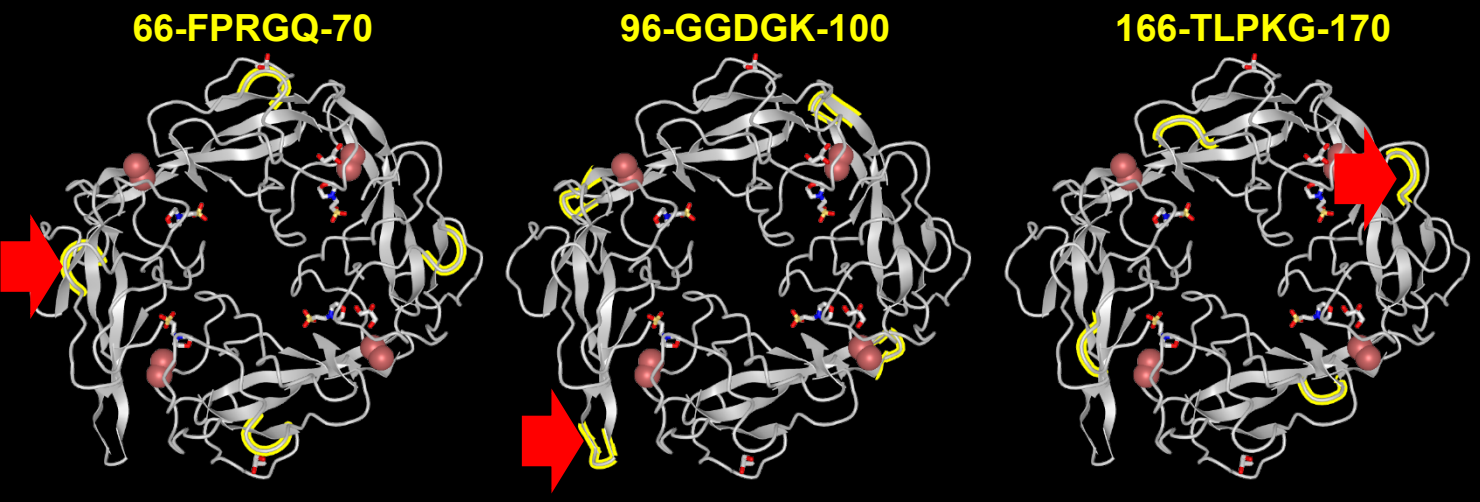


https://doi.org/10.1101/2020.03.26.994756

a

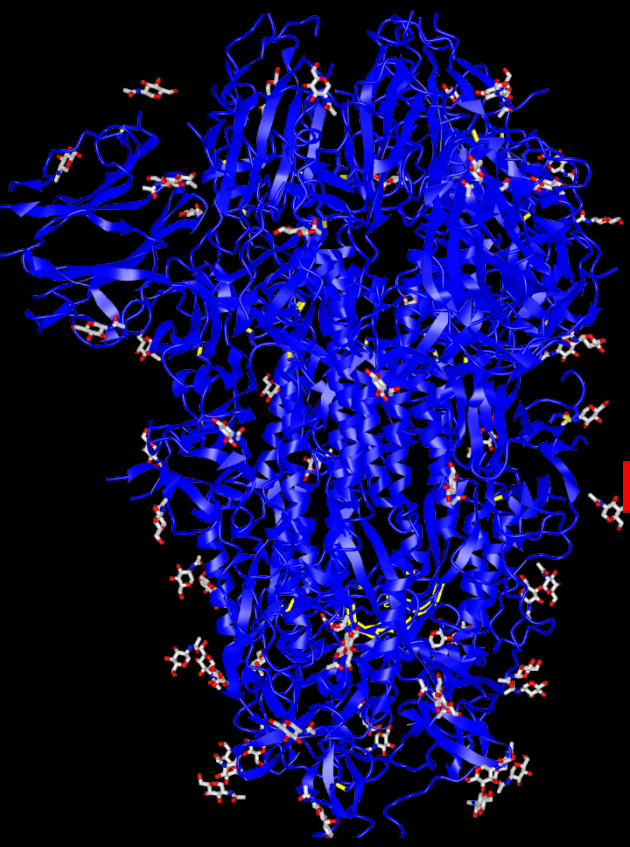
SARS-COV-2

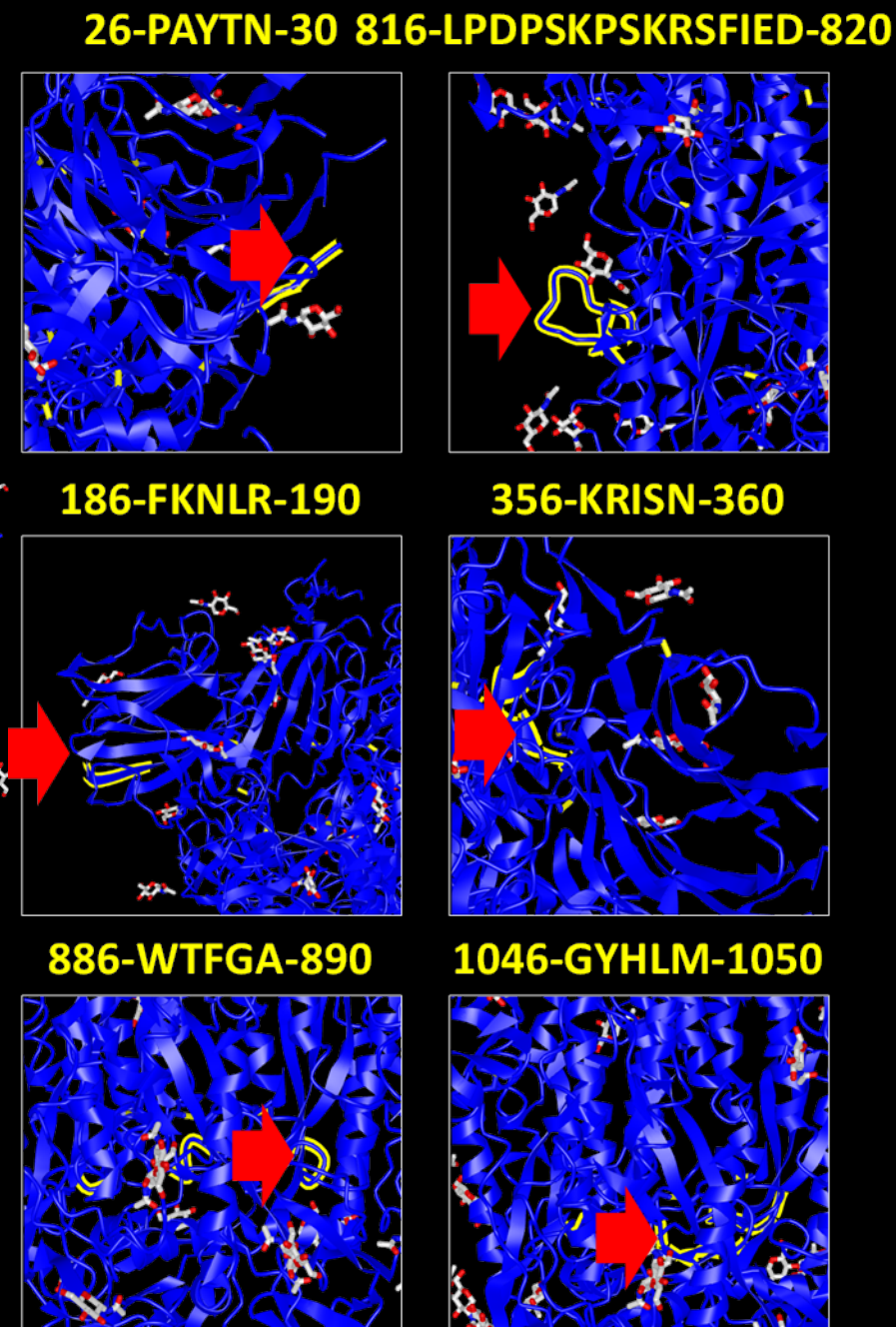
RNA binding domain of nucleocapsid phosphoprotein

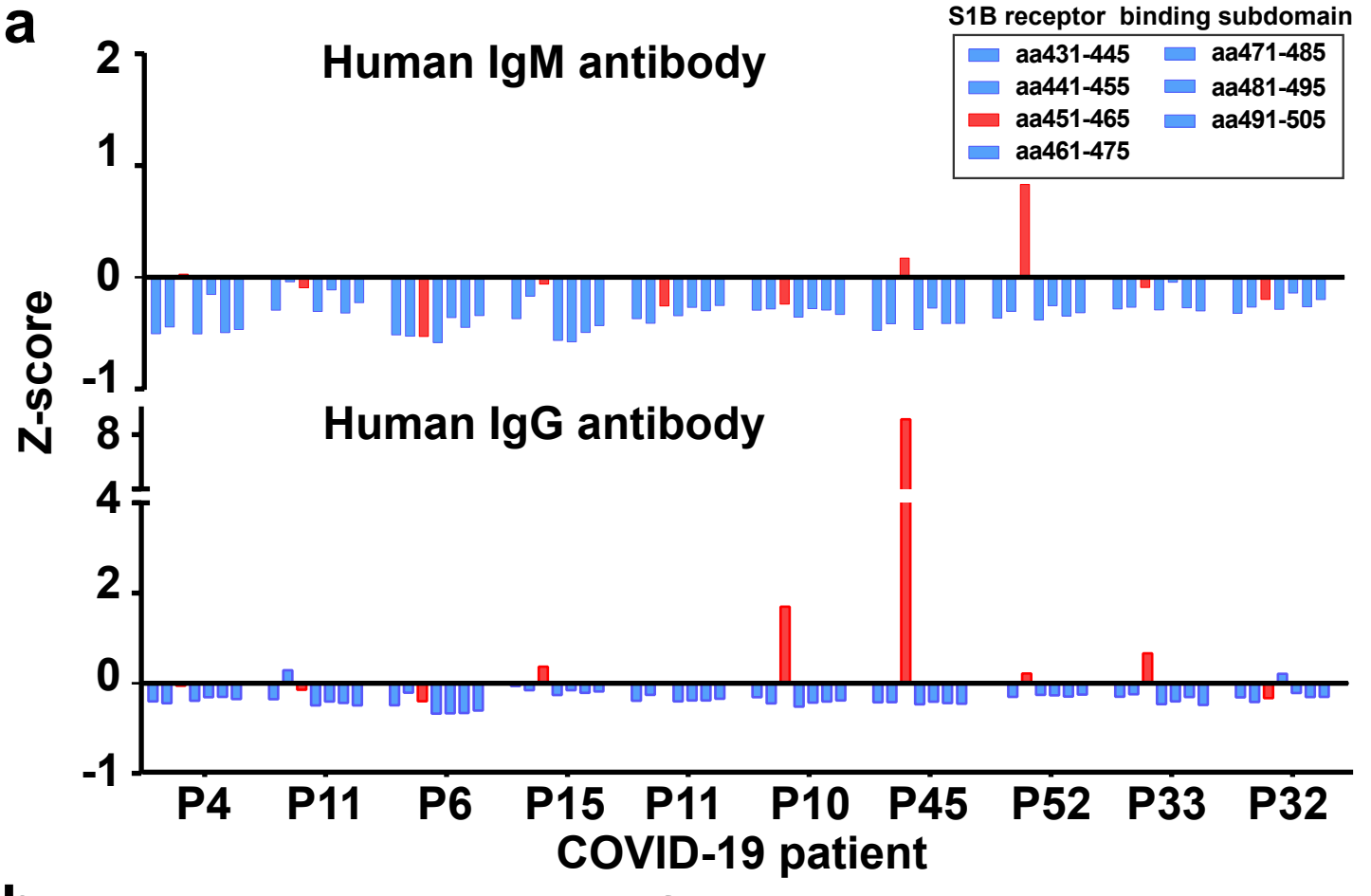


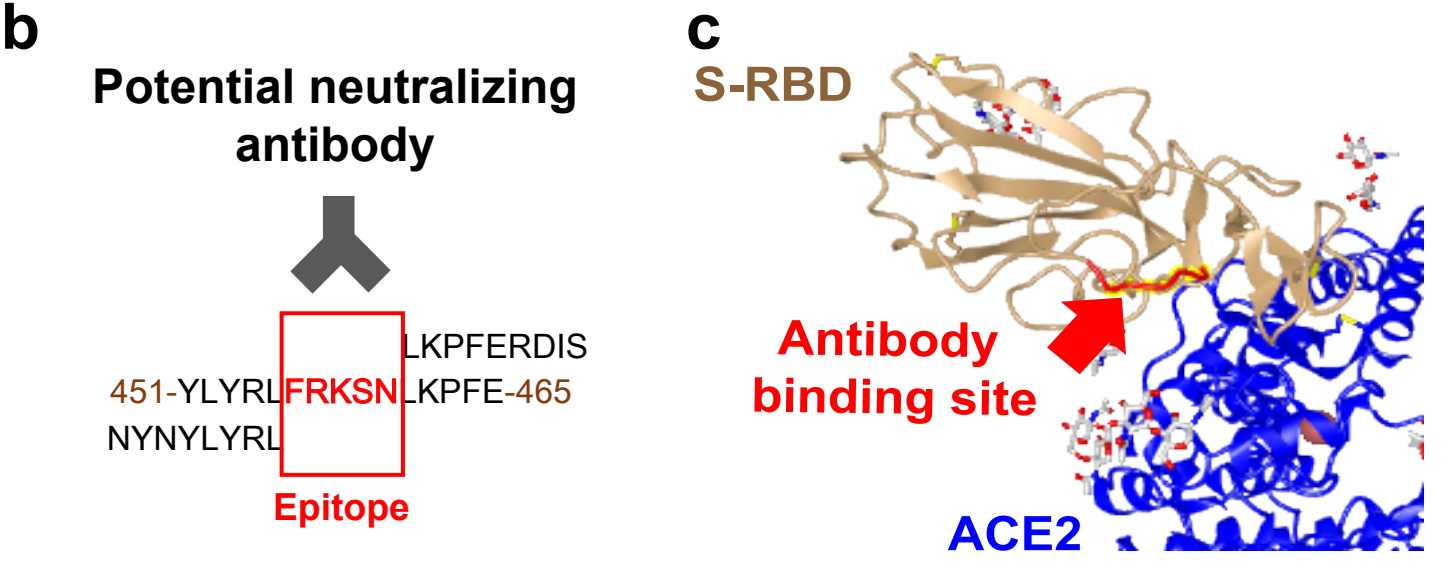
b

SARS-CoV-2
spike glycoprotein
(closed state)






a
S1B receptor binding subdomain
aa431-445
aa441-455
aa451-465
aa461-475
aa471-485
aa481-495
aa491-505
Human IgM antibody
Human IgG antibody
Z-score
2
1
0
-1
8
4
2
0
-1
P4
P11
P6
P15
P11
P10
P45
P52
P33
P32
COVID-19 patient


b
Potential neutralizing antibody
LKPFERDIS
451-YLYRLFRKSNLKPFE-465
NYNYLYRL
Epitope
c
S-RBD
Antibody binding site
ACE2

-Supplementary information-

SARS-CoV-2 proteome microarray for mapping COVID-19 antibody interactions at amino acid resolution

Supplementary methods

1 Supplementary Table

5 Supplementary Figures

Supplementary methods

Preparation of SARS-CoV-2 proteome microarray

All biotin labeled peptides were obtained from China Peptides (Shanghai, China) and Guoping Pharmaceutical Company (Anhui, China). The SARS-COV-2 N and S proteins were obtained from Sino Biological, Inc. (Beijing, China). These peptides were printed onto a 3D modified slide surface (Capital Biochip Corp, Beijing, China) in parallel and in duplicate using an Arrayjet microarrayer (Roslin, UK). Phosphate buffered saline (PBS) and bovine serum albumin (BSA, 100 μg/mL) (Sigma-Aldrich, MO, USA) were used as negative controls. Biotinylated BSA (100 μg/mL), human IgG and IgM (10 μg/mL) were used as positive controls. The peptide microarrays were stored at -20 ℃ until ready to use.

Characterization of anti-SARS antibody using SARS-CoV-2 proteome microarray

The peptide microarrays were assembled in an incubation tray and blocked with 5% (w/v) milk in 1xPBS with 0.2% (v/v) Tween-20 (PBST) for 1 min at room temperature. After washing with PBST three times, the array was incubated with rabbit anti-SARS-CoV-1 N-protein antibody (1 μg/mL) (Sino Biological, Beijing, China) for 30 min at room temperature. After washing again, the array was incubated with an Alexa Fluor 555 labeled goat anti-Rabbit IgG (H+L), cross-adsorbed, secondary antibody (Jackson ImmunoResearch, USA)

for 30 min. The arrays were washed, dissembled from the tray, and dried with centrifugation for 2 min at 2,000 rpm. The resulting array was scanned with a GenePix 4300A microarray scanner (Molecular Devices, Sunnyvale, CA, USA). The median fluorescent signal intensity of each spot was extracted using GenePix Pro7 software (Molecular Devices, Sunnyvale, CA, USA). The median background signal was subtracted from the median spot signal intensity.

Detection of serum antibody using SARS-CoV-2 proteome microarray

All COVID-19 patients were diagnosed according to the Diagnosis and management plan of pneumonia with new coronavirus infection (trial version 7). The serum samples were collected with written informed consent under the approval of the intuitional review board (IRB) from Peking Union Medical College Hospital and Beijing Proteome Research Center. All experiments were performed according to the standards of the Declaration of Helsinki.

Prior to antibody detection, the peptide microarrays were assembled in an incubation tray and blocked with 5% (w/v) milk in 1xPBS with 0.2% (v/v) Tween-20 (PBST) for 1 min at room temperature. After washing with PBST three times, the array was incubated with 1:300 diluted serum for 30 min at room temperature. After washing again, the array was then incubated for 30 min with a mixture containing Cy3 Affinipure donkey anti-human IgG(H+L) and Alexa fluor 647 Affinipure goat anti-human IgM FC5u antibody (Jackson

ImmunoResearch, USA) (2μg/mL). Finally, the array was washed with PBST and water, dissembled from the tray and dried with centrifugation for 2 min at 2,000 rpm. The array was scanned with a GenePix 4300A microarray scanner (Molecular Devices, Sunnyvale, CA, USA), and the signal was read using the laser at 532 and 635 nm, respectively. The median of fluorescent signal intensity with the deduction of background was extracted using GenePix Pro7 software (Molecular Devices, Sunnyvale, CA, USA).

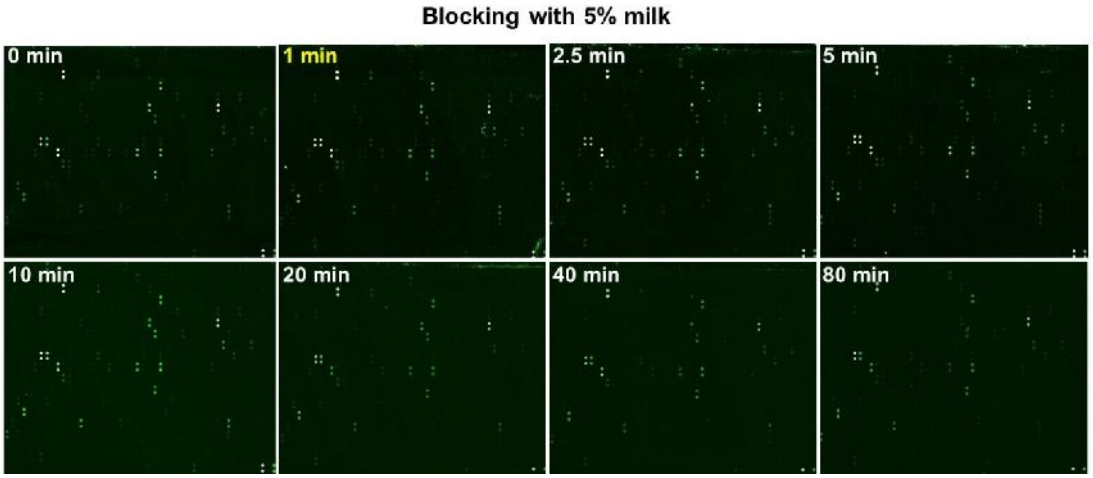
Data analysis

All microarray signals were normalized with a Z-score[1]. The hierarchical cluster analysis of serological antibody response to peptides was performed using the MultiExperiment Viewer 4.1 software (www.tigr.org)[2].

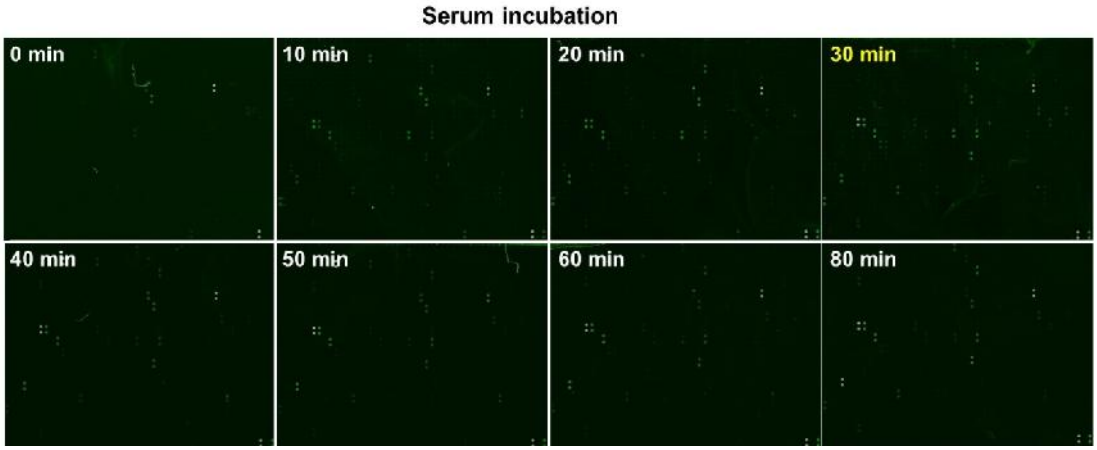
Supplementary Tables

Supplementary Table1. The layout of SARS-COV-2 proteome microarray.

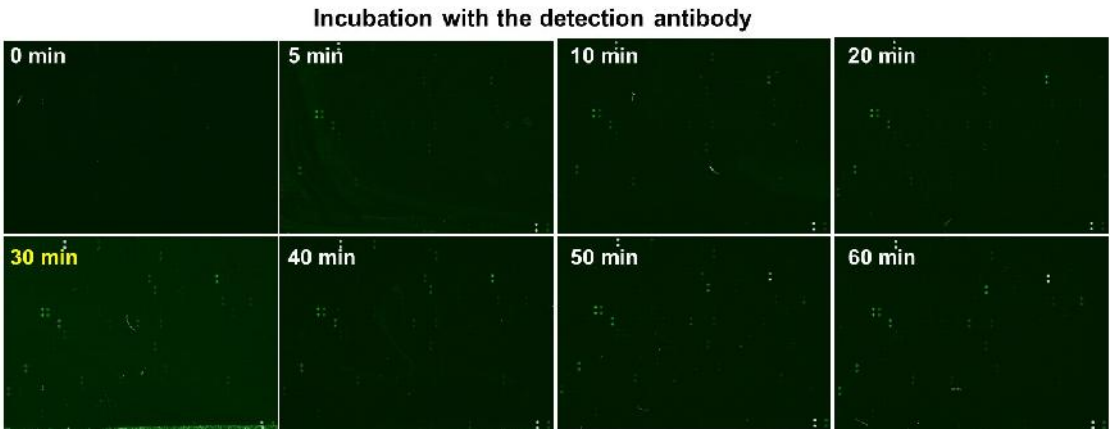
Supplementary Figures



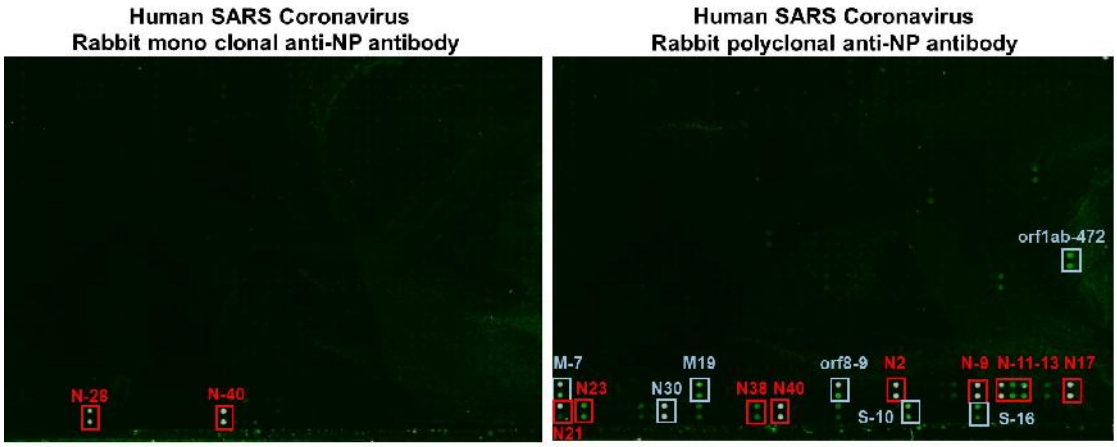
Supplementary Figure 1. A comparison of different blocking incubation lengths on the SARS-COV-2 proteome microarray



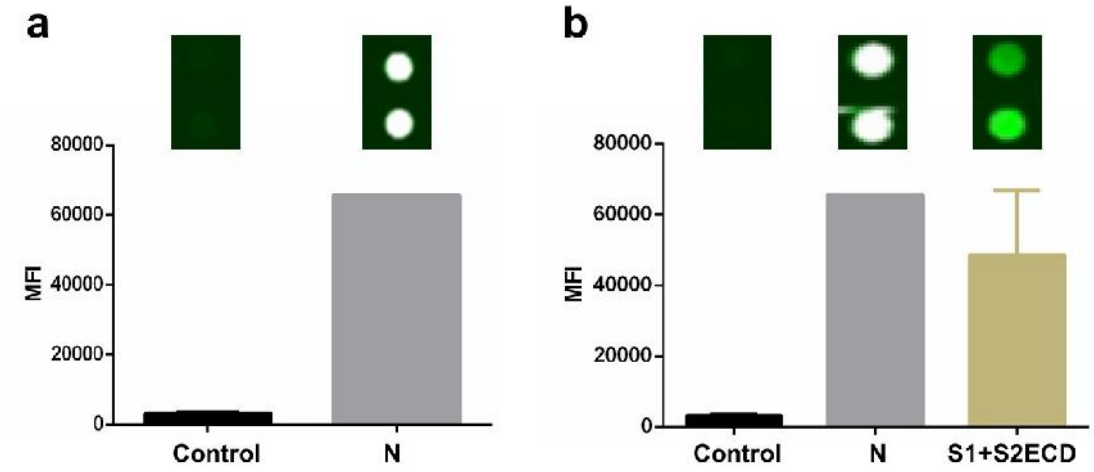
Supplementary Figure 2. A comparison of different serum sample incubation lengths on the SARS-COV-2 proteome microarray



Supplementary Figure 3. A comparison of different detection antibody incubation lengths on the SARS-COV-2 proteome microarray



Supplementary Figure 4. Fluorescent detection image of rabbit anti-SARS-CoV-1 N-protein antibodies using the SARS-CoV-2 proteome microarray. Red = antibody-peptide interactions with peptides from N proteins. Blue = cross-reactive antibody-peptide interactions with peptides from non-N proteins.



Supplementary Figure 5. Binding of anti-SARS-CoV-1 antibodies to full-length SARS-CoV-2 proteins. Interactions of the rabbit (a) monoclonal and (b) polyclonal antibodies with the full-length target proteins. A streptavidin protein was used as the control. S1+S2ECD = S-protein consisting of subunit 1 (S1) and subunit 2 (S2) extracellular domain (ECD).

References

1. Cheadle, C., Vawter, M.P., Freed, W.J. & Becker, K.G. Analysis of microarray data using Z score transformation. *J Mol Diagn* **5**, 73-81 (2003).
2. Chu, V.T., Gottardo, R., Raftery, A.E., Bumgarner, R.E. & Yeung, K.Y. MeV+R: using MeV as a graphical user interface for Bioconductor applications in microarray analysis. *Genome Biol* **9**, R118 (2008).